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Article

WUSCHEL Overexpression Promotes Callogenesis and Somatic Embryogenesis in *Medicago truncatula* Gaertn

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Abstract: The induction of plant somatic embryogenesis is often a limiting step for plant multiplication and genetic manipulation in numerous crops. It depends on multiple signaling developmental processes involving phytohormones and the induction of specific genes. The *WUSCHEL* gene (*WUS*) is required for the production of plant embryogenic stem cells. To explore a different approach to induce somatic embryogenesis, we have investigated the effect of the heterologous *Arabidopsis WUS* gene overexpression under the control of the jasmonate responsive *vsp1* promoter on the morphogenic responses of *Medicago truncatula* explants. *WUS* expression in leaf explants increased callogenesis and embryogenesis in the absence of growth regulators. Similarly, *WUS* expression enhanced the embryogenic potential of hairy root fragments. The *WUS* gene represents thus a promising tool to develop plant growth regulator-free regeneration systems or to improve regeneration and transformation efficiency in recalcitrant crops.

Keywords: callogenesis; embryogenic potential; growth regulators; *Medicago truncatula*; somatic embryogenesis; *WUSCHEL*



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1. Introduction

Somatic embryogenesis is a promising approach and a powerful tool for the mass propagation of plants. This process also provides a potential model to investigate the early regulatory and developmental events in plant embryogenesis [1]. The somatic embryogenesis system is characterized by a sequence of events that includes stimulation of cell proliferation, acquisition of embryogenic competence and induction of embryogenesis. Explant cells can be induced to an embryogenic state by a variety of procedures that usually include exposition to exogenous stimuli like plant growth regulators, certain stress conditions (pH shock, low or high temperature, osmotic shock, drought) or treatments with various chemical substances [2,3]. In response to these signals, somatic cells acquire an embryogenic competence resulting from the action of a complex signaling network and from the reprogramming of gene expression patterns.

Studies of factors and genes controlling *in vitro* plant morphogenesis are important for the development of improved regeneration systems and for the analysis of molecular mechanisms underlying plant embryogenesis. Genes regulating the plant stem cell development have been identified, like the *Arabidopsis* LEAFY COTYLEDON genes (*LEC1*, *LEC2*), *FUSCA3* (*FUS3*), *SOMATIC EMBRYOGENESIS RECEPTOR KINASE 1* (*SERK1*), *BABY BOOM* (*BBM*) and *WUSCHEL* (*WUS*) playing key roles in controlling embryo development [4–7]. Similarly, several transcription factors were used to induce ectopic formation of somatic embryos in *Arabidopsis*. These include LEAFY COTYLEDON genes (*LEC1*, *LEC2* and *LEC1-LIKE*), *WUSCHEL* (*WUS*), *PLANT GROWTH ACTIVATOR 37* (*PGA 37*) and *EMBRYOMAKER* (*EMK*) gene [8–12]. Similar attempts have been reported in other species using *BABY BOOM* (*BBM*) in tobacco [13], pepper [14] cacao [15] and rice [16], *LEC1* and *LEC2* in tobacco [17], *AGAMOUS-LIKE15* in soybean [18], *BBM* and *WUS2* in maize [19].

The *WUSCHEL*-related homeobox (*WOX*) gene family is a unique transcription factor family in plants and belongs to the homeobox (HB) superfamily. This *WOX* family is characterized by the phylogenetic relatedness of its homeodomains [20]. The analysis of *WOX* gene expression and function shows that *WOX* family members fulfill specialized functions in key developmental processes in plants, including embryonic development, maintenance of meristematic stem cells, development of lateral organs, seed formation and regeneration of isolated tissues and organs [21,22].

The *Arabidopsis* *WUS* gene is essential for regulating cell division and differentiation during plant development [7]. *WUS* expression is confined to a small group of cells in the lower part of the central zone of the shoot apical meristem, but can drive signals across cell layers and is expressed non autonomously [23]. The *WUS* gene is required to specify stem-cell identity and to maintain a pool of pluripotent stem cells in the shoot apical meristem (SAM). Thus, *wus* mutants fail to organize a functional SAM. During embryogenesis, *WUS* plays a key role by promoting the vegetative-to-embryonic transition and maintaining the identity of the embryonic stem cells [10,24].

Early expression of *WUS* is characteristic of somatic embryogenesis in *Arabidopsis*, *Medicago* and *Zea* [20,25,26]. *WUS* overexpression has been reported to enhance somatic embryogenesis in species such as *N. tabacum* [27], *Coffea canephora* [28], *Capsicum chinense* [29], *Picea glauca* [30], *Gossypium hirsutum* [31] and more recently in *Medicago truncatula* [32]. *WUS* also promotes the formation of embryogenic calli in *G. hirsutum* [33]. However, *WUS* overexpression can result in abnormal somatic embryos formation and can prevent seedling generation [29,30,33].

In this work, we have investigated the effect of the ectopic overexpression of the *A. thaliana* *WUS* (*AtWus*) gene expressed from the *vsp1* jasmonate inducible promoter on somatic embryogenesis from *Medicago truncatula* (Gaertn.) leaflets and hairy root segments, in the presence or absence of growth regulators. The use of the jasmonate inducible promoter *vsp1* was explored to ectopically express *WUS*. We studied the effect of *WUSCHEL* expression on callogenesis and somatic embryogenesis of plants with the aim of using it as a substitute for plant growth regulators, in the model plant for somatic embryogenesis *M. truncatula*. Remarkably, transgenic tissues (over)expressing *WUS* present an initiation of callogenesis and an increase of embryogenesis even in the absence of growth regulators in the culture medium.

2. Results

The effect of *WUSCHEL* overexpression during regeneration via somatic embryogenesis was discernable when comparing *WUS*-expressing explants to control explants transformed with the pCambia-*bar* vector. Developmental and morphological characteristics were observed from the appearance of proembryogenic calli till the development of somatic embryos. The transgenic status of the plants selected by phosphinothricin was confirmed by polymerase chain reaction (PCR) amplification of the *BAR* and *WUS* sequences.

2.1. The Designed Construct Allows WUS Gene Expression but Not Induction by Jasmonate

The leaves and root segments transformed by *WUS* as well as those of the control were cultured on media treated with or without jasmonate for callogenesis and embryogenesis induction. In order to follow the expression of the transgene in the explants, we tested *WUS* expression via quantitative real-time polymerase chain reaction (qRT-PCR) in 7- and 14-day-old calli formed in the presence or absence of jasmonate. The experiment showed that the *WUS* relative expression level was not significantly different between transgenic calli produced in the presence or absence of jasmonate on leaf explants (2.7 and 2.4, respectively) and on roots (2.4 and 2.1, respectively) ($p > 0.05$). However, *WUS* calli and embryos showed a significantly higher expression level than the calli transformed with pCambia1301-bar (0.07) used as negative controls demonstrating expression of the transgene. We deduced from this experiment that the *vsp1-WUS* gene was expressed in the presence and in the absence of the inducible agent (jasmonate). Accordingly, no significant differences were observed in the callogenesis and embryogenesis of *WUS* explants (leaves and root segments) cultured on media with or without jasmonate (see below). The results presented below are those obtained for *WUS* explants and controls cultured in the presence of jasmonate.

2.2. WUSCHEL Enhances Callogenesis in *M. truncatula* Leaf Explants

In order to test the effect of the *WUS* gene expression on *Medicago* regeneration, leaf fragments were tested for regeneration on media with SH1 medium (supplemented with $4 \text{ mg}\cdot\text{L}^{-1}$ 2,4-D and $0.5 \text{ mg}\cdot\text{L}^{-1}$ BAP) or without plant growth regulators (SH0).

On the media without plant growth regulators (SH0), only *WUS* leaves produced calli while the controls did not produce any callus. Strikingly, these *WUS* leaves began callogenesis after 2 weeks of culture and presented a percentage of callus production reaching 59%. Calli formed small clusters near leaf incisions. This result shows that the expression of the *WUS* gene is alone able to induce cell proliferation in absence of plant growth regulators.

As for the leaf explants cultured on SH1, calli production took place two weeks earlier on *WUS* leaf explants (within 10 days of culture) than on the controls (three weeks of culture). The percentage of explants giving rise to calli was statistically higher in *WUS* transgenic leaves ($p < 0.1$), with 74% of *WUS* leaves producing calli clusters against 46% for the controls (Figure 1). Calli were observed around the edge of the incisions and later covered the entire leaf surface, forming large embryogenic cell clusters. These calli were translucent-white and friable [34]. Thus, calli production was also enhanced on *WUS* expressing leaf explants in presence of callus inducing plant growth regulators.

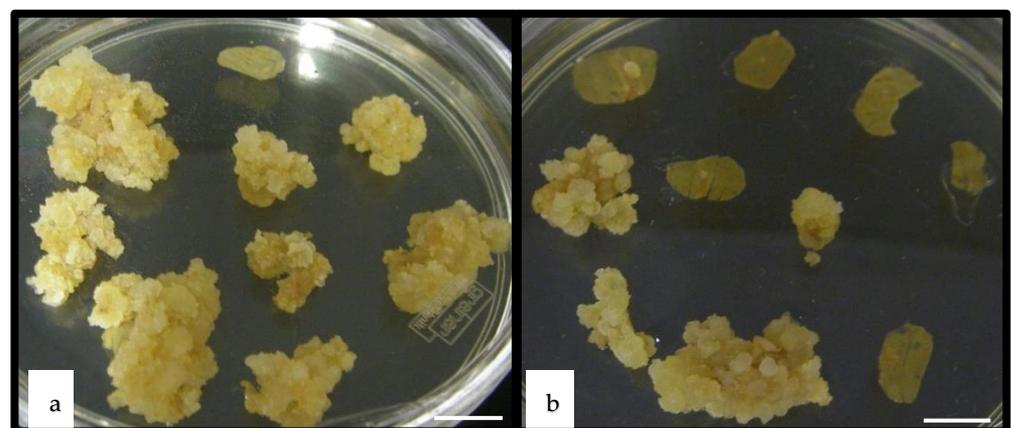


Figure 1. Callus production from *Medicago truncatula* leaf explants. Callus production from *Medicago truncatula* leaf explants transformed with pCambia1301-*WUS*-BAR leaves (a) and from leaf explants transformed with pCambia-BAR (control, b), cultured on SH1 medium ($4 \text{ mg}\cdot\text{L}^{-1}$ 2,4-D, $0.5 \text{ mg}\cdot\text{L}^{-1}$ BAP) during 3 weeks. Scale bar represent 1.0 cm.

