



HAL
open science

Host dispersal shapes the population structure of a tick-borne bacterial pathogen

Ana Cláudia Norte, Gabriele Margos, Noémie Becker, Jaime Albino Ramos, Maria Sofia Nuncio, Volker Fingerle, Pedro Miguel Araújo, Peter Adamík, Haralambos Alivizatos, Emilio Barba, et al.

► **To cite this version:**

Ana Cláudia Norte, Gabriele Margos, Noémie Becker, Jaime Albino Ramos, Maria Sofia Nuncio, et al.. Host dispersal shapes the population structure of a tick-borne bacterial pathogen. *Molecular Ecology*, 2020, 29 (3), pp.485-501. 10.1111/mec.15336 . hal-02990550

HAL Id: hal-02990550

<https://cnrs.hal.science/hal-02990550>

Submitted on 19 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

MOLECULAR ECOLOGY

Host dispersal shapes the population structure of a tick-borne bacterial pathogen

Journal:	<i>Molecular Ecology</i>
Manuscript ID	MEC-18-1428.R1
Manuscript Type:	Original Article
Date Submitted by the Author:	n/a
Complete List of Authors:	<p>Norte, Ana; University of Coimbra, MARE - Marine and Environmental Sciences Centre, Department of Life Sciences Margos, Gabriele; Bavarian Health and Food Safety Authority, German National Reference Centre for <i>Borrelia</i> Becker, Noemie; LMU Munich, Division of Evolutionary Biology, Faculty of Biology Ramos, Jaime; University of Coimbra, MARE - Marine and Environmental Sciences Centre, Department of Life Sciences Nuncio, Maria ; National Institute of Health Dr. Ricardo Jorge, Infectious Department Fingerle, Volker; Bavarian Health and Food Safety Authority, German National Reference Centre for <i>Borrelia</i> Araújo, Pedro; MARE – Marine and Environmental Sciences Centre, Dep. Life Sciences Adamik, Peter; Palacky University, Department of Zoology Alivizatos, Haralambos; Hellenic Bird Ringing Center Barba, Emilio; University of Valencia, Cavanilles' Institute of Biodiversity and Evolutionary Biology Barrientos, Rafael; Universidad Complutense de Madrid Cauchard, Laure; Université de Montréal, Département des Sciences Biologiques Csorgo, Tibor; Eötvös Loránd University, Budapest, Department of Anatomy, Cell and Developmental Biology Diakou, Anastasia; Aristotle University of Thessaloniki Faculty of Health Sciences Dingemanse, Niels; Behavioural Ecology, Department of Biology, Ludwig Maximilians University of Munich Doligez, Blandine; CNRS - Department of Biometry and Evolutionary Biology (LBBE) - University Lyon 1 Dubiec, Anna; Polska Akademia Nauk Muzeum i Instytut Zoologii Eeva, Tapio; University of Turku, Department of Biology, Section of Ecology Flaisz, Barbara; University of Veterinary Medicine Grim, Tomas; Palacky University Hau, Michaela; Max Planck Institute for Ornithology, Evolutionary Physiology Heylen, Dieter; University of Antwerp Hornok, Sandor; Department of Parasitology and Zoology, University of Veterinary Medicine Kazantzidis, Savas; Forest Research Institute</p>

	<p>Kovats, David; Hungarian Biodiversity Research Society Krause, František Literak, Ivan; Department of Biology and Wildlife Diseases, Faculty of Veterinary Hygiene and Ecology, University of Veterinary and Pharmaceutical Sciences Brno Mand, Raivo; Department of Zoology, University of Tartu Mantesana, Lucia; Max-Planck-Institute for Ornithology Morinay, Jennifer; University Lyon Mutanen, Marko; Department of Ecology and Genetics, University of Oulu Neto, Julio; Molecular Ecology and Evolution Lab, University of Lund Nováková, Markéta; University of Veterinary and Pharmaceutical Sciences Brno Sanz, Juan José; Museo Nacional de Ciencias Naturales (CSIC), Ecología Evolutiva da Silva, Luis; CIBIO, Centro de Investigação em Biodiversidade e Recursos Genéticos, InBIO Laboratório Associado, Universidade do Porto; Center for Functional Ecology - Science for People & the Planet, Department of Life Sciences, University of Coimbra Sprong, Hein; National Institute for Public Health and the Environment (RIVM), Centre for Infectious Disease Control Tirri, Ina; University of Helsinki Török, János; Eötvös Loránd University Trilar, Tomi ; Slovenian Museum of Natural History Tyller, Zdeněk; Palacky University; Museum of the Moravian Wallachia region Visser, Marcel; Department of Animal Ecology, Netherlands Institute of Ecology (NIOO-KNAW) Lopes Carvalho, Isabel; National Institute of Health Dr. Ricardo Jorge, Infectious Department</p>
Keywords:	Birds, ticks, host-parasite interactions, migration, <i>Borrelia garinii</i> , Lyme borreliosis

Host dispersal shapes the population structure of a tick-borne bacterial pathogen

Running title: Population structure of a tick-borne pathogen

Authors:

Ana Cláudia Norte^{1,2}, Gabriele Margos³, Noémie S. Becker⁴, Jaime Albino Ramos¹, Maria Sofia Nuncio², Volker Fingerle³, Pedro Miguel Araújo¹, Peter Adamík⁵, Haralambos Alivizatos⁶, Emilio Barba⁷, Rafael Barrientos⁸, Laure Cauchard⁹, Tibor Csörgő^{10,11}, Anastasia Diakou¹², Niels J. Dingemans¹³, Blandine Doligez¹⁴, Anna Dubiec¹⁵, Tapio Eeva¹⁶, Barbara Flaisz¹⁷, Tomas Grim⁵, Michaela Hau¹⁸, Dieter Heylen¹⁹, Sándor Hornok¹⁷, Savas Kazantzidis²⁰, David Kováts^{10,21}, František Krause²², Ivan Literak²³, Raivo Mänd²⁴, Lucia Montesana¹⁸, Jennifer Morinay^{14,25}, Marko Mutanen²⁶, Júlio Manuel Neto²⁷, Markéta Nováková²³, Juan José Sanz²⁸, Luís Pascoal da Silva^{29,30}, Hein Sprong³¹, Ina-Sabrina Tirri³², János Török³³, Tomi Trilar³⁴, Zdeněk Tyller^{5,35}, Marcel E. Visser³⁶ and Isabel Lopes de Carvalho²

Affiliations:

