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Commentary

Deregulation of Slug/Snail2 and TGF-β crosstalk in airway epithelial stem/progenitor cells: A key link between COPD and lung cancer?

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Abstract

This commentary focuses on two recent publications showing deregulation of the transcription factor Slug/Snail2 and TGF- β function in primary bronchial basal/progenitor cells of patients with Chronic Obstructive Pulmonary Disease (COPD) and the impact on proliferation and the expression of genes involved in stem cell maintenance. We discuss the molecular mechanisms related to the exhaustion of airway basal stem/progenitor cells in tobacco smoke-induced COPD, as well as putative links between COPD and lung cancer at the molecular level. The focus of the commentary is a potential central role of the crosstalk between Slug/Snail2 and TGF- β mechanisms, and perspectives for identification of new biomarkers and/or therapeutic targets or therapy for preventing and treating these diseases.

Keywords: Chronic obstructive pulmonary disease (COPD), Lung cancer, Human primary bronchial basal stem/progenitor cells, Transforming growth factor (TGF)-β, Slug/Snail2 transcription factor

Introduction to COPD and Airway Remodeling

Chronic Obstructive Pulmonary Disease (COPD) is a respiratory disease characterized by progressive and irreversible loss of respiratory capacity. Different stages of the disease have been defined based on the decrease in respiratory capacity of the patient that can range from mild to severe. COPD results essentially from inhalation of toxic gas and particles, and a major cause is tobacco smoke. Continual exposure to the compounds in tobacco smoke induces tissue damage, leading to irreversible morphological and functional alterations in the lungs.

COPD is a complex disease and is characterized by a combination of structural and inflammatory changes. In addition to a high level of inflammation, a remodeling of the airways in the COPD patients is leading to an increase in the thickness of the airway wall and a narrowing of the airway lumen. A central issue in airway remodeling is that chronic exposure to tobacco smoke directly affects the morphology and function of the epithelium lining the airways. [1]. Whereas COPD has been initially seen as an inflammation-driven disease, and research and therapy were focused mainly on the treatment of airway inflammation, only recently airway remodeling is increasingly considered as the primary feature underlying COPD development [2,3].

This change of paradigm, from COPD being essentially an inflammatory disease to COPD being primarily an airway remodeling disease, changed the focus of the research toward better understanding of the airway remodeling and its mechanisms with a particular emphasis on the impact of tobacco smoke on the occurrence of the epithelial abnormalities and their link with COPD initiation and development [3]. The work that we have recently published [4,5] is part of this area of research and stems from several recent advances in understanding the causes of airway epithelium deregulations in COPD smokers.

Background/State of the Art on COPD and Airway Epithelial Abnormalities

Basal stem/progenitor cell reprogramming

A mix of basal, ciliated, and mucus-producing goblet cells essentially composes the pseudostratified

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airway epithelium. Basal cells contain the adult stem/progenitor cells of the airway epithelium that can either self-renew and/or differentiate in ciliated or goblet cells to repair the epithelium after injury [6].

The abnormalities in the COPD airway epithelium consist mainly of areas of squamous metaplasia and goblet cell hyperplasia at the epithelial luminal side and, at the basal membrane, of wall fibrosis and thickening [7,8]. Recently, it has been reported that basal stem/progenitor cell number is reduced in COPD and that they regenerate an epithelium with a reduced number of ciliated cells and relatively more basal and goblet cells. Several studies provide evidence that tobacco smoke has a major impact on basal stem/progenitor cells causing aberrant repair processes and abnormalities in airway epithelium of COPD. This would be the result of a deregulation of their genetic program leading to an imbalance in their fate and a pathological differentiation process. [6,9-11]

Aberrant expression of epigenetic markers and methylation modifications have been linked to COPD, suggesting that epigenetic modifications are involved in this reprogramming [12-14].