2.3. WUSCHEL Enhances Embryogenesis in *M. truncatula* Leaf Explants

The *WUS* calli produced on SH0 and transferred to embryogenic medium (SH2 without growth regulators) started to produce embryos after three weeks of culture. Only 7.5% of these calli were embryogenic and produced on average 26 embryos per calli.

As early as two weeks after the transfer of calli cultured on SH1 to embryogenic medium (SH2 without growth regulators), early-stage embryos were produced on cultures transformed with the *WUS* construct. Somatic embryos started to form one week later on the control explants. The histological longitudinal section of a *WUS* embryonic calli showed these embryos of different stage of development (Figure 2). After 4–5 weeks of culture, the percentage of embryogenic calli was not significantly different between *WUS* calli and the controls. In fact, 55% of the *WUS* transgenic calli were embryogenic against 57% for the control explants (Figures 3 and 4). There were also no differences in the production and the phenotype of the embryonic mass between empty vector transformation and non-transformed tissue. However, the average number of embryos produced from each responsive *WUS* calli (120 embryos) was almost 2.3-fold higher than that of control (51 embryos) (Figure 5). Together, these results showed that *WUS* expression allowed embryogenesis from leaf explants cultured in absence of plant growth regulators in the culture medium and increased embryogenesis for leaf explants cultured in the presence of growth regulators.

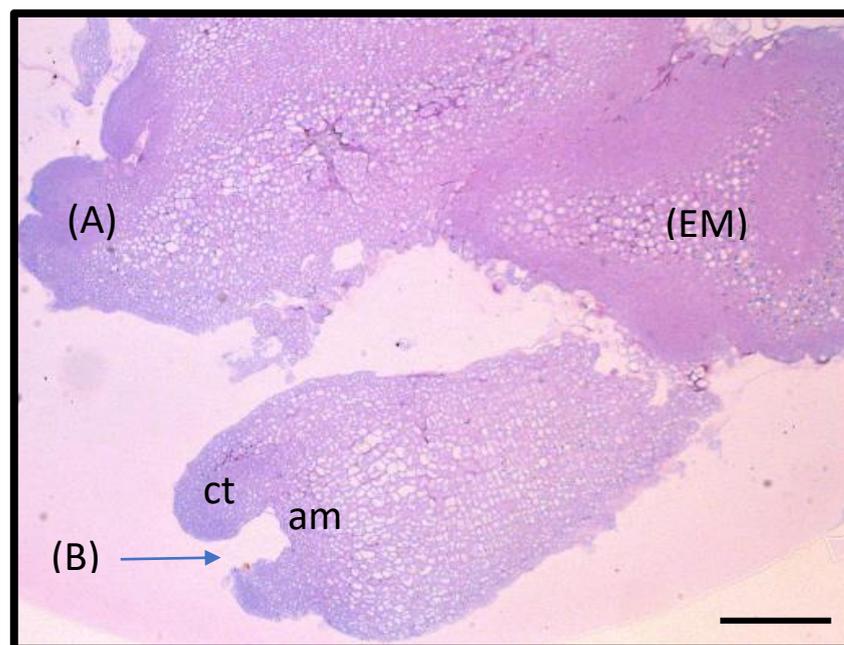


Figure 2. Histological longitudinal section of *WUS* embryogenic calli (EM) after 3 weeks of culture on SH1 medium showing the produced embryos at various stages of development: (A) globular stage somatic embryo; (B) cotyledonary-stage somatic embryo with two distinct cotyledons (ct) and apical meristem (am). The bar scale represents 200 μ m

2.4. WUSCHEL Enhances Callogenesis in *M. truncatula* Hairy Root (HR) Explants

In order to test the effect of the *WUS* gene expression on regeneration of *Medicago* hairy root (HR) fragments regeneration, root segments of *M. truncatula* were first cultured under two different regeneration conditions, either directly on M1 medium under photoperiod or first on a callogenesis C medium in the dark for two weeks. The explants from the C medium were then transferred to M0 (without growth regulators) or M1 media. *WUS* and control root explants showed callus production from the first week of culture. Calli were produced along the entire length of the root segments on the two media but the callus

sizes of the root segments grown in the dark (C medium) were bigger than those under the photoperiod (Figure 6).

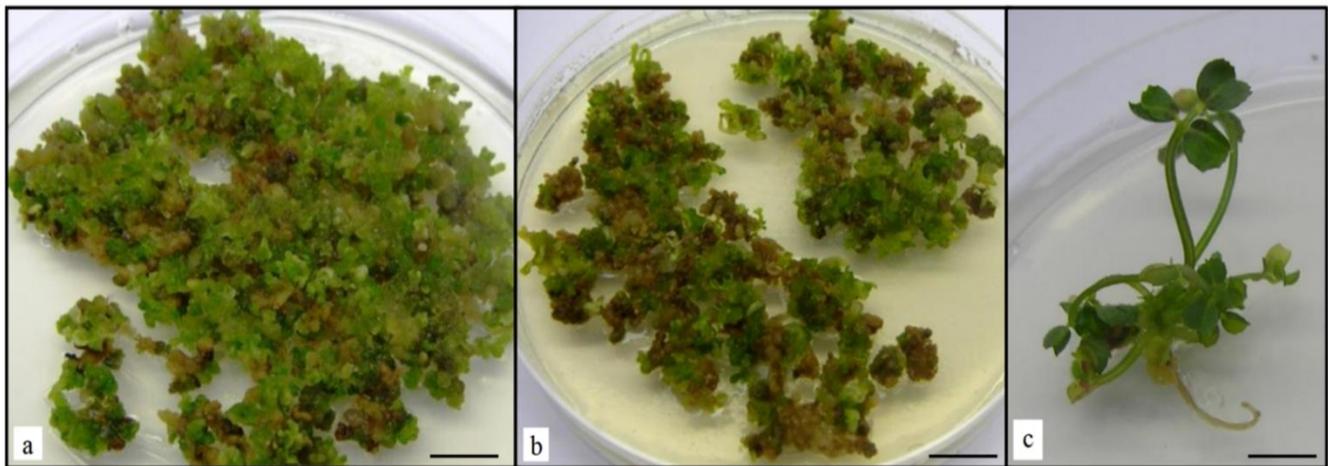


Figure 3. Embryo production from *Medicago truncatula* leaf explants. Embryos production on embryogenic calli formed from *Medicago truncatula* leaf explants transformed with pCambia1301-*WUS*-*BAR* (a) and from leaf explants transformed with pCambia-*BAR* (control) (b) after four weeks of culture on SH2 medium (without plant growth regulators). Plantlet developed from a *WUS* embryo of *Medicago* on SH2 medium (c). Bar 1.0 cm.

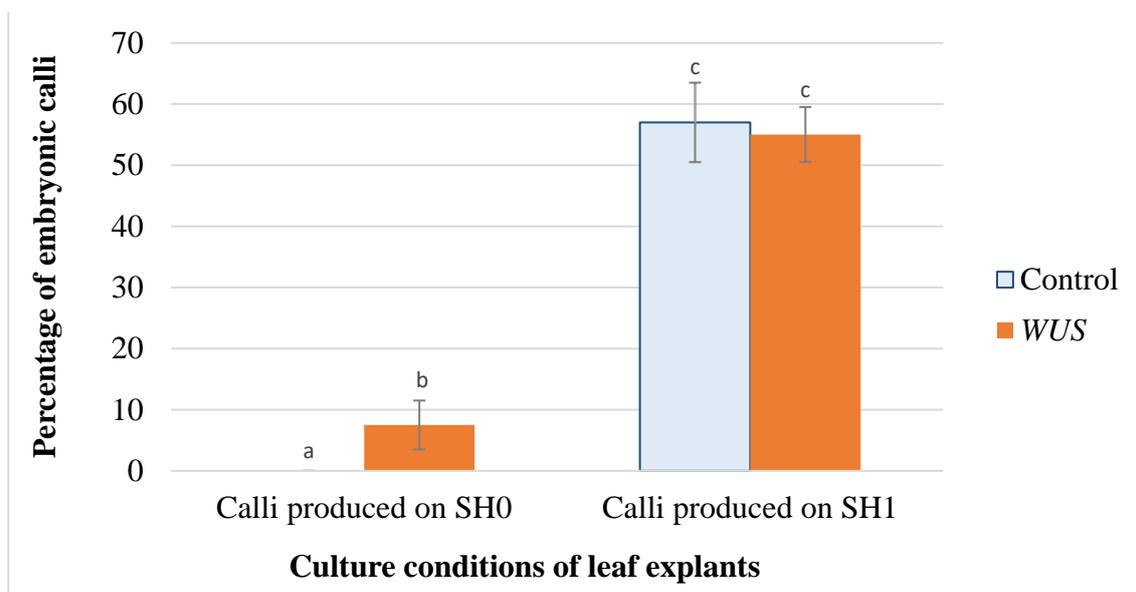


Figure 4. Embryogenic calli production from leaf explants of *Medicago truncatula*. Percentage of embryonic calli formed on leaf explants of *M. truncatula* transformed using pCambia-*BAR* (Control) and pCambia1301-*WUS*-*BAR* 1 (*WUS*) cultured on SH2 (without growth regulators). These calli were produced on SH0 (without growth regulators) or on SH1 (4 mg·L⁻¹ 2,4-D, 0.5 mg·L⁻¹ BAP). For each condition, 60 root explants were cultured per replicate. Three replicates were performed for each condition. Bars with different letters were significantly different at $p < 0.05$.