1. MARE – Marine and Environmental Sciences Centre, Dep. Life Sciences, University of Coimbra, Coimbra, Portugal

2. Centre for Vector and Infectious Diseases Research, Instituto Nacional de Saúde Dr. Ricardo Jorge, Lisbon, Portugal

3. German National Reference Centre for *Borrelia* (NRZ), Bavarian Health and Food Safety Authority (LGL), Oberschleissheim, Germany
4. Division of Evolutionary Biology, Faculty of Biology, LMU Munich, Planegg-Martinsried, Germany
5. Department of Zoology, Palacky University, Olomouc, Czech Republic
6. Hellenic Bird Ringing Center, Athens, Greece
7. Instituto Cavanilles de Biodiversidad y Biología Evolutiva (ICBiBE), Universidad de Valencia, Spain
8. Department of Biodiversity, Ecology and Evolution, Universidad Complutense de Madrid, Madrid, Spain
9. Département des Sciences Biologiques, Université de Montréal, Montréal, Canada
10. Ócsa Bird Ringing Station, Ócsa, Hungary
11. Department of Anatomy, Cell and Developmental Biology, Eötvös Loránd University, Budapest, Hungary
12. Laboratory of Parasitology and Parasitic Diseases School of Veterinary Medicine Faculty of Health Sciences Aristotle University of Thessaloniki, Thessaloniki, Greece
13. Behavioural Ecology, Department of Biology, Ludwig Maximilians University of Munich, Planegg-Martinsried, Germany
14. CNRS - Department of Biometry and Evolutionary Biology (LBBE) - University Lyon 1, University of Lyon, Villeurbanne, France

15. Museum and Institute of Zoology, Polish Academy of Sciences, Warszawa, Poland
16. Department of Biology, University of Turku, Finland
17. Department of Parasitology and Zoology, University of Veterinary Medicine, Budapest, Hungary
18. Evolutionary Physiology Lab, Max Planck Institute for Ornithology, Seewiesen, Bavaria, Germany
19. Evolutionary Ecology Group, Department of Biology, University of Antwerp, Antwerp, Belgium
20. Forest Research Institute, Hellenic Agricultural Organization "DEMETER", Vassilika, Thessaloniki, Greece
21. Hungarian Biodiversity Research Society, Hungary
22. Břetislavova 8, 690 02 Břeclav, Czech Republic
23. Department of Biology and Wildlife Diseases, Faculty of Veterinary Hygiene and Ecology, University of Veterinary and Pharmaceutical Sciences Brno, Brno, Czech Republic
24. Department of Zoology, University of Tartu, Tartu, Estonia
25. Department of Ecology and Evolution, Animal Ecology, Evolutionary Biology Centre, Uppsala University, Uppsala, Sweden
26. Department of Ecology and Genetics, University of Oulu, Oulu, Finland

27. Molecular Ecology and Evolution Lab, Department of Biology, University of Lund, Ecology Building, Lund, Sweden
28. Departamento de Ecología Evolutiva, Museo Nacional de Ciencias Naturales (CSIC), Madrid, Spain
29. CFE - Centre for Functional Ecology - Science for People & the Planet, Department of Life Sciences, University of Coimbra, Coimbra, Portugal
30. CIBIO-InBIO, Research Center in Biodiversity and Genetic Resources, University of Porto, Portugal
31. National Institute of Public Health and Environment (RIVM), Laboratory for Zoonoses and Environmental Microbiology, Bilthoven, Netherlands
32. Finnish Museum of Natural History, University of Helsinki, Finland
33. Behavioural Ecology Group, Department of Systematic Zoology and Ecology, Eötvös Loránd University, Budapest, Hungary
34. Slovenian Museum of Natural History, Ljubljana, Slovenia
35. Museum of the Moravian Wallachia region, Vsetín, Czech Republic
36. Department of Animal Ecology, Netherlands Institute of Ecology (NIOO-KNAW), Wageningen, The Netherlands

Corresponding author:

Isabel Lopes de Carvalho

National Institute of Health, Portugal

Av. Padre Cruz, 1649-016 Lisboa

Email: isabel.carvalho@insa.min-saude.pt

1 **Abstract**

2 **Keywords:** birds; ticks; host-parasite interactions; migration; *Borrelia garinii*; Lyme

3 borreliosis

4

5 1. INTRODUCTION

6

7 Wild birds are relevant for public health because of their role in the spread of emerging
8 zoonotic pathogens that cause newly recognised diseases or diseases which are rapidly
9 increasing in incidence or geographical range (Reed, Meece, Henkel, & Shukla, 2003). Some
10 birds act as reservoirs of pathogens such as *Borrelia burgdorferi* sensu lato (s.l.),
11 enterobacteria, flavivirus and influenza A virus, being significantly involved in the direct
12 infection of humans or arthropod vectors that transmit the disease agents to humans (Thomas,
13 Hunter, & Atkinson, 2007). Wild birds, especially migratory species, may also carry the
14 arthropod vectors (e.g. ticks) to different geographic areas creating new foci of disease (Reed
15 et al., 2003). Studies that monitored tick infestation of birds during migration estimated that
16 birds are responsible for the transport of 6.8 - 175 million ticks each spring between wintering
17 and breeding areas (Ogden et al., 2008; Olsen, Jaenson, & Bergstrom, 1995), which may
18 greatly impact the distribution and population structure of ticks and their associated
19 pathogens. The life cycle of tick-borne pathogens is complex and their evolutionary ecology
20 is shaped by the interactions with vertebrate hosts and tick vectors (Kurtenbach et al., 2006).
21 This study focused on the ecology and genetic diversity of *Borrelia burgdorferi* s.l. as a
22 model to investigate the drivers of the population structure and to understand the role of host-
23 associated dispersal on the evolution of tick-borne pathogens. This represents a consequential
24 question in the ecology and evolution of any pathogen.

25 *Borrelia burgdorferi* s.l. is a bacterial complex of over 20 known genospecies,
26 including the etiologic agents of Lyme borreliosis (Casjens et al., 2011; Margos et al., 2015),
27 whose main vectors are ticks of the genus *Ixodes* (Eisen & Lane, 2002). These bacteria are
28 widespread in Europe, Asia and North America and are also present in North Africa (Margos,

29 Vollmer, Ogden, & Fish, 2011; Zhioua et al., 1999). Different *Borrelia* genospecies have
30 different patterns of association with vertebrate reservoir hosts (Humair & Gern, 2000;
31 Kurtenbach, Peacey, et al., 1998) because of the immunological host response, mediated by
32 the action of the host's complement system (Kurtenbach et al., 2002). While *B. burgdorferi*
33 sensu stricto (s.s.) is a generalist genospecies, *B. afzelii* is mostly associated with mammalian
34 hosts such as rodents, whereas *B. valaisiana*, *B. garinii* and *B. turdi* are mostly associated
35 with birds (Heylen, 2016; Margos et al., 2011). Because tick vectors cannot move large
36 distances independent of hosts, it has been suggested that host specialization determines the
37 spread and dispersal of *B. burgdorferi* s.l. genospecies (Kurtenbach et al., 2010; Sonenshine
38 & Mather, 1994). Because birds are both important hosts for some *Borrelia* genospecies and
39 for various species of vector ticks, they act as a driving force shaping *B. burgdorferi* s.l.
40 distribution and phylogeographical patterns (Margos et al., 2011; Vollmer et al., 2011).