Field of injury implicating large and small airways

The loss of respiratory capacity characterizing COPD has been linked to small airways. A correlation between disease severity and the degree of small airway remodeling, in particular epithelial anomalies, has been shown [15,16]. However, several studies have also revealed that basal stem/progenitor cells exposed to tobacco smoke keep the memory of the exposure, likely through epigenetic modifications, and that a "field of injury" is established all along the airway epithelium with both small and large airway epithelia presenting lesions. In addition, previous studies imply that basal progenitors from large airways reflect the dysfunction of progenitors of the small airways and that molecular changes in large airways are also seen in small airways. This makes cells from large airways a good model to study the molecular mechanisms leading to epithelial abnormalities and their link with COPD pathogenesis [10,11]

TGF- β signaling and epithelial-mesenchymal transition at the crossroad of COPD and lung cancer

Studies have shown that mature epithelium from COPD airways present features of Epithelial-Mesenchymal Transition (EMT), including myofibroblast formation. Research that followed has investigated the possibility that EMT is active and pathogenic in the airways of smokers with COPD. The data obtained suggest that EMT plays a major role in COPD as an airway remodeling disease and this led to the hypothesis that COPD may be an EMT driven process [17,18].

COPD smokers have an increased risk of lung cancer compared to non-COPD smokers, in particular of squamous carcinoma in the proximal bronchus [19,20]. Deregulation of the COPD airway epithelium is likely involved in this association and it has also increasingly been linked with EMT. In a recent study, EMT activity in non-small cell lung cancers has been directly related to EMT in non-cancerous airways of the same patients [21].

Transforming Growth Factor (TGF)- β is known to be, in certain conditions, an EMT-inducing factor [22,23]. It is found at high levels in COPD lung tissues and clinicopathologic studies have shown that TGF- β is likely to have a major EMT-inducing role

in COPD [24,25]. TGF- β is particularly interesting as its function is complex and it has a wide spectrum of effects, which can be antagonistic depending on the cell state. It has been shown to play a role in the fate of stem/progenitor cells. Whereas it is a cell cycle inhibitor in normal cells, it can act as a tumor promoter in malignant cells [26-28].

Aims and Main Findings

Experimental approach: Design and hypothesis

Our aim was to further understand the reprogramming of basal stem/progenitor cells in COPD, and to search for molecular links between COPD and lung cancer. For this, we first wanted to identify and compare the genes that are involved in the genetic differentiation program of normal and COPD basal stem/progenitor cells. To identify key genes involved in the regulation of basal stem/progenitor cell fate and differentiation process, we choose to identify the genes involved at the onset of differentiation rather than genes involved in the mature cell functions.

As an experimental model we used primary bronchial epithelial progenitors cells cultured in the Air-liquid Interface (ALI) system. Bronchial progenitor cells, which originate from large airways, are easier to obtain and their molecular dysfunctions reflect to some extend that of progenitors of the small airways [10,11]. Primary bronchial epithelial progenitors cultured at the ALI are able to reconstitute the pseudostratified bronchial epithelium. Importantly, they retain the memory of the tissue of origin and when isolated from COPD patients, they reconstitute an epithelium with abnormalities characteristic of COPD [18,29,30].

Since we anticipated that many genes would be involved in the onset of differentiation, we choose to particularly focus on genes downstream of the transcription factor Slug/Snail2. Like other members of the Snail family of EMT-inducing transcription factors, Slug/Snail2 is overexpressed and involved in EMT-related processes in several cancers, including lung. [31]. In contrast to other EMT-inducing transcription factors, Slug/Snail2 is expressed in adult stem/progenitor cells in normal airway epitheliums. It was found enriched in both mouse and human airway basal cells and expressed in spheroids of stem/progenitor cells isolated from human adult airway epithelia [32-34]. Moreover, TGF- β , that is at high levels in COPD lung tissues and involved in EMT, is also known to regulate the expression of Slug/Snail2 [24,25,35]. We then hypothesized that Slug/Snail2, in combination with TGF-B, could be a key transcription regulator involved in adult airway basal stem/ progenitor cell fate, and that the deregulation of this crosstalk could lead to the airway epithelial dysfunction observed in COPD.

Data generated and main results

We have generated two datasets from our experiments. One corresponds to expression data from primary human bronchial epithelial basal cells from non-COPD and COPD smokers that had Slug/Snail2 knocked down and the other to expression data from primary human bronchial epithelial basal cells from smokers non-COPD and COPD in ALI culture at the onset of differentiation in absence or presence of TGF- β .