2.5. *WUSCHEL* Enhances Embryogenesis in *M. truncatula* HR Explants

Calli produced on HR segments grown directly on M1 medium under photoperiodic conditions started to produce proembryos during the second week of culture. After 5 weeks of culture, *WUS* calli showed a significantly higher embryogenic percentage (92%) than controls (57%). In addition, *WUS* calli produced a higher mean number of embryos (9.8 embryos per explant) as compared to the controls (3.8 embryos/explant; Figures 7–9). Thus,

the expression of the *WUS* gene strongly enhances embryogenesis in these HR fragments in the absence of the callogenesis step.

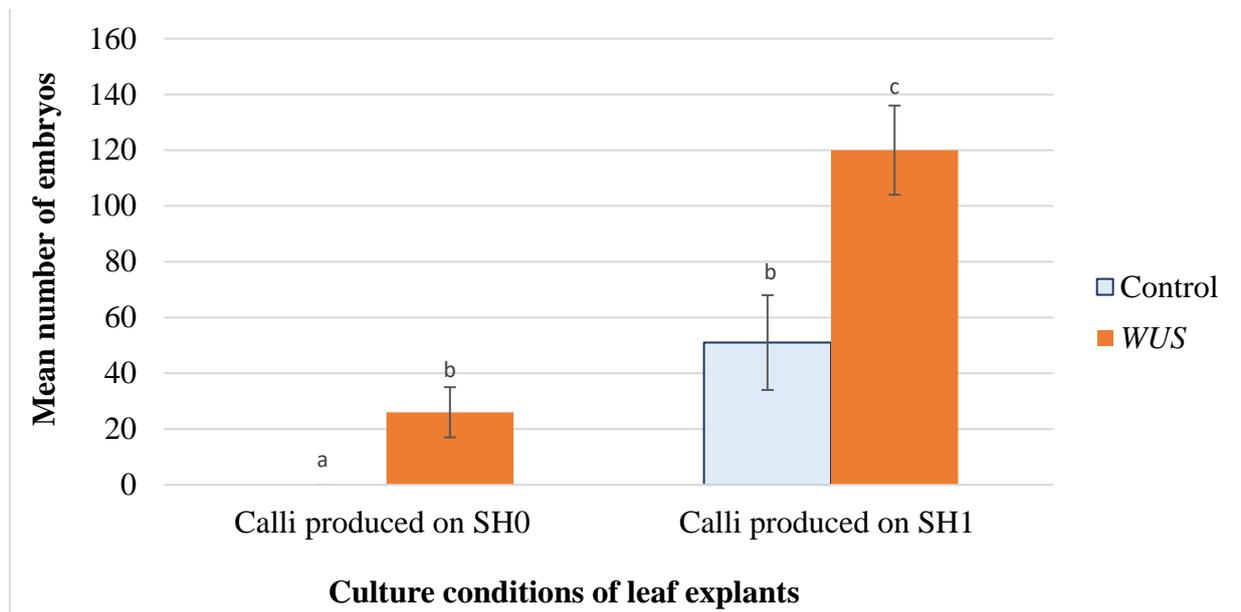


Figure 5. Embryo production from leaf explants of *Medicago truncatula*. Mean number of embryos produced on calli formed leaf explants of *M. truncatula* transformed using pCambia-BAR (Control) and pCambia1301-*WUS*-BAR I (*WUS*) cultured on SH2 (without growth regulators). These calli were produced on SH0 (without growth regulators) or on SH1 (4 mg·L⁻¹ 2,4-D, 0.5 mg·L⁻¹ BAP). Data represent the mean ± standard errors of three replicates. Bars with different letters were significantly different at $p < 0.05$.

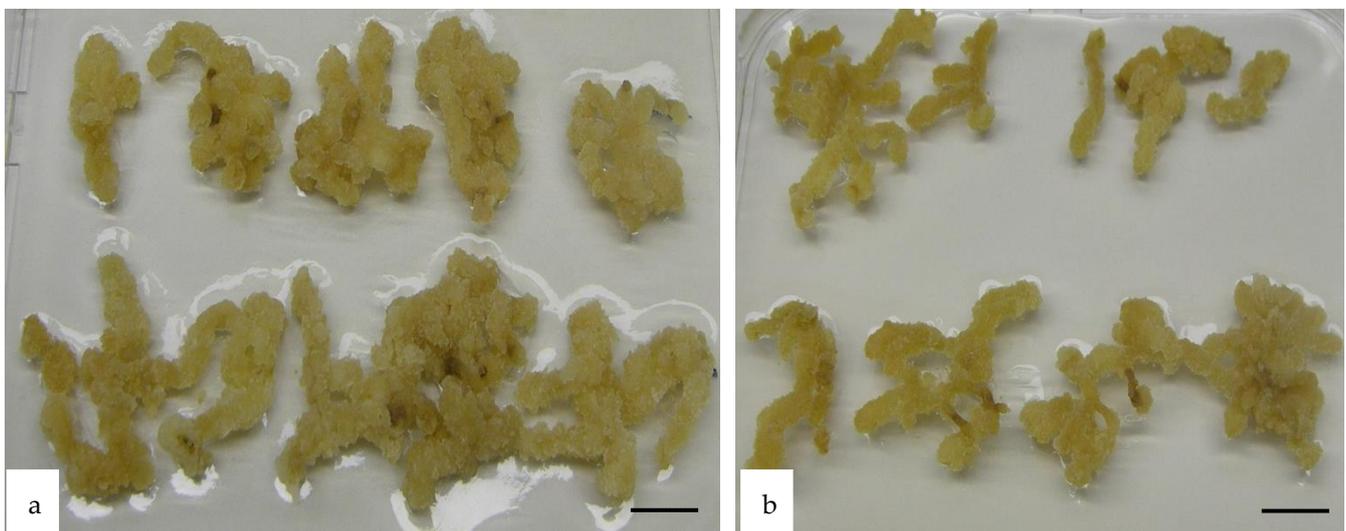


Figure 6. Callus production from *Medicago truncatula* hairy root (HR) explants. Callus production from *Medicago truncatula* leaf explants, transformed with pCambia1301-*WUS*-BAR, two weeks after their transfer to M0 medium (without growth regulator). These calli were produced on a callogenesis medium C (5 mg·L⁻¹ 2,4-D and 0.5 mg·L⁻¹ BAP) in the dark for two weeks (a) or under photoperiod on M1 medium (50 mg·L⁻¹ NAA and 1.5 mg·L⁻¹ BAP). Scale bar 1.0 cm (b).

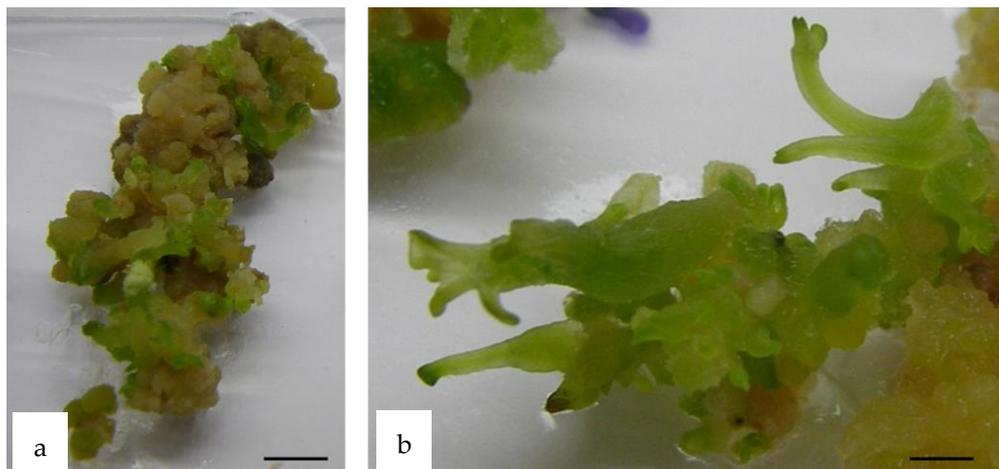


Figure 7. Embryo production from *Medicago truncatula* HR explants. (a) *WUS* embryogenic calli produced on root segments of *M. truncatula* after 3 weeks of culture and (b) *WUS* somatic embryos at different developmental stages (c: cotyledonary stage and g: globular stage) produced on embryogenic calli after 4 weeks of culture on M0 medium (without plant growth regulators). Bar 1.0 cm (a) Bar 0.5 cm (b).