41 Here, we assessed the role of passerine birds as hosts and dispersers of *B. burgdorferi*
42 s.l. We tested the hypothesis that infection prevalence with *Borrelia* genospecies would differ
43 among bird species due to differences in their ecological niche occupancy and reservoir
44 competence for *B. burgdorferi* s.l. We also evaluated whether the avian-associated and human
45 pathogenic genospecies *B. garinii* would show lack of geographical structuring, because of
46 the large distance range of movements of its avian hosts and potential for dispersal and
47 consequent spatial mixing of strains. To achieve this, we collected ticks feeding on common
48 passerine species with different ranges of migratory movements in a coordinated study
49 covering a broad geographical area in Europe (11 countries), and assessed the infection status
50 of these ticks with *B. burgdorferi* s.l. The diversity of the common avian-associated *B. garinii*
51 genospecies and potential phylogeographical patterns were determined using a multilocus
52 sequence typing scheme (MLST) of eight house-keeping genes (Margos et al., 2008).

53 Analyses based on these conserved genetic markers have been previously used to estimate
54 and describe the degree of population genetic structure over the largest geographical range
55 studied so far, and have the potential to reveal signatures of demographic processes, dispersal
56 and migration (Hoen et al., 2009; Margos et al., 2012; S. Vollmer et al., 2013).

57

58 **2. METHODS**

59

60 **2.1 Birds and ticks**

61

62 Birds were captured in collaboration with ornithologists and ringers in 11 European countries
63 (Czech Republic, Estonia, Finland, Germany, Greece, Hungary, Netherlands, Portugal,
64 Slovenia, Spain and Sweden) covering an area from 8°23'W to 24°57'E and from 40°35'N to
65 62°14'N. Capturing effort directed to tick collection was concentrated during the year of 2015,
66 but additional data (including data from the years 2005-2008, 2013-2014 and 2016) was
67 included for 5 of the 21 study sites (Sup. Mat 1). These collection sites correspond to ringing
68 stations or sites where bird populations of particular species have been studied in a long-term
69 perspective and with which we could establish a collaborative protocol. Therefore, some
70 European areas are missing from this study due to sampling limitations. Birds were captured
71 using mist-nets or, when breeding in nest boxes: incubating females were captured by hand
72 (in Estonia, in Gotland, Sweden and in Harjavalta, Finland), and both parents were caught
73 when feeding the nestlings using spring or wire traps. Nestlings were sampled in the nest
74 between 8 and 15 days of age. Countries, collection sites, range of capturing dates and bird
75 species sampled are detailed in Fig.1 and Sup. Mat. 1. Although classification of collection

76 sites according to country has no biological meaning, for convenience, and because countries
77 are related to geographical positioning, we refer to country of collection when reporting some
78 results for an easy identification of sample origin, for comparability with previous localised
79 studies in different countries, and for an integration with sequence data available in public
80 databases. Birds were carefully inspected for attached ticks with special attention to the head
81 (around the eyes, beak, ears, chin and crown) and neck, where ticks are most often attached.
82 We removed infesting ticks with fine forceps and collected them into tubes containing 70 -
83 99% ethanol according to each individual host. Because we were interested in *B. burgdorferi*
84 s.l. infection prevalence in ticks feeding on birds, our analyses use ticks as sampling units.
85 Therefore, we did not collect data on non-infested birds nor on tick infestation intensity. Our
86 statistical analyses take into account sampling site, bird life cycle stage and month, to account
87 for differences in sampling effort and uneven sample distribution across sites and time of
88 year. The ticks were identified morphologically using identification keys (Estrada-Peña,
89 Bouattour, Camicas, & Walker, 2004; Estrada-Peña, Nava, & Petney, 2014; Pérez-Eid, 2007).

90

91 **2.2 Molecular analysis**

92

93 We extracted tick DNA in a subset of *Ixodes* spp. ticks ($n = 656$ ticks; mean ticks \pm SE = 65.6
94 ± 11.5 per country, 38.6 ± 14.28 per bird species, 1.54 ± 0.03 per bird), using a column DNA
95 extraction kit (DNeasy, Qiagen, Hilden, Germany). Tick (nymph and adult) exoskeleton was
96 broken by piercing followed by incubation with proteinase K, for 24h. We tested a sub-
97 sample of these *Ixodes* spp. ($n = 58$, mean ticks \pm SE = 5.8 ± 0.79 randomly selected per
98 country) using a conventional PCR targeting the mitochondrial 16S rRNA gene of ticks using
99 the primers described by (Mangold, Bargues, & Mas-Coma, 1998) and an annealing

100 temperature of 56°C, to confirm morphological identification of ticks by BLASTn search
101 (<https://blast.ncbi.nlm.nih.gov/>). For specimens (n = 4) for which the % of identity in
102 BLASTn search was less than 98% we built a Maximum Likelihood phylogenetic tree,
103 together with reference sequences (Chitimia-Dobler et al., 2018; Estrada-Peña et al., 2014)
104 retrieved from Genbank, and confirmed the species identification with the obtained clustering
105 patterns. We assessed infection of ticks by a nested PCR targeting the *flaB* gene of *B.*
106 *burgdorferi* s.l. using the primers described in (Johnson, Happ, Mayer, & Piesman, 1992),
107 with an annealing temperature of 52°C. We used the Invitrogen PCR Reagent System mix
108 (Life Technologies, Waltham, USA), according to manufacturer's instructions, and a positive
109 and a negative control were used in all PCR runs. *Borrelia* genospecies were identified by
110 sequencing. The procedures listed above were performed at the Portuguese National Institute
111 of Health Doutor Ricardo Jorge, Portugal.

112 A sub-sample of *B. garinii*-positive specimens were tested using MLST targeting eight
113 housekeeping genes (*clpA*, *clpX*, *nifS*, *pepX*, *pyrG*, *recG*, *rplB*, *uvrA*) according to Margos et
114 al. (2008) at the German National Reference Centre for *Borrelia*, Bavarian Health and Food
115 Safety Authority, Germany. Those isolates were selected from common bird species (*Turdus*
116 spp. and hole-nesting birds) that covered the geographical range in which *B. garinii* was
117 detected in our study. All samples were analysed following the same protocol (see
118 <https://pubmlst.org/borrelia/sequencing> for protocol and primer information). We compared
119 the obtained sequences with those available in the *Borrelia* MLST database
120 (<https://pubmlst.org/borrelia/>) located at the University of Oxford, to obtain allele and
121 sequence type (ST) numbers for each isolate (Jolley & Maiden, 2010). Novel alleles or STs
122 were given novel numbers. Samples that contained mixed infections were discarded from

123 further MLST/MLSA analyses because allele numbers and sequence types could not be
124 identified for these samples. We submitted all data to the *Borrelia* MLST database.