By combining these two datasets, we have selected the genes that are repressed downstream of Slug/Snail2 and that respond to both differentiation and TGF- β . These genes, either from normal or from

COPD cells, were classified in 4 groups according to their response to differentiation and TGF- β (Figure 3a in [4]; Figure 1a in [5]). This allowed us to visualize important differences between normal and COPD cells in the genes repressed downstream of Slug/Snail2.

In normal cells, we identified a large group of proliferation genes that are downregulated with differentiation and repressed by TGF- β , and that are not downstream of Slug/Snail2 in COPD. In COPD cells, we identified a small set of genes coding for transcription factors involved in stem cell maintenance that are downregulated with differentiation and repressed by TGF- β , and that are not downstream of Slug/Snail2 in normal cells. The study of the levels of expression of both proliferation and stem cell maintenance genes revealed that they differ between normal and COPD basal stem/progenitor cells only in presence of TGF- β . At the onset of differentiation, proliferation genes are less expressed in normal cells due in part to a higher repressive effect of TGF- β in these cells than in COPD cells. Instead, stem cell maintenance genes are less expressed in COPD cells as they are repressed by TGF- β in these cells and not in normal cells (Figure 1). In addition, we found a positive correlation between Slug/Snail2 expression levels and the repressive effect of TGF- β on the proliferation genes in normal cells only, as well as on the stem cell maintenance genes in COPD cells only.

Perspectives for New Directions of Research

Our results reveal that the genes downstream of Slug/ Snail2, which represent a mix of direct and indirect targets of this transcription factor, are deregulated in COPD basal stem/progenitor cells. In these cells, we found that, at the onset of differentiation and in presence of TGF- β , the genes involved in proliferation, independently of Slug/Snail2 pathway, are expressed at higher levels, while stem cell maintenance genes, downstream of Slug/Snail2 pathway, have their expression inhibited. We have also found that the response of COPD basal stem/progenitor cells to TGF- β differs with in particular an increase of expression of β -Catenin, which is part of a non-canonical TGF- β signaling. In addition, we showed an increase of Slug protein levels in COPD cells in presence of TGF- β , but that it was not associated with a decrease of the EMT marker E-cadherin. No difference between normal and COPD cells for the expression of EMT markers was found in undifferentiated cells and at the onset of differentiation in presence or absence of TGF- β .

At this stage of the study, our findings are mainly correlative and hypothesis generating. They will provide a base for new directions of COPD research.

Molecular mechanisms involved in exhaustion of basal stem/ progenitor cells in COPD

The identification of genes involved in stem cell maintenance genes that are repressed by TGF- β in COPD basal stem/progenitor cells brings a new insight in the molecular mechanisms of the disease. They could be directly involved in the decrease of self-renewal and differentiation capacity of basal stem/progenitor cells and/or epithelium abnormalities observed in the airway epithelium of COPD patients [11,12]. Since these genes all code for transcription factors, they are likely among the key regulators of basal stem/progenitor cells. Their specific function and implication in the exhaustion of COPD basal stem/progenitor cells need to be further studied.





Our data also suggest that a crosstalk between Slug/Snail2 and TGF- β is important to further understanding the etiopathology of COPD, Slug/Snail2 may act as a mediator of TGF- β in the regulation of basal stem/progenitor cells, as it has been shown in other cells [36,37]. However, further experiments are required to confirm this.

Slug/Snail2 and TGF- $\pmb{\beta}$ signaling as molecular links between COPD and lung cancer

The implication of Slug/Snail2 and TGF- β in the regulatory changes suggests that this is a good starting point to identify molecular mechanisms involved in the association between COPD and lung cancer. The finding that Slug/Snail2 in combination with TGF- β is involved in the repression of proliferation genes at the onset of differentiation in normal basal stem/progenitor cells and that this repression is deregulated in COPD cells seems particularly interesting for the search of the molecular links between COPD and lung cancer.