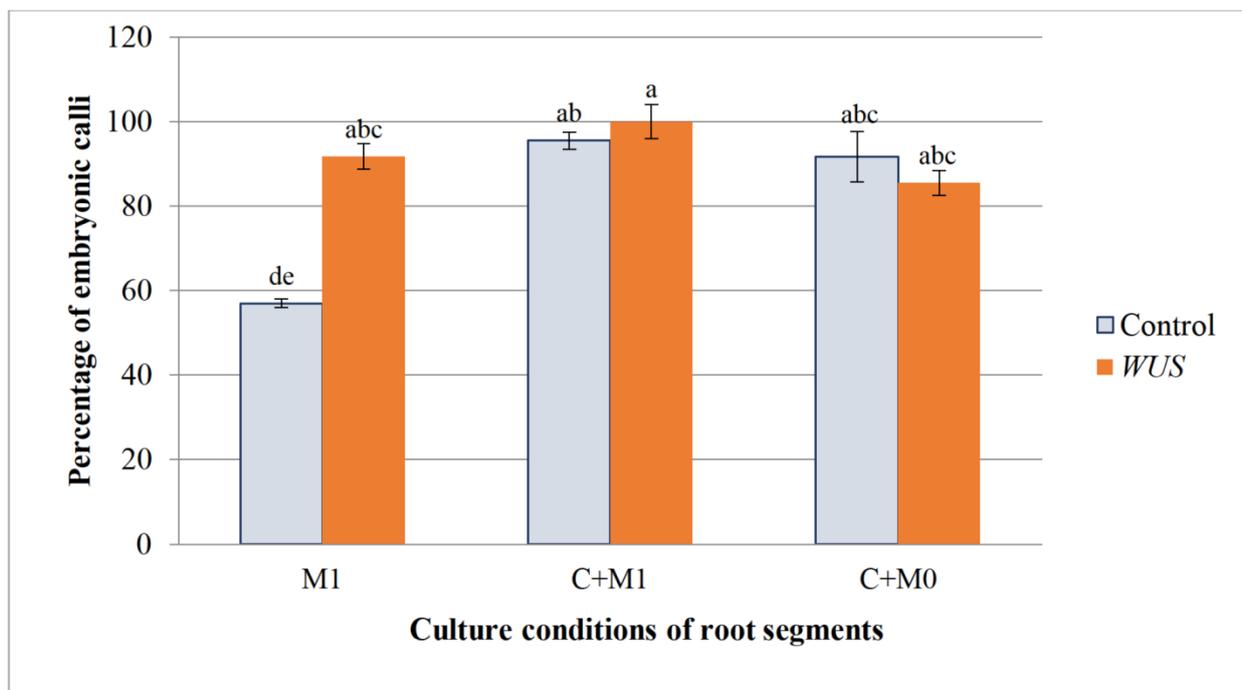


Figure 8. Embryogenic calli production from HR fragments of *Medicago truncatula*. Percentage of embryonic calli formed on hairy root segments of *M. truncatula* transformed by pCambia-BAR (Control) and pCambia1301-*WUS*-BAR I (*WUS*) cultured in 3 different conditions: directly on M1 medium containing $50 \text{ mg}\cdot\text{L}^{-1}$ NAA and $1.5 \text{ mg}\cdot\text{L}^{-1}$ BAP (M1); first, a callogenesis stage on C medium ($5 \text{ mg}\cdot\text{L}^{-1}$ 2,4-D and $0.5 \text{ mg}\cdot\text{L}^{-1}$ BAP) and then transferred to M1 medium (C+M1); first a callogenesis stage on C medium and then transferred to M0 medium without growth regulator (C+M0). For each condition, 60 root explants were cultured per replicate. Three replicates were performed for each condition. Bars with different letters were significantly different at $p < 0.05$.

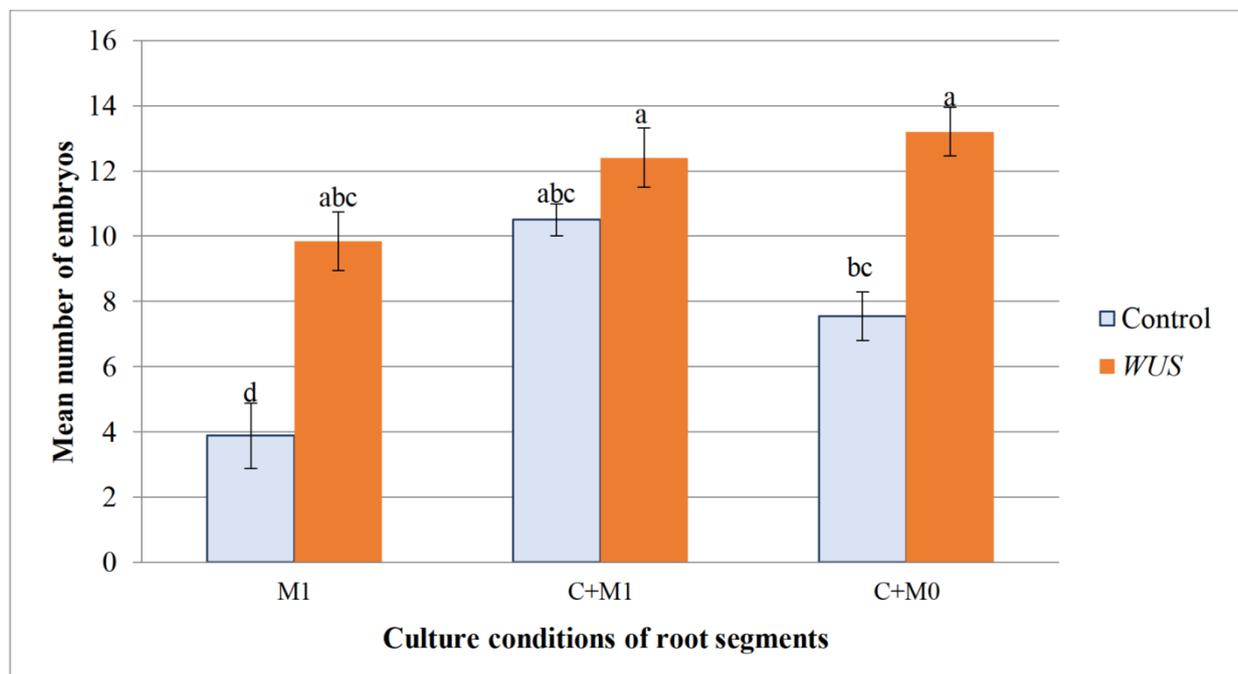


Figure 9. Embryo production from HR fragments. Mean number of embryos produced on calli formed on hairy root segments of *M. truncatula* transformed by pCambia-BAR (Control) and pCambia1301-WUS-BAR 1 (*WUS*) cultured in 3 different conditions: directly on M1 medium containing $50 \text{ mg}\cdot\text{L}^{-1}$ NAA and $1.5 \text{ mg}\cdot\text{L}^{-1}$ BAP (M1); first, a callogenesis stage on C medium ($5 \text{ mg}\cdot\text{L}^{-1}$ 2,4-D and $0.5 \text{ mg}\cdot\text{L}^{-1}$ BAP) and then transferred to M1 medium (C+M1); first a callogenesis stage on C medium and then transferred to M0 medium without growth regulator (C+M0). Data represent the mean \pm standard errors of three replicates. Bars with different letters were significantly different at $p < 0.05$.

For the explants first cultured two weeks in the dark on the callogenesis C medium, embryogenesis did not start until a week after the calli were transferred to M0 or M1 media under the photoperiod. Both controls and *WUS* calli developed a high percentage of embryogenesis. Under these two culture conditions (M0, M1), the percentage of embryonic calli was not significantly different between *WUS* calli and the controls (respectively 100% and 95.5% on M1; 85.5% and 91.7% on M0) (Figure 8). Similarly, *WUS* and control calli produced a high number of embryos (12.4 and 10.5 embryos respectively) following their transfer on M1. In contrast, on the hormone-free M0 medium the number of embryos produced per *WUS* callus (13.2 embryos) was higher than that of the control (7.5 embryos) (Figure 9). It should be noted that under these three culture conditions, *WUS*-expressing HR explants maintained a high number of embryos per explants, whereas the controls showed a significant decrease of the embryos' mean number when they were cultured directly on M1 or M0 media after a callogenesis step. Again, the expression of the *WUS* gene favored embryogenesis in the presence or absence of plant growth regulators.

2.6. Embryo Development and Plantlet Production

The *WUS* and control embryogenic calli followed the typical stages of embryogenesis observed in *Medicago truncatula* [34,35]. They presented a proliferation of embryonic clumps and differentiation of somatic embryo simultaneously (Figure 7). Less than 25% of the produced somatic embryos, from both *WUS* and *BAR* transgenic embryos, were abnormal and later degenerated. Both *WUS* and control somatic embryos produced normal plantlets after being transferred to maturation medium (1/2 MS medium). These plantlets did not show any morphological abnormalities even though they expressed the transgenic *WUS* gene in their leaves (not shown).

3. Discussion

Stem cell induction and somatic embryo development involve complex mechanisms including the central role of growth regulators and transcriptional regulators. Mendez-Hernandez et al. [2] emphasized the interactions between the different plant growth regulators during the induction of somatic embryogenesis. Rose [36] suggested the presence of connections between specific plant growth regulators and up-regulated genes during early phases of somatic embryogenesis in *M. truncatula*. The *WUS* gene is essential for somatic embryogenesis and it is designated as being a primary candidate for a gene promoting regeneration in a wide range of species [10,37]. Overexpression of *WUS* has indeed been found to improve the embryonic potential in transgenic plants [10,24,32]. In this work, we used a different approach to induce somatic embryogenesis in order to overcome the hormone dependence of this process. For this, we studied the effect of *WUS* ectopic expression on the somatic embryogenesis capacity of *M. truncatula* (Gaertn.) leaf and hairy root explants under different tissue culture conditions, including plant growth regulators free tissue culture conditions.