125 A novel *Borrelia* genospecies was detected and characterised by PCR amplification of
126 the 5S-23S rRNA intergenic spacer (Rijpkema, Molkenboer, Schouls, Jongejan, &
127 Schellekens, 1995), and subsequent sequencing of PCR amplicons of the 16S rRNA locus
128 (Radulović, Milutinovic, Tomanovic, & Mulenga, 2010) and of the *clpX* gene (Margos et al.,
129 2008).

130

131 **2.3 Statistical analyses**

132

133 Factors affecting the prevalence of *B. burgdorferi* s.l. in ticks from birds (each tick feeding on
134 a bird was used as a sampling unit) were tested using a Generalized Linear Mixed Model
135 (GLMM) with a binomial error distribution (logit function). The full model included bird
136 species (10 levels), latitude, longitude, tick stage (larvae or other), tick species (4 levels), bird
137 life cycle stage (breeding or non-breeding) and month (12 levels) as fixed effects. Bird
138 identity (bird_ID, to control for ticks tested from the same individual), nested within
139 collection site, was included as a random effect. We ran a set of models with different fixed
140 effects structures and used the Akaike Information Criterion (AIC) to select the best model
141 (best model selection table is presented in Sup. Mat 2). After ranking the models using AIC,
142 we used those with a $\Delta\text{AIC} < 2$ with respect to the top model to calculate conditional model-
143 averaged parameter estimates. Significance level was defined at $P = 0.05$.

144 In these models, only bird species for which we had information on *Borrelia* infection
145 for at least eight ticks (from different individuals) were used. Tick stage was considered in

146 statistical analyses and was divided into larvae and other tick stages because nymphs
147 represented the majority whilst adults were too infrequently found to justify their own group
148 for statistical analysis. This procedure was adopted because transovarial transmission (i.e. the
149 acquisition of *Borrelia* by larvae via vertical transmission from the parent) is considered to
150 be very low in *Ixodes* spp. (Eisen & Lane, 2002). For this reason, an infection of a larva is
151 highly unlikely if they were collected from an uninfected or incompetent reservoir bird host.
152 On the other hand, nymphs/adults may have acquired an infection during a previous blood
153 meal on an infected host. In such case *Borrelia* DNA may be detected in the tick independent
154 of the bird hosts they were collected from. Tick species was also included in the models
155 because within the genus *Ixodes*, different species differ in their vector competence for *B.*
156 *burgdorferi* s.l. (Eisen & Lane, 2002; Heylen, Krawczyk, et al., 2017; Heylen, Sprong, et al.,
157 2014). We controlled for the effects of timing of tick collection throughout the year including
158 month of collection as explanatory variable and also by grouping those ticks collected during
159 the birds' breeding season (April – July) and those collected outside the breeding season
160 (August – March), because breeding is one of the most stressful periods in the birds' life cycle
161 and the stress associated with breeding duties may suppress their immune system potentially
162 leading to spirochetemia. Therefore, this could affect the probability of infection of the ticks
163 feeding on the birds. We used the same statistical approach to test factors affecting prevalence
164 of infection by the most prevalent *Borrelia* genospecies in our study (*B. garinii*). The models
165 were run in R (R core team, 2013) using the packages lme4, lmerTest, bbmle, MuMin and
166 arm (Bates, Mächler, Bolker, & Walker, 2015; Kuznetsova, Brockhoff, & Christensen, 2017).

167

168 **2.4. Multilocus Sequence Typing and Multilocus Sequence Analysis (MLST/MLSA)**

169

170 In order to study the population structure and the phylogenetic relationships of *B. garinii* in a
171 global context, we selected a subset of 82 *B. garinii*- positive samples, as to include isolates
172 from all European countries sampled in this study and represent all bird species sampled with
173 *B. garinii*- infected ticks ($n \geq 5$ infected ticks), and tested those by MLST.

174 Complete allelic profiles of *B. garinii*- positive specimens obtained in our study were
175 analysed with goeBURST analysis using PhyloViz (Francisco, Bugalho, Ramirez, & Carriço,
176 2009) together with other *B. garinii* genotypes detected worldwide (identification of the
177 isolates used for MLST/MLSA is given in Sup. Mat. 3; we included each ST only once for
178 each country). Relationships among STs were evaluated through triple locus variants (TLV),
179 and founder clonal complexes were identified to infer patterns of descent.

180 We estimated nucleotide diversity (π ; Nei, 1987) and Tajima D (Tajima, 1989) for
181 each gene using R packages pegas v. 0.9 (Paradis, 2010) and ape v. 3.5 (Paradis, Claude, &
182 Strimmer, 2004) on each continent for a sample set including 304 isolate sequences (198 from
183 Europe, 85 from Asia and 21 from North America), and on each country ($n = 11$) for which
184 more than five isolates were available (see Sup. Mat. 3 for identification of the isolates
185 included in this analysis). The sequences for gene *clpX* were realigned using MAFFT v7.205
186 (Kato & Standley, 2013) as there was a deletion of three bases in some isolates.

187 An ancestry recombination graph for the 85 STs present in Europe was reconstructed with
188 BEAST2 software v. 2.5 (Bouckaert et al., 2019) and package bacter v. 2.2 (Didelot, Lawson,
189 Darling, & Falush, 2010; Vaughan et al., 2017) using sequences of the eight housekeeping
190 genes. BEAST2 was run three times with a unique tree and substitution model for the eight
191 loci but with a lognormal-relaxed clock model for each locus. We used the following priors:

192 HKY substitution model (Hasegawa, Kishino, & Yano, 1985), Gamma site heterogeneity
193 model with four gamma categories, Tree prior: Coalescent Constant Populations. Due to the
194 high number of STs and the complex model including recombination, the chain was slow to
195 converge (as was shown by Tracer v. 1.6 (Rambaut, Suchard, Xie & Drummond, 2014) and
196 we thus extended the original 10M states to 17M for one run and 19M for the other two. A
197 consensus tree was reconstructed with the tool ACGAnnotator present in the bacter package
198 after removing 40% to 70% burn-in depending on the run.

199
200

201 3. RESULTS

202

203 3.1 Ticks collected from birds

204 In total, 2,308 ticks were collected from 843 infested birds, belonging to 28 bird species (Sup.
205 Mat. 1). Ticks collected from these birds belonged to three genera: *Haemaphysalis* (n = 3),
206 *Hyalomma* (n = 48) and *Ixodes* (n = 2,255). Two ticks could not be identified to genus by
207 morphological criteria as they were damaged. We identified four species of *Ixodes*: *I. ricinus*
208 (n = 1,779), *I. arboricola* (n = 214), *I. frontalis* (n = 164), and *I. ventalloi* (n = 24) but 74
209 *Ixodes* ticks could not be identified to species because they lacked critical body structures
210 needed for morphological identification (Sup. Mat. 4). The vast majority of collected ticks
211 were immatures (2,175 out of 2,255 *Ixodes* spp.), and from these, 63% were nymphs. Adults
212 belonged to *I. arboricola* (n = 63), *I. frontalis* (n = 11), *I. ricinus* (n = 1) and *I. ventalloi* (n =
213 4). Amplification and sequencing of the ribosomal 16S rRNA gene of ticks (Mangold,
214 Barges, & Mas-Coma, 1998) confirmed tick morphological identification in 84% of the
215 cases, corresponding to a misidentification rate of 16%.