TGF- β function is complex with a wide spectrum of effects. It can have antagonistic effects depending on cell state: it can act as a cell cycle inhibitor in normal cells and a tumor promoter in malignant cells [26,27]. We speculate that the difference between normal and COPD cells that we found could reflect such antagonistic effects, with COPD cells being in a premalignant state and imprinted with epigenetic modifications that make them permissive for oncogenic mutations to initiate lung cancer. The change in Slug/ Snail2 downstream genes could reflect difference in TGF- β signaling between normal and premalignant cells. Further studies in that direction could lead to understand at least part of the association of COPD and lung cancer.

It is also important to note that in COPD cells we did not observe a decrease of the EMT marker E-cadherin, even when treated with TGF-B. Other EMT-inducing transcription factors had a similarly low expression in both normal and COPD cells, suggesting that Slug/Snail2 is not an EMT-inducer in undifferentiated basal stem/ progenitor cells or at the onset of their differentiation. However, we speculate that the deregulations observed in the basal stem/ progenitor cells in COPD could ultimately lead to a shift in Slug/ Snail2 function, which then starts acting as an EMT-inducing factor. It has been reported that Slug levels of expression define its function and overexpression of Slug induces EMT in epithelial cells [38,39]. Slug protein is at higher levels in COPD cells in the presence of TGF- β . The high levels of TGF- β found in the lungs of COPD patients increase with the progression of the disease and could increase during repeated injury. This could explain the higher EMT features found in mature COPD epithelium and the increased risk for COPD patients to develop lung cancer.

More studies are required to better understand the central role of TGF- β and its link with Slug/Snail2 transcription factor in the deregulation of airway basal stem/progenitor cells. However, all our findings point to a key role of these deregulations in the transition from normal to COPD cells induced by tobacco smoke. In addition, both TGF- β and Slug/Snail2 have been shown to be involved in lung cancer, making them good candidates to better understand both the molecular mechanism involved in the exhaustion of basal stem/ progenitor cells in COPD and the molecular link between COPD and Lung cancer. Such studies could open up new directions to identify new markers or therapeutic targets or therapy for preventing

and treating these diseases.

Conflict of Interest

The author declares no conflict of interest.

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References

- Vogelmeier C, Criner G, Martinez F, Anzueto A, Barnes P, Bourbeau J, et al. Global Strategy for the Diagnosis, Management, and Prevention of Chronic Obstructive Lung Disease 2017 Report. GOLD Executive Summary. American Journal of Respiratory and Critical Care Medicine. 2017;195(5):557-582.
- Martinez F, Han M, Allinson J, Barr R, Boucher R, Calverley P, et al. At the Root: Defining and Halting Progression of Early Chronic Obstructive Pulmonary Disease. American Journal of Respiratory and Critical Care Medicine. 2018;197(12):1540-1551.
- Walters E, Shukla S, Mahmood M, Ward C. Fully integrating pathophysiological insights in COPD: an updated working disease model to broaden therapeutic vision. European Respiratory Review. 2021;30(160):200364.
- Ben Brahim C, Courageux C, Jolly A, Ouine B, Cartier A, de la Grange P, et al. Proliferation Genes Repressed by TGF-β Are Downstream of Slug/Snail2 in Normal Bronchial Epithelial Progenitors and Are Deregulated in COPD. Stem Cell Reviews and Reports. 2021;17(3):703-718.
- de la Grange P, Jolly A, Courageux C, Ben Brahim C, Leroy P. Genes coding for transcription factors involved in stem cell maintenance are repressed by TGF-β and downstream of Slug/Snail2 in COPD bronchial epithelial progenitors. Molecular Biology Reports. 2021;48(10):6729-6738.
- Rock J, Randell S, Hogan B. Airway basal stem cells: a perspective on their roles in epithelial homeostasis and remodeling. Disease Models & Mechanisms. 2010;3(9-10):545-556.
- Rigden H, Alias A, Havelock T, O'Donnell R, Djukanovic R, Davies D, et al. Squamous Metaplasia Is Increased in the Bronchial Epithelium of Smokers with Chronic Obstructive Pulmonary Disease. PLOS ONE. 2016;11(5):e0156009.
- Sohal S, Walters E. Role of epithelial mesenchymal transition (EMT) in chronic obstructive pulmonary disease (COPD). Respiratory Research. 2013;14(1):120.
- 9. Shaykhiev R, Crystal R. Basal cell origins of smoking-induced airway epithelial disorders. Cell Cycle. 2013;13(3):341-342.
- Steiling K, van den Berge M, Hijazi K, Florido R, Campbell J, Liu G, et al. A Dynamic Bronchial Airway Gene Expression Signature of Chronic Obstructive Pulmonary Disease and Lung Function Impairment. American Journal of Respiratory and Critical Care Medicine. 2013;187(9):933-942.
- Ghosh M, Miller Y, Nakachi I, Kwon J, Barón A, Brantley A, et al. Exhaustion of Airway Basal Progenitor Cells in Early and Established Chronic Obstructive Pulmonary Disease. American Journal of Respiratory and Critical Care Medicine. 2018;197(7):885-896.
- 12. Staudt MR, Buro-Auriemma LJ, Walters MS, Salit J, Vincent T,