Previous studies have shown that the continuous overexpression of *WUS* resulted in malformations and growth alteration [10,33]. Moreover, the level of *WUS* transgene expression was related to the frequency of somatic embryos with aberrant phenotype in white spruce [30]. Similarly, calli produced on *WUS* expressing tobacco explants maintained in induced condition, darkened and the frequency of regeneration was reduced [37]. An inducible system based on the estrogen 17- β -estradiol was used to trigger the expression of *WUS* in different plants [10]. However, somatic embryos formed in the absence of the estrogen in coffee and white spruce [28,30]. In our study, we planned to use a jasmonate-inducible *WUS* gene expressed from the *vsp1* promoter. The aim was to transitory express *WUS* in order to trigger a developmental switch to embryonic callogenesis. Jasmonate was supposed to induce ectopic expression of *WUS* without affecting the in vitro development and growth of transgenic *M. truncatula* explants and embryos. However, in our experiment, *WUS* gene expression was also observed in the absence of jasmonate. According to Arroyo-Herrera et al. [28] using a 17- β -estradiol inducible system, the transcription could be attributed to the position effect of the T-DNA insertion or to a cross reactivity of the inducer receptor (estrogen) interacting with an estrogen-like endogenous molecule. Similar position effect or production of jasmonate in tissue culture can explain our results. The constitutive expression of *WUS* could be due to the influence of the 35S promoter enhancer located next to the *vsp1* promoter in the plasmid construct. *WUS* expression could be also induced by endogenous jasmonate produced as a response to the stress subjected to explants cultured in vitro.

Plant growth regulators are required for the induction of somatic embryogenesis from cultured explants in the majority of plants [38]. In many in vitro culture systems and especially for *Medicago* leaf culture, auxin is required as a pulse to induce callus production. High levels of auxin in the culture medium promote cell proliferation and embryogenic callus formation. The produced calli are then transferred to auxin-free medium for the formation of somatic embryos [34,39]. In our study, the expression of *WUS* induced spontaneous embryogenic calli from *Medicago* leaves grown on plant growth regulators-free basal medium. Only *WUS*-expressing leaves produced embryogenic calli in the absence of growth regulators while leaves transformed with the empty vector did not produce any callus cluster. This is consistent with Chen et al. [25] who suggested that *WUS* is associated with the production of totipotent cells, similar to the way it is involved in stem cell formation and maintenance. Similarly, leaves-derived calli expressing *WUS* produced a higher percentage of embryogenic calli and showed a 2.3-fold improvement (120 embryos versus 51) in somatic embryo production as compared to control, on media without growth regulators. These experiments showed that *WUS* transgenic embryos could be obtained from *Medicago* leaves explants without the use of growth regulators. Zuo et al. [10] suggested that *WUS* can reprogram cell identity, bypassing the auxin requirement or simply taking advantage of the endogenous auxin flux. However, Gallois et al. [24]

reported that ectopic expression of *WUS* in roots generated embryo-like structures only in the presence of auxin. In *Arabidopsis thaliana*, *WUS* transient overexpression caused highly embryogenic callus formation in the presence of auxin [10]. The expression of *AtWUS* induced calli formation as well as a 400% increase in somatic embryo production in coffee plants and a threefold increase in cotton explants in the presence of plant growth regulators [28,31]. Recently, Tvorogova et al. [32] also reported that the overexpression of *MtWOX9-1*, a *WUSCHEL*-related homeobox transcription factor, led to an increase in embryogenic capacity of calli produced from leaf explants in *Medicago*. However, these calli were produced on a callogenesis-inducing medium in the presence of plant growth regulators before being transferred to a plant growth regulator-free medium.

Hairy root (HR) cultures show interesting features such as high growth rate on hormone-free media and genetic stability. HR cultures have also shown promising biosynthetic ability as producers of various biologically active substances and provided insights into root metabolism [40,41]. Moreover, *A. rhizogenes*-mediated hairy root transformation systems are suitable for species recalcitrant to transformation by *A. tumefaciens*, as higher transformation efficiencies are obtained in comparison to *A. tumefaciens*-mediated transformation systems [42]. They may also help faster production of mutants using the CRISPR/CAS9 technology [43]. With the aim of enhancing HR callogenesis and embryogenesis, we studied the effect of *WUS* expression on HR. HR regeneration however requires specific protocols [35]. Using the HR explants with standard in vitro culture protocol, *WUS* and control *Medicago* explants showed similar embryogenesis potential. However, under culture conditions that normally do not support embryogenesis (without the passage through a callogenesis phase or in absence of growth regulators in the embryonic media), *WUS* calli produced a high number of embryos per explant, while the number of embryos produced by control calli was very low. This result showed that somatic embryos can be produced on explants expressing *WUS*, bypassing the need for callus initiation and maintenance.

Chen et al. [25] found that during callogenesis, the clusters of cells expressing *WUS* were the source of cells that formed embryos. Moreover, the ectopic expression of *WUS* resulted in up regulation of other embryogenic regulators such as *LEC1*, *LEC2*, and *FUS3* [44]. Identifying these regulators and elucidating their exact roles in embryogenesis will advance the molecular understanding of plant embryogenesis.

4. Materials and Methods

4.1. Construction of Binary Plasmids

Two binary vectors were generated, pCambia1301-*bar* and pCambia1301-*WUS-bar*. These plasmids contain the reporter gene (*GUS*) and the plant selectable marker gene conferring resistance to phosphinothricin (*BAR*), both expressed from the CaMV35S promoter. In addition, pCambia1301-*WUS-bar* contains the *WUSCHEL* gene under control of the *vsp1* jasmonate inducible promoter (Figure 10).

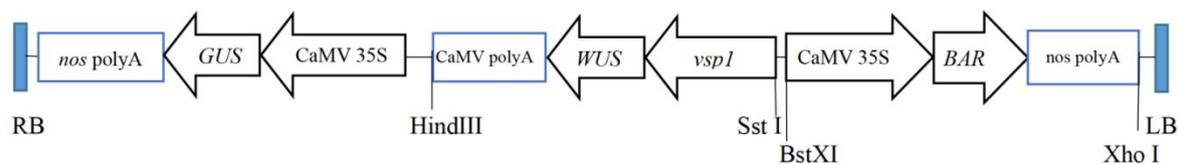


Figure 10. Organization of the T-DNA of plasmid pCambia1301-*WUS-bar*. Arrows represent promoters and coding sequences. Rectangles represent polyA regions. Restriction sites used for the construction are also indicated.

The vector pCambia1301-*bar* was generated by using pCambia 1301/Pubi-*bar*-Tnos (provided by AFOCEL, Nangis, France) containing the *BAR* gene under the control of the *Pubi* promoter [45]. To place the *BAR* gene under control of the CaMV35S promoter, the *BAR*::Tnos was recovered from pCambia 1301/Pubi-*bar*-Tnos and inserted into the pBLTI221 vector [46] under the control of CaMV35S promoter using *Bam*HI and *Eco*RI.

Restriction sites *Bst*XI and *Xho*I were present on either sides of the cassette CaMV35S-*bar*-*Tnos*. The *hpt* gene was removed from pCAMBIA1301 (Center for the Application of Molecular Biology to the International Agriculture of Canberra, Australia) by digesting the plasmid with *Bst*XI/*Xho*I and the CaMV35S: *bar* gene, was ligated into the digested pCambia1301 plasmid, between the *Bst*XI and *Xho*I sites.

As a first step to construct pCambia1301-*WUS*-*bar*, the *WUS* coding sequence was amplified from cDNAs synthesized from RNA extracted from *Arabidopsis thaliana* Columbia stem tips carrying flower buds, flowers and young siliques, using the 5'-GAGCTGCAGAAC AATGGAGCCGCCACAGC-3' and 5'-GAGGAATTCTAGTTCAGACGTAGCTC-3' primers. The PCR product was inserted into pGEM[®]-T Easy (Promega) and the *WUS* sequence was verified by nucleotide sequencing. To place the *WUS* coding sequence under the control of the *vsp1* promoter, an expression cassette was constructed as follows. The *vsp1* promoter, recovered from pAM35 [47] as an *Sst* I-*Pst* I fragment, was inserted into pJIT117 [48] in place of the 35S promoter, generating pAM44. The *WUS* coding sequence was inserted into pAM44 using *Pst* I and *Eco*RI, generating p44wus. In this construct, the *WUS* coding sequence is placed between the *vsp1* promoter and the CaMV polyadenylation signal. The *Bgl* II fragment of p44wus carrying the *vsp1*-*WUS*-CaMV polyA fusion was inserted into the *Bam*HI site of pUC19, generating pUC44wus. The gene fusion was then moved into the binary vector pCambia 1301 *bar* using *Sst* I and *Hind*III, generating pCambia1301-*WUS*-*bar*. Molecular cloning was performed according to standard methods [49].

4.2. Bacterial Strains and Growth Conditions

The pCambia1301-*bar* and pCambia1301-*WUS*-*bar* vectors were separately transferred into *Agrobacterium tumefaciens* strains AGL0 for *M. truncatula* leaf transformation and into *Agrobacterium rhizogenes* strain A4Tc24 for *M. truncatula* hairy root transformation, using a CaCl₂ method [50]. For each condition, cultures of *A. tumefaciens* were initiated from a single colony or from glycerol stocks and grown overnight at 28 °C with shaking (150 rpm) in liquid YEB medium [51] containing 50 mg·L⁻¹ kanamycin, to mid log phase (OD₆₀₀ = 0.9–1.2). For leaf coculture, the *A. tumefaciens* cells strain AGL0 were collected by centrifugation and resuspended in SH based liquid inoculation medium [52]. For radicle coculture, a single resistant colony of A4Tc24 was streaked on solid YEB medium containing 100 mg·L⁻¹ kanamycin and incubated at 28 °C for two days.