216 The blackbird *Turdus merula*, the song thrush *T. philomelos*, the redwing *T. iliacus*,
217 the great tit *Parus major*, the collared-flycatcher *Ficedula albicollis*, and the Eurasian jay
218 *Garrulus glandarius* presented co-infestations by ticks of different species (Sup. Mat. 4).

219

220 3.2 Prevalence of *Borrelia burgdorferi* s.l. in ticks collected from birds

221

222 Out of 656 *Ixodes* ticks collected from birds and analysed for *B. burgdorferi* s.l. infection, 244
223 (37.2%) were positive. Of these, 22 were larvae (prevalence of *B. burgdorferi* s.l. in larvae =
224 20%, 22/110), and 214 were nymphs (prevalence of *B. burgdorferi* s.l. in nymphs = 41%,
225 214/521). *Ixodes ricinus* was the most infected tick species (40.2%, 210/522), followed by *I.*
226 *ventalloi* (31.3%, 5/16), *I. arboricola* (29.7%, 14/47) and *I. frontalis* (20.5%, 9/44).

227 *Borrelia burgdorferi* s.l. prevalence differed significantly between ticks collected from
228 different bird species and was affected by latitude ($\chi_{10,616}^2 = 90.10$, $P < 0.0001$; Fig. 2, Fig. 3).
229 Longitude, tick stage, tick species, month and the birds' life cycle stage did not affect *B.*
230 *burgdorferi* s.l. prevalence. The model selection table is presented in Sup. Mat. 2 and the
231 conditional model averaged coefficients parameters obtained from the generalized linear
232 mixed models (GLMMs) that best explained the prevalence of *B. burgdorferi* s.l. in ticks
233 collected from birds are presented in Table 1. In comparison with the reference species (the
234 blue tit *Cyanistes caeruleus*), ticks collected from *T. merula* (estimate \pm SE = 2.50 ± 0.91 , $z =$
235 2.76 , $P = 0.006$) and *Turdus pilaris* (estimate \pm SE = 4.29 ± 1.51 , $z = 2.84$, $P = 0.005$) showed
236 higher infection rates, whereas those collected from the robin *E. rubecula* had the lowest
237 infection rates (estimate \pm SE = -2.41 ± 1.25 , $z = -1.93$, $P = 0.054$). *B. burgdorferi* s.l.
238 prevalence increased with latitude (estimate \pm SE = 0.08 ± 0.03 , $z = 2.28$, $P = 0.022$). The

239 fieldfare *T. pilaris* was the bird species that carried ticks with the highest *Borrelia* prevalence
240 (92%), followed by the blackbird *T. merula* (58%). Only two out of 53 (3.8%) ticks feeding
241 on the robin *E. rubecula* were positive for *B. burgdorferi* s.l. (Fig. 2).

242 The genospecies of 193 positive samples was identified by sequencing the *flaB* gene.
243 The most prevalent genospecies was *B. garinii* (60.7%, 116/191), followed by *B. valaisiana*
244 (23.6%, 45/191), *B. afzelii* (9.4%, 18/191) and *B. turdi* (5.2%, 10/191). *B. lusitaniae* (0.5%,
245 1/191) and a novel genospecies (0.5%, 1/191) were also detected (Fig. 2 and 3; Sup. Mat. 5).

246 The most abundant genospecies associated with *T. pilaris*, *T. philomelos*, *P. major* and
247 *F. albicollis* was *B. garinii* with a prevalence varying between 100% (13/13) in *T. pilaris* and
248 53% (8/15) in *T. philomelos* (Fig. 2), whereas *B. valaisiana* was the most prevalent
249 genospecies detected in ticks collected from *T. merula* (50%, 30/60; Fig. 2). The model
250 explaining the variation in *B. garinii* prevalence, the most common genospecies detected in
251 ticks feeding on birds, was identical to the one explaining *B. burgdorferi* s.l. prevalence, with
252 the exception that the prevalence of *B. garinii* in ticks feeding on *T. merula* and *E. rubecula*
253 was not significantly different from the reference bird species. Under the assumption that
254 there was no co-feeding transmission (i.e. when larvae acquire the infection due to feeding in
255 close proximity to other infected tick stages; Randolph, Gern, & Nuttall, 1996), data on larval
256 infection suggested that *F. albicollis* and *T. merula* may act as reservoirs for *B. garinii* and *B.*
257 *valaisiana*, *T. iliacus* for *B. garinii*, *B. valaisiana* and *B. turdi*, *P. major* for *B. garinii*, and the
258 willow warbler *P. trochilus* and *E. rubecula* for *B. garinii* (Fig. 2; Sup. Mat. 5).

259 *Borrelia afzelii* DNA was detected only in nymphs, mostly feeding on *P. major*, and
260 to a lesser extent on other bird species (*C. caeruleus*, *F. albicollis*, *T. iliacus* and *T. merula*).
261 *Borrelia lusitaniae* DNA was detected in one *I. ricinus* nymph feeding on *P. major*. *Borrelia*

262 *turdi* DNA was detected in all stages of two tick species, *I. frontalis* (66.7%; 6 out of 9
263 positive ticks), and *I. ventalloi* (80%; 4 out of 5 positive ticks), feeding on *T. iliacus*, *T.*
264 *merula*, *T. philomelos* and *P. major*.

265 DNA of the new *Borrelia* genospecies (*Candidatus* *Borrelia aligera*) was detected in
266 an *I. ventalloi* nymph feeding on a *T. iliacus* in Portugal. Its *flaB* sequence was 100%
267 identical to a *flaB* sequence previously detected in one *I. ricinus* nymph feeding on a
268 Sardinian warbler *Sylvia melanocephala* in Portugal (isolate T794A; accession number
269 KT207789; Norte et al., 2015). The PCR targeting the 5S-23S rRNA intergenic spacer was
270 positive showing that this genospecies belongs to the *B. burgdorferi* s.l. group. Its 16S rRNA
271 sequence showed only 97% similarity to several *B. burgdorferi* genospecies, including *B.*
272 *bissettiae* and *B. mayonii*. The sequence of the housekeeping gene *clpX* showed 36 nucleotide
273 differences from all previously detected alleles available at the MLST database
274 (<http://pubmlst.org/borrelia>). Detailed information on specimens from which different
275 *Borrelia* genospecies were detected in this study is presented in Sup. Mat. 5.