Shaykhiev R, et al. Airway Basal stem/progenitor cells have diminished capacity to regenerate airway epithelium in chronic obstructive pulmonary disease. American Journal of Respiratory and Critical Care Medicine. 2014 Oct 15;190(8):955-8.

- 13. Shaykhiev R, Crystal RG. Early events in the pathogenesis of chronic obstructive pulmonary disease. Smoking-induced reprogramming of airway epithelial basal progenitor cells. Annals of the American Thoracic Society. 2014 Dec;11(Supplement 5):S252-8.
- 14. Vucic EA, Chari R, Thu KL, Wilson IM, Cotton AM, Kennett JY, et al. DNA methylationis globally disrupted and associated with expression changes in chronic obstructive pulmonary disease small airways. Am. J. Respir. Cell Mol Biol. 2014; 50: 912–22.
- 15. Baraldo S, Turato G, Saetta M. Pathophysiology of the small airways in chronic obstructive pulmonary disease. Respiration. 2012;84(2):89-97.
- 16. Hogg J. Peripheral lung remodelling in asthma and chronic obstructive pulmonary disease. Eur Respir J. 2004. 24(6), 893-894.
- 17. Mahmood MQ, Sohal SS, Shukla SD, Ward C, Hardikar A, Noor WD, et al. Epithelial mesenchymal transition in smokers: large versus small airways and relation to airflow obstruction. International Journal of Chronic Obstructive Pulmonary Disease. 2015;10:1515.
- Gohy ST, Hupin C, Fregimilicka C, Detry BR, Bouzin C, Chevronay HG, et al. Imprinting of the COPD airway epithelium for dedifferentiation and mesenchymal transition. European Respiratory Journal. 2015 May 1;45(5):1258-72.
- Powell HA, Iyen-Omofoman B, Baldwin DR, Hubbard RB, Tata LJ. Chronic obstructive pulmonary disease and risk of lung cancer: the importance of smoking and timing of diagnosis. Journal of Thoracic Oncology. 2013 Jan 1;8(1):6-11..
- Young RP, Hopkins RJ, Christmas T, Black PN, Metcalf P, Gamble GD. COPD prevalence is increased in lung cancer, independent of age, sex and smoking history. European Respiratory Journal. 2009 Aug 1;34(2):380-6..
- Mahmood MQ, Ward C, Muller HK, Sohal SS, Walters EH. Epithelial mesenchymal transition (EMT) and non-small cell lung cancer (NSCLC): a mutual association with airway disease. Medical Oncology. 2017 Mar;34(3):1-0.
- 22. Xu J, Lamouille S, Derynck R. TGF-beta-induced epithelial to mesenchymal transition. Cell Res. 2009. 19(2), 156-172
- 23. Gregory PA, Bracken CP, Smith E, Bert AG, Wright JA, Roslan S, et al. An autocrine TGF-β/ZEB/miR-200 signaling network regulates establishment and maintenance of epithelial-mesenchymal transition. Molecular Biology of the Cell. 2011 May 15;22(10):1686-98.
- Mahmood MQ, Reid D, Ward C, Muller HK, Knight DA, Sohal SS, et al. Transforming growth factor (TGF) beta1 and Smad signallingpathways: A likely key to EMT-associated COPD pathogenesis. Respirology. 2017. 22(1), 133-140.
- 25. Mahmood MQ, Walters EH, Shukla SD, Weston S, Muller HK, Ward C, et al. β -catenin, Twist and Snail: Transcriptional regulation of EMT in smokers and COPD, and relation to airflow obstruction. Scientific Reports. 2017 Sep 7;7(1):1-2.
- 26. Fynan TM, Reiss M. Resistance to inhibition of cell growth by transforming growth factor-beta and its role in oncogenesis. Critical Reviews in Oncogenesis. 1993 Jan 1;4(5):493-540.
- Derynck R, Akhurst RJ, Balmain A. TGF-β signaling in tumor suppression and cancer progression. Nature Genetics. 2001 Oct;29(2):117-29.