4.3. *Agrobacterium tumefaciens*-Mediated Leaf Transformation of *M. truncatula*

Agrobacterium tumefaciens-mediated transformation and regeneration via somatic embryogenesis of *M. truncatula* was done as described by Trinh et al. (1998) and Cosson et al. (2015) [34,39]. Plantlets of *M. truncatula* (Gaertn.) line R108-1 (c3) [39,53] were grown in Magenta boxes containing half strength of SH based media [52]. The plantlets were cultured under a 16 h light (200 μE/m²/s)/8h dark photoperiod at 24 ± 2 °C. Leaves from 2- to 3-week-old in vitro plantlets were cut off and each foliole was wounded with 3 to 4 scalpel cuts.

Leaf explants were transferred into two flasks, one containing a suspension of AGL0 cells carrying pCambia1301-*bar* and the other containing AGL0 with pCambia1301-*WUS*-*bar*. A vacuum (760 mm Hg) was generated in the flasks for 20 min using a tap water pump.

4.4. Coculture and Culture Media of *M. truncatula* Leaves

The infected explants were placed adaxial side facing up on the SH0 and SH1 coculture media. These media are based on SH media (N6 macroelements, SH microelements, SH vitamins, 100 mg·L⁻¹ *myo*-inositol, 30 g·L⁻¹ sucrose, and 3 g·L⁻¹ phytigel (Sigma-Aldrich, Saint-Louis, Missouri, USA), pH 5.8) and differ respectively by the absence of plant growth regulators (SH0) or the presence of 2,4-D at 4 mg·L⁻¹ and of BAP at 0.5 mg·L⁻¹ (callogenic medium SH1). For each condition, 60 explants were incubated in the dark at 20 °C for 2 days and then cultured in the dark on selection SH0 and SH1 media supplemented with

augmentin at $800 \text{ mg}\cdot\text{L}^{-1}$ to suppress *Agrobacterium* growth and with phosphinothricin at $3 \text{ mg}\cdot\text{L}^{-1}$ to select transformed plant cells.

After 5 weeks of culture, explants and their developing calli were transferred onto SH2 embryonic medium (SH0 medium with $20 \text{ mg}\cdot\text{L}^{-1}$ of sucrose and without plant growth regulators), supplemented with augmentin at $800 \text{ mg}\cdot\text{L}^{-1}$ and phosphinothricin at $3 \text{ mg}\cdot\text{L}^{-1}$. The explants (10 explants per Petri dish) were cultivated at 24°C under a photoperiod of 12 h ($75\text{--}110 \mu\text{E m}^{-2} \text{ s}^{-1}$).

4.5. *Agrobacterium rhizogenes*-Mediated Hairy Root Production in *M. truncatula*

Agrobacterium rhizogenes-mediated transformation was done as described by Boisson-Dernier et al. (2001) [54]. Scarified and surface sterilized seeds of *M. truncatula* (Gaertn.) line R108-1 (c3) [53] were germinated on inverted agar plates at 14°C in the dark. After 2 days of germination, the radicle was sectioned approximately 3 mm from the root tip with a sterile scalpel. Sectioned radicles were inoculated by coating the freshly cut surface with *A. rhizogenes* A4Tc24 carrying pCambia1301-*bar* or pCambia1301-*WUS-bar* grown on YEB solid medium [51]. The inoculated sectioned seedlings were then placed on Fahraeus medium [55] in square Petri dishes ($12 \times 12 \text{ cm}$). The Petri dishes containing the inoculated seedlings (10 seedlings per Petri dish) were placed at an angle of approximately 45° in a growth chamber at 20°C for one week (16 h light/8 h dark photoperiod and a light intensity of $75 \mu\text{E m}^{-2} \text{ s}^{-1}$). After 7 days of co-culture, seedlings were transferred onto $1/2$ MS medium [56] supplemented with Augmentin at $400 \text{ mg}\cdot\text{L}^{-1}$ for HR development.

4.6. Culture Media of *M. truncatula* Hairy Roots

For HR regeneration, hairy root fragments of approximately 1 to 2 cm in length were excised and sub-cultured every 2 weeks in horizontal Petri dishes under different culture conditions:

(a) Directly on MS medium [56] containing $50 \text{ mg}\cdot\text{L}^{-1}$ NAA and $1.5 \text{ mg}\cdot\text{L}^{-1}$ BAP (M1 medium) under a 16 h light/8 h dark photoperiod.

(b) On a callogenesis inducing MS medium [56] containing $5 \text{ mg}\cdot\text{L}^{-1}$ 2,4-D and $0.5 \text{ mg}\cdot\text{L}^{-1}$ BAP (C medium) for two weeks in the dark. Then, the produced calli were transferred to two media: MS medium without plant growth regulators (medium M0) or MS medium containing $50 \text{ mg}\cdot\text{L}^{-1}$ NAA and $1.5 \text{ mg}\cdot\text{L}^{-1}$ BAP (medium M1) and placed under a 16 h light/8 h dark photoperiod.

4.7. Induction of *WUS* Transgene Expression

For enhancing *WUS* expression, sterile capsules containing 10 mM of methyl jasmonate ($100 \mu\text{L}$) were placed in each Petri dish. Explants were treated with jasmonate from the first day of culture till the production of the first proembryos, i.e., for at least two to three weeks.

4.8. Molecular Analysis, Polymerase Chain Reaction (PCR) Analysis

Total genomic DNA was isolated from leaves and roots of transformed plants and control plants using the DNeasy plant Mini Kit (Qiagen, Les Ulis, France) according to the manufacturer recommendations. Polymerase chain reaction (PCR) amplification was performed in a $20 \mu\text{L}$ reaction volume consisting of 10X buffer (Promega, Madison, WI, USA), 50 mM KCl, 1.5 mM MgCl_2 , 100 mM dNTPs, 0.5 U Taq DNA polymerase (Promega, Madison, WI, USA), 250 nM primers, and 20 ng template DNA. The primers 5'-GCCATTTGAAGCCGATGTCACGCC-3' and 5'-GTATCGGTGTGAGCGTCGCAGAAC-3' were used to amplify a 1050 bp *GUS* fragment. The primers 5'-CTACATCGAGACAAGCACGGTCAA-3' and 5'-GCTGAAGTCCAGCTGCCAGAAA-3' were used to amplify a 427 bp *BAR* fragment. The primers 5'-CCGCCACAGCATCAGCATCAT-3' and 5'-CACCACATTCA GTACCTGAGCT-3' were used to amplify a 529 bp the *WUS* fragment. Cycling parameters for *BAR* and *WUS* amplification began with an initial denaturation at 94°C for 2 min, followed by 30 cycles of denaturation (94°C for 2 min), annealing (55°C for 30 s) and extension (60°C for 30 s), then a final extension at 72°C for 5 min. The cycling parameters

for *GUS* amplification were similar except that the annealing temperature was 60 °C. PCR amplification products were analyzed by electrophoresis in 1% agarose gels.

4.9. Real-Time PCR (qRT-PCR) Analysis

WUS expression was detected by RT-PCR. Total RNA was extracted from calli and embryos of *M. truncatula* plants transgenic for *WUS* at different time points using the RNeasy Mini Kit (Qiagen, Les Ulis, France) according to the manufacturer instructions. cDNA synthesis was performed using the QuantiTect[®] Reverse Transcription (Qiagen, Les Ulis, France) with integrated removal of genomic DNA contamination. Real-time PCR was performed using the QuantiTect[®] SYBR[®] Green PCR (Qiagen, Les Ulis, France). Primers 5'-ATCCAGAAGATCACTGCAAG-3' and 5'-TGGTGTAATTGGTTCTCTTAGTAAT-3' were designed to quantify the *WUS* expression. Normalization was done using the *GLYCERALDEHYDE-3-P DEHYDROGENASE (GAPDH)* gene using the primers 5'-ACAAA CATGGGAGCATCCTTACTAG-3' and 5'-GTTTTTACCGACAAGGACAAAGCT-3'. Reverse transcription was performed at 50 °C for 30 min, followed by PCR activation at 90 °C for 15 min and then 25 cycles of PCR amplification (94 °C for 1 min, 52 °C for 1 min and 72 °C for 1 min). Transcript abundance was estimated using the comparative threshold cycle (Ct) method and was calculated as $2^{-\Delta\Delta Ct}$, where $\Delta\Delta Ct = (Ct_{\text{target}} - Ct_{\text{GAPDH}})_{\text{Time x}} - (Ct_{\text{target}} - Ct_{\text{GAPDH}})_{\text{Calibrator}}$.

4.10. Histology and Microscopy

For embryo observation, samples of embryogenic calli were fixed in a mixture of 3.5% (*w/v*) of glutaraldehyde, 0.2 M of cacodylic acid and 2.6% (*w/v*) of sucrose, and then dehydrated in an ethanol series (15%, 30%, 50% for 5 min each and 70%, 90%, 100% for 2 h each). The fixed tissues samples were embedded in Technovit 7100 (Heraeus Kulzer). Histological slices (3 µm) were obtained by using a microtome Jung RM 2045 Leica. Sections were treated with 0.5% (*w/v*) periodic acid and stained with Schiff reagent (20 min; colors polysaccharides in purple) and Naphthol Blue Black (1 min, colors soluble proteins in blue).