276

277

278 **3.3 Multilocus Sequence Typing / Multilocus Sequence Analysis (MLST/MLSA)**

279

280 Twenty-nine complete allelic profiles with sequences for all eight genes were obtained from a
281 subset of 82 *B. garinii*- positive samples selected as to include isolates from all European
282 countries sampled in this study. Some samples (n = 25) were excluded because they
283 represented *B. garinii* mixed infections, and, therefore, allelic profiles could not be
284 determined. These complete 29 profiles represented all countries from which *B. garinii* was
285 detected in ticks from birds in this study, apart from Greece, for which we did not obtain any

286 complete profiles. Comparison of alleles from an incomplete ST (i.e. not obtaining sequences
287 for all alleles) from a tick feeding on a bird in Greece showed that they were identical to
288 samples previously reported from the UK.

289 The 29 *B. garinii* allelic profiles were resolved into 20 STs, nine of which were new.
290 These sequence data were supplemented with sequences of *B. garinii* isolates available at the
291 MLST database (see Sup. Mat. 3 for identification of isolates included in this analysis) and
292 used for goeBURST (n = 172; Fig.4) and phylogenetic analyses (n = 110; Fig. 5).

293 At a global scale, out of the 201 *B. garinii* isolates (137 STs) analysed (downloaded
294 from the MLST database and our own data), 2% (three STs: ST244, ST86 and ST575) were
295 found in more than one continent, 21% (29/137) were found in more than one country and
296 9.5% (13/137) were found in three or more countries. When a ST was detected in more than
297 one country, those countries were generally distant (i.e. did not border each other, 96.6%).
298 Eleven STs (out of the 20) found in ticks feeding on birds, and typed during the course of this
299 study, were detected in more than one country, and included two STs that were found in more
300 than one continent and four widespread STs (detected in 5 to 9 countries; Fig. 4). Among
301 these 20 STs described as part of this study, two were shared between migrant bird species
302 and species with both resident and mixed populations.

303 In the goeBURST analysis of the global collection (137 STs), using TLV as
304 parameter, 16 major clonal complexes and 4 minor clonal complexes (consisting of only two
305 associated STs) were found. Thirty-three isolates formed singletons (Fig 4). In five out of the
306 16 major clonal complexes, a clonal founder could be identified - those were ST86, ST88,
307 ST184, ST244 and ST251. The goeBURST analysis further revealed that STs from different
308 continents belonged to different clonal complexes, with only a few exceptions: seven out of

309 the 137 STs were shared between continents, or clustered together (e.g. ST364, ST694; Fig.
310 4). Two of the STs found in more than one continent were also detected in ticks from
311 European birds investigated during the course of this study.

312 Focusing on European STs, the pattern of clonal complexes was not related to
313 geographical distribution – there was no evidence that STs from different countries formed
314 separate clonal complexes, except for four STs detected only in Norway (ST487, ST488,
315 ST498 and ST516; Tveten, 2013) - Sup. Mat. 6, marked with an *). There was also no
316 indication of clustering according to bird species (Sup. Mat. 6).

317 STs detected in ticks from birds worldwide did not cluster tightly as clonal complex
318 but were distributed amongst clonal complexes, including those from migrant and birds which
319 have both resident and mixed populations (Sup. Mat. 7).

320 The averaged nucleotide diversity of *B. garinii* for all eight genes together was of the
321 same order for the three continents and all countries (ranging from 0.007 to 0.010), except
322 Norway and Sweden which presented the lowest nucleotide diversity ($\pi = 0.005$; Table 2).
323 Tajima's *D* was close to zero for most countries, showing that there is no specific sign of
324 selection or expansion in these genes. However, the population in Norway showed a
325 comparatively high Tajima's *D* of 1.085, which could be a sign of a bottleneck, being in
326 agreement with the low genetic diversity observed in this population.

327 At the European scale, *B. garinii* showed no spatial structuring in goeBURST analysis
328 (Sup. Mat. 6): STs for which more than one isolate has been obtained (e.g. ST86, ST187,
329 ST251, ST94, ST82) were not regionally restricted, but originated from distant countries such
330 as UK, Latvia, Slovenia, Hungary or Austria.

331 This is also what we observed on the ancestry recombination graph reconstructed
332 using sequences of the 85 European STs and generated in BEAST2 (Bouckaert et al., 2019);
333 Fig.5). This method was used because we suspected recombination between housekeeping
334 loci. Indeed 12 occurrences of recombination (dashed lines) were identified in at least 50% of
335 the sampled graphs in the phylogeny showing that recombination does occur but that there is
336 a global clonal frame. These recombination events concerned six out of the eight loci. The
337 ancestry recombination graph shows no evident geographical clustering for four main
338 European regions (Northern Europe - Estonia, Finland, Latvia, Norway and Sweden; Eastern
339 Europe - Czech Republic, Hungary, Serbia, Slovenia and Yugoslavia; Central Europe -
340 France, Germany, Italy, Netherlands; and British islands - United Kingdom) except for the 11
341 STs present in Norway (marked with square brackets on Fig.5) that cluster into three
342 monophyletic groups and one isolated ST. Out of the 85 STs present in Europe, 25% (n = 22)
343 were present in at least two of the geographical regions defined (Northern, Eastern, Central
344 Europe and the British islands). *Borrelia garinii* STs can thus disperse very far at the
345 continental scale. STs detected in birds (this study) were dispersed among other STs isolated
346 from ticks or humans.

347

348

349 4. DISCUSSION

350

351 In this study *I. ricinus* was the most abundant tick collected from common passerine birds
352 across a large geographical area in Europe. Overall, *B. burgdorferi* s.l prevalence in ticks
353 collected from birds was 37%. Thrushes (*Turdus* spp.) were the most important carriers of
354 infected *Ixodes* spp., supporting the notion that different bird species contribute differently to
355 *B. burgdorferi* s.l. genospecies complex maintenance and dispersal. Our genetic

356 characterization of the most prevalent genospecies detected in ticks feeding on birds, *B.*
357 *garii*, showed that this tick-borne pathogen presents little overlap of STs among continents,
358 but no geographical population structuring was detected in Europe, or according to isolation
359 source (bird-derived ticks or questing ticks/ human isolates). Taken together this provides
360 supporting evidence that birds act as important reservoirs for *B. garii* and are a main source
361 of infection of this genospecies to ticks and ultimately humans (through the bite of an infected
362 tick). Given the importance of birds as main hosts of this tick-borne pathogen, they have the
363 potential to modulate its phylogeography by homogenising the distribution of STs within the
364 European continental range through dispersal and migratory movements. Studying the
365 different factors in action driving this complex host-vector-parasite system is important for a
366 full understanding of *B. burgdorferi* s.l. enzootic cycle and potentially other (not only tick-
367 borne) bird-associated zoonotic pathogens.