- 28. Yumoto K, Thomas PS, Lane J, Matsuzaki K, Inagaki M, Ninomiya-Tsuji J, et al. TGF-β-activated kinase 1 (Tak1) mediates agonistinduced Smad activation and linker region phosphorylation in embryonic craniofacial neural crest-derived cells. Journal of Biological Chemistry. 2013 May 10;288(19):13467-80..
- 29. Fulcher ML, Gabriel S, Burns KA, Yankaskas JR, Randell SH. Welldifferentiated human airway epithelial cell cultures. InHuman cell culture protocols 2005. 2005: 183-206.
- Mertens TC, Karmouty-Quintana H, Taube C, Hiemstra PS. Use of airway epithelial cell culture to unravel the pathogenesis and study treatment in obstructive airway diseases. Pulmonary Pharmacology & Therapeutics. 2017 Aug 1;45:101-13.
- 31. Nieto MA, Huang RY, Jackson RA, Thiery JP. EMT: 2016. Cell. 2016 Jun 30;166(1):21-45.
- 32. Rock JR, Onaitis MW, Rawlins EL, Lu Y, Clark CP, Xue Y, et al. Basal cells as stem cells of the mouse trachea and human airway epithelium. Proceedings of the National Academy of Sciences. 2009 Aug 4;106(31):12771-5.
- Hackett NR, Shaykhiev R, Walters MS, Wang R, Zwick RK, Ferris B, et al. The human airway epithelial basal cell transcriptome. PloS One. 2011 May 4;6(5):e18378.
- Tesei A, Zoli W, Arienti C, Storci G, Granato AM, Pasquinelli G, et al. Isolation of stem/progenitor cells from normal lung tissue of adult humans. Cell Proliferation. 2009 Jun;42(3):298-308.
- 35. Slabáková E, Pernicová Z, Slavíčková E, Staršíchová A, Kozubík A, Souček K. TGF Beta 1-induced EMT of non-transformed prostate hyperplasia cells is characterized by early induction of SNAI2/Slug. The Prostate. 2011 Sep;71(12):1332-43.
- 36. Geismann C, Arlt A, Bauer I, Pfeifer M, Schirmer U, Altevogt P, et al. Binding of the transcription factor Slug to the L1CAM promoter is essential for transforming growth factor-β1 (TGF-β)-induced L1CAM expression in human pancreatic ductal adenocarcinoma cells. International Journal of Oncology. 2011 Jan 1;38(1):257-66.
- 37. Joseph MJ, Dangi-Garimella S, Shields MA, Diamond ME, Sun L, Koblinski JE, et al. Slug is a downstream mediator of transforming growth factor-β1-induced matrix metalloproteinase-9 expression and invasion of oral cancer cells. Journal of Cellular Biochemistry. 2009 Oct 15;108(3):726-36.
- Mistry DS, Chen Y, Wang Y, Zhang K, Sen GL. SNAI2 controls the undifferentiated state of human epidermal progenitor cells. Stem Cells. 2014 Dec;32(12):3209-18.
- Bolós V, Peinado H, Pérez-Moreno MA, Fraga MF, Esteller M, Cano A. The transcription factor Slug represses E-cadherin expression and induces epithelial to mesenchymal transitions: a comparison with Snail and E47 repressors. Journal of Cell Science. 2003 Feb 1;116(3):499-511.