4.11. Statistical Analysis

All the experiments were performed by using a completely randomized design (CRD) with three replicates per treatment. In each treatment, 60 explants of leaves and radicles were used. Percentage of callogenesis [(number of explants producing callus per total number of explants), percentage of embryogenic calli [(number of embryogenic calli per number of calli] and mean number of somatic embryos produced per callus (total number of somatic embryos per number of embryogenic calli) were determined. Mean numbers were calculated with their respective standard errors. Data of these observations were analyzed by using standard analysis of variance (ANOVA). The significant difference among treatments was determined using Duncan's multiple range test at $p \leq 0.05$.

5. Conclusions

The efficiency of in vitro embryogenesis in plants is modulated by many central regulators of regeneration and enhanced by exogenously supplied plant growth regulators. Several key regulators are induced during somatic embryogenesis and control downstream physiological responses to promote callogenesis and embryo production. Our work showed that the expression of the *WUS* gene in *Medicago* explants induced callogenesis and the production of highly embryogenic calli. *WUS*-expressing leaf explants produced embryos and generated plantlets in the absence of growth regulators in the media. This confirmed that the overexpression of the *WUS* gene can be useful for improving tissue culture-based regeneration systems and transformation frequencies of recalcitrant species. Further elucidation of the exact roles of the *WUS* stem cell signaling pathway and of related regulator networks is crucial to understand the diverse strategies of somatic embryogenesis.

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References

- Gulzar, B.; Mujib, A.; Malik, M.Q.; Sayeed, R.; Mamgain, J.; Ejaz, B. Genes, proteins and other networks regulating somatic embryogenesis in plants. *J. Genet. Eng. Biotechnol.* **2020**, *18*, 31. [[CrossRef](#)] [[PubMed](#)]
- Méndez-Hernández, H.A.; Ledezma-Rodríguez, M.; Avilez-Montalvo, R.N.; Juárez-Gómez, Y.L.; Skeete, A.; Avilez-Montalvo, J.; De-La-Peña, C.; Loyola-Vargas, V.M. Signaling overview of plant somatic embryogenesis. *Front. Plant Sci.* **2019**, *10*, 1–15. [[CrossRef](#)]
- Nic-Can, G.I.; Loyola-Vargas, V.M. The role of the auxins during somatic embryogenesis. In *Somatic Embryogenesis. Fundamental Aspects and Applications*; Loyola-Vargas, V.M., Ochoa-Alejo, N., Eds.; Springer: Cham, Switzerland, 2016; pp. 171–181. [[CrossRef](#)]
- Stone, S.L.; Kwong, L.W.; Yee, K.M.; Pelletier, J.; Lepiniec, L.; Fisher, R.L.; Goldberg, R.B.; Harada, J.J. *LEAFY COTYLEDON2* encodes a B3 domain transcription factor that induces embryo development. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 11806–11811. [[CrossRef](#)]
- Hecht, V.; Vielle Calzada, J.P.; Hartog, M.V.; Schmidt, E.D.; Boutilier, K.; Grossniklaus, U.; de Vries, S.C. The Arabidopsis *SOMATIC EMBRYOGENESIS RECEPTOR KINASE 1* gene is expressed in developing ovules and embryos and enhances embryogenic competence in culture. *Plant Physiol.* **2001**, *127*, 803–816. [[CrossRef](#)]
- Boutilier, K.; Offringa, R.; Sharma, V.K.; Kieft, H.; Ouellet, T.; Zhang, L.; Hattori, J.; Liu, C.M.; van Lammeren, A.A.; Miki, B.L.; et al. Ectopic expression of *BABY BOOM* triggers a conversion from vegetative to embryonic growth. *Plant Cell* **2002**, *14*, 1737–1749. [[CrossRef](#)] [[PubMed](#)]
- Laux, T.; Mayer, K.F.; Berger, J.; Jürgens, G. The *WUSCHEL* gene is required for shoot and floral meristem integrity in *Arabidopsis*. *Development* **1996**, *122*, 87–96.
- Lotan, T.; Ohto, M.; Yee, K.M.; West, M.A.; Lo, R.; Kwong, R.W.; Yamagishi, K.; Fischer, R.L.; Goldberg, R.B.; Harada, J.J. *Arabidopsis LEAFY COTYLEDON1* is sufficient to induce embryo development in vegetative cells. *Cell* **1998**, *93*, 1195–1205. [[CrossRef](#)]
- Wójcikowska, B.; Jaskóła, K.; Gąsiorek, P.; Meus, M.; Nowak, K.; Gaj, M.D. *LEAFY COTYLEDON2 (LEC2)* promotes embryogenic induction in somatic tissues of *Arabidopsis*, via YUCCA-mediated auxin biosynthesis. *Planta* **2013**, *238*, 425–440. [[CrossRef](#)]
- Zuo, J.; Niu, Q.W.; Frugis, G.; Chua, N.H. The *WUSCHEL* gene promotes vegetative-to-embryonic transition in *Arabidopsis*. *Plant J.* **2002**, *30*, 349–359. [[CrossRef](#)] [[PubMed](#)]
- Wang, X.; Niu, Q.W.; Teng, C.; Li, C.; Mu, J.; Chua, N.H.; Zuo, J. Overexpression of *PGA37/MYB118* and *MYB115* promotes vegetative-to-embryonic transition in *Arabidopsis*. *Cell Res.* **2009**, *19*, 224–235. [[CrossRef](#)]
- Tsuwamoto, R.; Yokoi, S.; Takahata, Y. Arabidopsis *EMBRYOMAKER* encoding an AP2 domain transcription factor plays a key role in developmental change from vegetative to embryonic phase. *Plant Mol. Biol.* **2010**, *73*, 481–492. [[CrossRef](#)]
- Srinivasan, C.; Liu, Z.; Heidmann, I.; Supena, E.N.J.; Fukuoka, H.; Joosen, R.; Lambalk, J.; Angenent, G.; Scorza, R.; Custers, J.B.M.; et al. Heterologous expression of the *BABY BOOM* AP2/ERF transcription factor enhances the regeneration capacity of tobacco (*Nicotiana tabacum* L). *Planta* **2007**, *225*, 341–351. [[CrossRef](#)]
- Heidmann, I.; de Lange, B.; Lambalk, J.; Angenent, G.C.; Boutilier, K. Efficient sweet pepper transformation mediated by the *BABY BOOM* transcription factor. *Plant Cell Rep.* **2011**, *30*, 1107–1115. [[CrossRef](#)]
- Florez, S.L.; Erwin, R.L.; Maximova, S.N.; Guiltinan, M.J.; Curtis, W.R. Enhanced somatic embryogenesis in *Theobroma cacao* using the homologous *BABY BOOM* transcription factor. *BMC Plant Biol.* **2015**, *15*, 121–133. [[CrossRef](#)]