368 *Ixodes ricinus* is a generalist tick and birds are known to be important hosts for its
369 immature stages (Norte et al., 2012; Santos-Silva et al., 2011). The other tick species (*I.*
370 *frontalis*, *I. arboricola* and *I. ventraloi*) and genera (*Haemaphysalis* and *Hyalomma*) collected
371 from birds in this study have also been previously reported on birds (Diakou et al., 2016;
372 Norte et al., 2012; Pérez-Eid, 2007) and differ in vector competence for *Borrelia* (Eisen &
373 Lane, 2002; Heylen, Krawczyk, et al., 2017; Heylen, Sprong, et al., 2014). Some bird species
374 such as hole-nesting birds (*P. major* and *F. albicollis*), *T. merula* and *T. philomelos* were
375 hosts for different tick species, however, the opportunities for co-feeding transmission of *B.*
376 *burgdorferi* s.l. between different tick species are limited by spatial and temporal tick species
377 distribution. The misidentification rate of ticks based on morphological features in our study
378 was similar to that reported for the genus *Ixodes* (14%; Estrada-Peña et al., 2017). Although *I.*
379 *persulcatus* occurs in part of the geographic range included in our study (e.g. Finland and

380 Estonia; ECDC 2018; Laaksonen et al., 2017), and its morphological distinction from *I.*
381 *ricinus* in immature stages is difficult, none of the tested samples were identified as *I.*
382 *persulcatus* by 16 rRNA sequencing.

383 Overall, the prevalence of *B. burgdorferi* s.l. (37%) was within the range reported for
384 ticks collected from hosts in The Netherlands and Belgium (34%; Heylen, Fonville, et al.,
385 2017). However, it was higher than in ticks collected from birds migrating through Italy
386 (30.7%; Toma et al., 2014) and Sweden (26.6%; Olsen et al., 1995), birds from central Europe
387 (25.8 – 28%; Dubska, Literak, Kocianova, Taragelova, & Sychra, 2009; Hanincova et al.,
388 2003; Taragel'ova et al., 2008), Germany (25%; Kipp, Goedecke, Dorn, Wilske, & Fingerle,
389 2006), Poland (13.3%; Michalik, Wodecka, Skoracki, Sikora, & Stanczak, 2008), Switzerland
390 (19.6-22.5%; Lommano, Bertaiola, Dupasquier, & Gern, 2012; Poupon et al., 2006), Spain
391 (9.2%; Palomar et al., 2016) and Portugal (7.3%; Norte et al., 2015). We cannot exclude the
392 possibility that this may be related to different methodologies used for *B. burgdorferi* s.l.
393 detection in different studies (real-time PCR *versus* conventional PCR and target genes). Our
394 results, using the same detection method across samples from different geographical origins
395 revealed that prevalence of *B. burgdorferi* s.l. in ticks from birds varied significantly
396 according to latitude. The fact that prevalence increased with latitude is in accordance with
397 Scandinavian countries such as Finland and Sweden showing relatively high prevalence when
398 compared with other European countries, as reported in previous studies on Lyme borreliosis
399 incidence and infection loads of questing ticks (Hubalek, 2009; Rauter & Hartung, 2005;
400 Wilhelmsson et al., 2013). A meta-analysis for Europe revealed an overall prevalence of
401 13.7% in questing ticks, higher in central Europe and Sweden, but with a significant increase
402 with longitude, rather than latitude, as in our study (Rauter & Hartung, 2005). Nonetheless, it
403 is also known that Lyme borreliosis presents a focal pattern of distribution, determined by the

404 heterogeneous spatial distribution of vector ticks, and also that the north-south gradient has a
405 greater influence on disease incidence at its distributional range limits (Hubalek, 2009). A
406 heterogeneous geographical distribution of *Borrelia* genospecies was also reported by Olsen
407 et al. (1995). These authors showed that *Borrelia* infections in ticks collected from birds
408 arriving to Sweden from the south or southeast in spring were mainly caused by *B. garinii*,
409 whereas the genospecies distribution was more heterogeneous in ticks from birds coming
410 from the southwest, and included *B. garinii*, *B. afzelii* and *B. burgdorferi* s.s.

411 The higher prevalence of *B. burgdorferi* s.l. in ticks removed from birds than that
412 reported from questing ticks is in accordance with birds acting as reservoirs for some *Borrelia*
413 genospecies and transmitting the infection to feeding ticks (Heylen, Matthysen, Fonville, &
414 Sprong, 2014; Humair, Postic, Wallich, & Gern, 1998b; Kurtenbach, Carey, Hoodless,
415 Nuttall, & Randolph, 1998; Norte, Lopes de Carvalho, Nuncio, Ramos, & Gern, 2013).
416 Additionally, *Borrelia* starts dividing in feeding ticks and may be more readily detected by
417 PCR (Schwan & Piesman, 2002). Our study revealed a non-homogeneous distribution of
418 *Borrelia* among bird species in bird-derived ticks. Thus, our data corroborate previous reports
419 that not all bird species contribute equally to the *Borrelia* enzootic cycle, as it is also known
420 for different mammal species (LoGiudice, Ostfeld, Schmidt, & Keesing, 2003; Talleklint &
421 Jaenson, 1994), and suggested by studies including different lizard species (Norte, Alves da
422 Silva, Alves, da Silva, Nuncio, et al., 2014; Szekeres, Majláthová, Majláth, & Földvári, 2016).
423 Contributing factors may not only be different infestation rates with vector ticks, which may
424 be related to foraging behaviour and consequent probability of exposure (Norte et al., 2012),
425 but also to other intrinsic factors related to host competence, including the host's adaptive and
426 innate immune system (Kurtenbach et al., 2006). *Turdus* species have been identified as main
427 reservoirs for *B. garinii* and *B. valaisana* in Europe (Dubska et al., 2011; Mannelli et al.,

428 2005; Michalik et al., 2008) and Asia (Miyamoto & Masuzawa, 2002). In addition, *T. merula*
429 has also been proven as competent reservoir for *B. turdi* through xenodiagnosis (Heylen,
430 Krawczyk, et al., 2017; Humair, Postic, Wallich, & Gern, 1998a; Norte et al., 2013). In an
431 experimental setup in which migratory restlessness was induced, latent *B. garinii* infections
432 were re-activated in *T. iliacus* (Gylfe, Bergstrom, Lunstrom, & Olsen, 2000). Although *B.*
433 *burgdorferi* s.l. infected ticks, including larvae, have occasionally been collected from *E.*
434 *rubecula* (this study; Poupon et al., 2006), our results suggest that this bird species, although
435 often infested by vector ticks (Norte et al., 2012), plays a minor role in *B. burgdorferi* s.l.
436 enzootic cycle because of the very low prevalence of *B. burgdorferi* s.l. in its ticks.