16. Khanday, I.; Santos-Medellin, C.; Sundaresan, V. Rice embryogenic trigger BABY BOOMI promotes somatic embryogenesis by Upregulation of auxin biosynthesis genes. *bioRxiv* **2020**. [[CrossRef](#)]
17. Guo, F.; Liu, C.; Xia, H.; Bi, Y.; Zhao, C.; Zhao, S.; Hou, L.; Li, F.; Wang, X. Induced expression of *AtLEC1* and *AtLEC2* differentially promotes somatic embryogenesis in transgenic tobacco plants. *PLoS ONE* **2013**, *8*, e71714. [[CrossRef](#)]
18. Harding, E.W.; Tang, W.; Nichols, K.W.; Fernandez, D.E.; Perry, S.E. Expression and maintenance of embryogenic potential is enhanced through constitutive expression of *AGAMOUS-Like 15*. *Plant Physiol.* **2003**, *133*, 653–663. [[CrossRef](#)] [[PubMed](#)]
19. Lowe, K.; Wu, E.; Wang, N.; Hoerster, G.; Hastings, C.; Cho, M.J.; Scelonge, C.; Lenderts, B.; Chamberlin, M.; Cushatt, J.; et al. Morphogenic regulators *Baby boom* and *Wuschel* improve monocot transformation. *Plant Cell* **2016**, *28*, 1998–2015. [[CrossRef](#)]
20. Haecker, A.; Gross-Hardt, R.; Geiges, B.; Sarkar, A.; Breuninger, H.; Herrmann, M.; Laux, T. Expression dynamics of WOX genes mark cell fate decisions during early embryonic patterning in *Arabidopsis thaliana*. *Development* **2004**, *131*, 657–668. [[CrossRef](#)] [[PubMed](#)]
21. Tadege, M. WOX3 in the scene: Intimacy with hormones. *J. Exp. Bot.* **2016**, *67*, 1605–1607. [[CrossRef](#)]
22. Ikeda, M.; Takahashi, M.; Fujiwara, S.; Mitsuda, N.; Ohme-Takagi, M. Improving the efficiency of adventitious shoot induction and somatic embryogenesis via modification of *WUSCHEL* and *LEAFY COTYLEDON 1*. *Plants* **2020**, *9*, 1434. [[CrossRef](#)] [[PubMed](#)]
23. Mayer, K.F.; Schoof, H.; Haecker, A.; Lenhard, M.; Jurgens, G.; Laux, T. Role of *WUSCHEL* in regulating stem cell fate in the *Arabidopsis* shoot meristem. *Cell* **1998**, *95*, 805–815. [[CrossRef](#)]
24. Gallois, J.L.; Nora, F.R.; Mizukami, Y.; Sablowski, R. *WUSCHEL* induces shoot stem cell activity and developmental plasticity in the root meristem. *Genes Dev.* **2004**, *18*, 375–380. [[CrossRef](#)]
25. Chen, S.K.; Kurdyukov, S.; Kereszt, A.; Wang, X.D.; Gresshoff, P.M.; Rose, R.J. The association of homeobox gene expression with stem cell formation and morphogenesis in cultured. *M. truncatula*. *Planta* **2009**, *230*, 827–840. [[CrossRef](#)]
26. Zhao, P.; Begcy, K.; Dresselhaus, T.; Sun, M.X. Does early embryogenesis in eudicots and monocots involve the same mechanism and molecular players? *Plant Physiol.* **2017**, *173*, 130–142. [[CrossRef](#)] [[PubMed](#)]
27. Rashid, S.Z.; Yamaji, N.; Kyo, M. Shoot formation from root tip region: A developmental alteration by *WUS* in transgenic tobacco. *Plant Cell Rep.* **2007**, *26*, 1449–1455. [[CrossRef](#)] [[PubMed](#)]
28. Arroyo-Herrera, A.; Ku-Gonzalez, A.; Canche-Moo, R.; Quiroz-Figueroa, F.R.; Loyola-Vargas, V.M.; Rodriguez-Zapata, L.C.; D'Hondt, C.B.; Suárez-Solís, V.M.; Castaño, E. Expression of *WUSCHEL* in *Coffea canephora* causes ectopic morphogenesis and increases somatic embryogenesis. *Plant Cell Tissue Organ. Cult.* **2008**, *94*, 171–180. [[CrossRef](#)]
29. Solís-Ramos, L.Y.; González-Estrada, T.; Nahuath-Dzib, S.; Zapata-Rodríguez, L.C.; Castaño, E. Overexpression of *WUSCHEL* in *C. chinense* causes ectopic morphogenesis. *Plant Cell Tissue Organ. Cult.* **2009**, *96*, 279–287. [[CrossRef](#)]
30. Klimaszewska, K.; Pelletier, G.; Overton, C.; Stewart, D.; Rutledge, R.G. Hormonally regulated overexpression of *Arabidopsis WUS* and conifer *LEC1 (CHAP3A)* in transgenic white spruce: Implications for somatic embryo development and somatic seedling growth. *Plant Cell Rep.* **2010**, *29*, 723–734. [[CrossRef](#)]
31. Bouchabke-Coussa, O.; Obellianne, M.; Linderme, D.; Montes, E.; Maia-Grondard, A.; Vilaine, F.; Pannetier, C. *Wuschel* overexpression promotes somatic embryogenesis and induces organogenesis in cotton (*Gossypium hirsutum* L.) tissues cultured in vitro. *Plant Cell Rep.* **2013**, *32*, 675–686. [[CrossRef](#)]
32. Tvorogova, V.E.; Fedorova, Y.E.; Potsenkovskaya, E.A.; Kudriashov, A.A.; Efremova, E.P.; Kvitkovskaya, V.A.; Wolabu, T.W.; Zhang, F.; Tadege, M.; Lutova, L.A. The *WUSCHEL*-related homeobox transcription factor MtWOX9-1 stimulates somatic embryogenesis in *Medicago truncatula*. *Plant Cell Tissue Organ Cult.* **2019**, *3*, 517–527. [[CrossRef](#)]
33. Zheng, W.; Zhang, X.; Yang, Z.; Wu, J.; Li, F.; Duan, L.; Liu, C.; Lu, L.; Zhang, C.; Li, F. *AtWuschel* Promotes Formation of the Embryogenic Callus in *Gossypium hirsutum*. *PLoS ONE* **2014**, *9*, e87502. [[CrossRef](#)]
34. Cosson, V.; Eschstruth, A.; Ratet, P. *Medicago truncatula* transformation using leaf explants. *Methods Mol. Biol.* **2015**, *1223*, 43–56. [[CrossRef](#)] [[PubMed](#)]
35. Crane, C.; Dixon, R.A.; Wang, Z.Y. *Medicago truncatula* transformation using root explants. *Methods Mol. Biol.* **2006**, *343*, 137–142. [[CrossRef](#)]
36. Rose, R.J. Somatic Embryogenesis in the *Medicago truncatula* Model: Cellular and Molecular Mechanisms. *Front. Plant Sci.* **2019**, *10*, 267. [[CrossRef](#)]
37. Kyo, M.; Maida, K.; Nishioka, Y.; Matsui, K. Coexpression of *WUSCHEL related homeobox (WOX) 2* with or *WOX9* promotes regeneration from leaf segments and free cells in *Nicotiana tabacum* L. *Plant Biotechnol.* **2018**, *35*, 23–30. [[CrossRef](#)]
38. Gaj, M.D. Factors influencing somatic embryogenesis induction and plant regeneration with particular reference to *Arabidopsis thaliana*. (L.) Heynh. *Plant Growth Reg.* **2004**, *43*, 27–47. [[CrossRef](#)]
39. Trinh, T.H.; Ratet, P.; Kondorosi, E.; Durand, P.; Kamate, K.; Bauer, P.; Kondorosi, A. Rapid and efficient transformation of diploid *Medicago truncatula* and *Medicago sativa* ssp. *falcata* lines improved in somatic embryogenesis. *Plant Cell Rep.* **1998**, *17*, 345–355. [[CrossRef](#)] [[PubMed](#)]
40. Häkkinen, S.T.; Moyano, E.; Cusidó, R.M.; Oksman-Caldentey, K.M. Exploring the metabolic stability of engineered hairy roots after 16 years maintenance. *Front. Plant Sci.* **2018**, *7*, 1486. [[CrossRef](#)]
41. Reis, A.; Boutet-Mercey, S.; Massot, S.; Ratet, P.; Zuanazzi, J.A.S. Isoflavone production in hairy root cultures and plantlets of *Trifolium pratense*. *Biotechnol. Lett.* **2019**, *41*, 427–442. [[CrossRef](#)]
42. Gomes, C.; Dupas, A.; Pagano, A.; Grima-Pettenati, J.; Paiva, J.A.P. Hairy Root transformation: A useful tool to explore gene function and expression in *Salix* spp. recalcitrant to transformation. *Front. Plant Sci.* **2019**, *10*, 1427. [[CrossRef](#)] [[PubMed](#)]

43. Zhang, H.; Cao, Y.; Zhang, H.; Xu, Y.; Zhou, C.; Liu, W.; Zhu, R.; Shang, C.; Li, J.; Shen, Z.; et al. Efficient generation of CRISPR/Cas9-mediated homozygous/biallelic *Medicago truncatula* mutants using a hairy root system. *Front. Plant Sci.* **2020**, *11*, 294. [[CrossRef](#)] [[PubMed](#)]
44. Elhiti, M.; Stasolla, C.; Wang, A. Molecular regulation of plant somatic embryogenesis. *Cell. Dev. Biol. Plant.* **2013**, *49*, 631–642. [[CrossRef](#)]
45. Christensen, A.H.; Quail, P.H. Ubiquitin promoter-based vectors for high-level expression of selectable and/or screenable marker genes in monocotyledonous plants. *Transgenic Res.* **1996**, *5*, 213–218. [[CrossRef](#)]
46. Pagny, S.; Cabanes-Macheteau, M.; Gillikin, J.W.; Leborgne-Castel, N.; Lerouge, P.; Boston, R.; Faye, L.; Gomord, V. Protein recycling from the Golgi apparatus to the endoplasmic reticulum is very active in plants but has a minor contribution to calreticulin retention. *Plant Cell* **2000**, *12*, 739–755. [[CrossRef](#)]
47. Guerineau, F.; Benjdia, M.; Zhou, D.X. A jasmonate-responsive element within the *A. thaliana vsp1* promoter. *J. Exp. Bot.* **2003**, *54*, 1153–1162. [[CrossRef](#)] [[PubMed](#)]
48. Guerineau, F.; Woolston, S.; Brooks, L.; Mullineaux, P. An expression cassette for targeting foreign proteins into chloroplasts. *Nucleic Acids Res.* **1988**, *16*, 11380. [[CrossRef](#)]
49. Sambrook, J.; Fritschi, E.F.; Maniatis, T. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: New York, NY, USA, 1989; p. 545.
50. Hanahan, D.; Jessee, J.; Bloom, F.R. Plasmid transformation of *Escherichia coli* and other bacteria. *Methods Enzymol.* **1991**, *20*, 63–113.
51. Vervliet, G.; Holsters, M.; Teuchy, H.; Van Montagu, M.; Schell, J. Characterization of different plaque forming and defective temperate phages in *Agrobacterium* strains. *J. Gen. Virol.* **1975**, *26*, 33–48. [[CrossRef](#)]
52. Schenk, R.V.; Hildebrandt, A.C. Medium and Techniques for Induction and Growth of Monocotyledonous and Dicotyledonous Plant Cell Cultures. *Can. J. Bot.* **1972**, *50*, 199–204. [[CrossRef](#)]
53. Hoffmann, B.; Trinh, T.H.; Leung, J.; Kondorosi, A.; Kondorosi, E. A new *Medicago truncatula* line with superior in vitro regeneration, transformation, and symbiotic properties isolated through cell culture selection. *Mol. Plant Microbe Interact.* **1997**, *10*, 307–315. [[CrossRef](#)]
54. Boisson-Dernier, A.; Chabaud, M.; Garcia, F.; Bécard, G.; Rosenberg, C.; Barker, D.G. Hairy roots of *Medicago truncatula* as tools for studying nitrogen-fixing and endomycorrhizal symbioses. *Mol. Plant Microbe Interact.* **2001**, *14*, 693–700. [[CrossRef](#)]
55. Fahraeus, G. The infection of clover root hairs by nodule bacteria studied by a simple glass slide technique. *J. Gen. Microbiol.* **1957**, *16*, 374–381. [[CrossRef](#)] [[PubMed](#)]
56. Murashige, T.; Skoog, F. A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiol. Plant.* **1962**, *15*, 473–497. [[CrossRef](#)]