437 Ticks associated with hole-nesting birds such as *P. major* and *F. albicollis* presented
438 infection rates of 33.8 - 36.4%. The most prevalent genospecies was also *B. garinii*, which
439 made up 64 - 77.9% of infections in these bird species. *Parus major* has been shown
440 experimentally to selectively amplify *B. garinii* and *B. valaisiana*, whereas *B. afzelii*
441 prevalence in moulted adult ticks that fed as nymphs on this bird species tended to decrease in
442 successive infestations of the birds with wild questing nymphs (Heylen, Matthysen, et al.,
443 2014). The finding of this mammal-associated genospecies in attached ticks derived from
444 birds has been suggested to result from previously acquired infection from another (mammal)
445 host because these spirochetes were found to be unviable by culturing (Heylen et al., 2017). In
446 our study, all *B. afzelii*- positive ticks were nymphs and we cannot comment on birds'
447 reservoir competence for *B. afzelii* with these findings because PCR does not allow
448 distinguishing between viable and non-viable bacteria. We cannot rule out that nymphs
449 acquired the infection during a previous blood meal as larvae from a mammalian host, or that
450 larvae were infected via transovarial transmission (Bellet-Edimo, Betschart, & Gern, 2005),
451 because larvae of *I. ricinus* have been shown to transmit *B. afzelii* and *B. miyamotoi* to

452 vertebrate hosts (van Duijvendijk et al., 2016). In Europe, the role of transovarial transmission
453 for different tick and *Borrelia* species has not been resolved (Bellet-Edimo et al., 2005; Eisen
454 & Lane, 2002; Humair & L. Gern, 2000; van Duijvendijk et al., 2016). Thus, the role of birds
455 in *B. afzelli* transmission needs to be further scrutinized.

456 *Borrelia turdi*, originally described in Japan (Fukunaga et al., 1996), has been
457 increasingly detected in Europe, often in association with the ornithophilic tick *I. frontalis* and
458 its bird hosts (Heylen, Tijssse, Fonville, Matthysen, & Sprong, 2013; Norte et al., 2015). In
459 this study, it was detected only in Portugal, although it is known to be present in Spain,
460 Belgium and Norway (Hasle, Bjune, Midthjell, Røed, & Leinaas, 2011; Heylen et al., 2013;
461 Palomar et al., 2016). In our study it has been detected in *I. frontalis* and *I. ventalloi* only,
462 which are host- specialised tick species (to birds and rabbits, respectively; Hillyard, 1996).
463 *Ixodes frontalis* has been proven to be a competent vector for *B. turdi* (Heylen, Krawczyk, et
464 al., 2017) but vector competence of *I. ventalloi* remains unknown. This *Borrelia* genospecies
465 may have been overlooked in the past in questing ticks such as *I. ricinus* probably because of
466 its low prevalence (Heylen, Krawczyk, et al., 2017). Because *B. turdi* prevalence in our study
467 was relatively low, we were unable to evaluate statistically its bird host and vector species'
468 associations. Furthermore, the small sample size for tick species other than *I. ricinus*, may
469 have hampered the detection of significant associations between *Borrelia* and tick species, to
470 infer tick vector competence. Such relationships may be better evaluated in experimental
471 transmission studies (Heylen, Fonville, et al., 2017; Heylen, Sprong, et al., 2014).

472 Besides the avian-associated genospecies *B. valaisiana*, *B. garinii* and *B. turdi*, we
473 also detected DNA of a new *Borrelia* genospecies that has not been previously described.
474 Although PCR amplification and sequencing of 16S rRNA, *flaB* and *clpX* was possible and
475 clearly indicated the genetic distinction of the isolate from other *Borrelia* species, it is

476 conceivable that its genetic dissimilarity precluded a deeper characterisation involving other
477 house-keeping genes (which could not be amplified). This finding adds to the growing
478 evidence of the diversity of genospecies in circulation in cryptic cycles in bird hosts.
479 Specificities of reservoir host and/or vector competence may explain why this novel *Borrelia*
480 sp. genospecies was not detected before.

481 *Borrelia lusitaniae*, a genospecies whose main reservoirs are lizards (De Sousa et al.,
482 2012; Dsouli et al., 2006; Norte, Alves da Silva, Alves, da Silva, Nuncio, et al., 2014), has
483 been occasionally detected in ticks feeding on birds, including larvae (Poupon et al., 2006). In
484 our study, only one tick feeding on a bird was positive for *B. lusitaniae*. However, the paucity
485 of these findings suggests that birds, at most, have a minor role as reservoirs for this
486 genospecies. These infections could be the result of a previous incomplete blood meal on a
487 lizard, transovarial or co-feeding transmission. Surveys in endemic areas in Italy and Portugal
488 in which hundreds of bird-derived ticks were tested revealed no *B. lusitaniae* positive
489 specimens and thus, did not provide evidence that birds may serve as reservoir hosts for *B.*
490 *lusitaniae* (Amore et al., 2007; Norte, Alves da Silva, Alves, da Silva, Nuncio, et al., 2014;
491 Norte, Ramos, Gern, Nuncio, & Lopes de Carvalho, 2013).

492 Focusing on the genetic diversity and geographical distribution of the most prevalent
493 genospecies detected in ticks feeding on birds, the avian-associated *B. garinii*, we found that
494 its STs clustered according to continent showing some spatial structuring at this very wide
495 geographical scale. However, there was one ST shared between Europe and Asia, one ST
496 shared between Europe and North America, and one ST shared between Europe, Asia and
497 North America providing evidence of overlap among distant areas at a global scale. One
498 would expect that finding identical STs on continents separated by the Atlantic would be less
499 likely than that between adjacent continents whose geographical barriers may be easily

500 crossed by migrating birds. The movement of long-distance migratory birds, such as seabirds,
501 which can travel thousands of miles and between hemispheres may be responsible for the
502 spread of some *B. garinii* STs to distant geographical regions. *Borrelia garinii* is known to
503 circulate in a marine cycle involving the ornithophilic tick *I. uriae* that infests seabirds at their
504 colonies (Comstedt, Jakobsson, & Bergström, 2011; Gómez-Díaz et al., 2011). Migratory
505 shorebirds such as the black-tailed godwit *Limosa limosa*, the common redshank *Tringa*
506 *totanus*, and the little stint *Calidris minuta* were also reported to carry *B. garinii* (Lopes de
507 Carvalho et al., 2012). To this point, *B. garinii* isolates sharing the same *flaB* sequence have
508 been found in both Campbell Island (New Zealand), the Crozet Islands, and in the northern
509 hemisphere (Egg and St. Lazaria Islands, USA; Comstedt et al., 2011). In our study, two of
510 the transcontinental *B. garinii* STs were indeed found in ticks feeding on birds. One of these
511 (ST244) was found in *I. uriae* on a Canadian island (Munro et al., 2017), in questing *I. ricinus*
512 in Europe and *I. persulcatus* in Russia (<https://pubmlst.org/borrelia/>), in human isolates in
513 Germany (<https://pubmlst.org/borrelia/>), and in *Ixodes* spp. feeding in terrestrial birds in
514 Finland and Germany (this study). Some passerine birds (e.g. the northern wheatear *Oenanthe*
515 *oenanthe*) can also perform long distance migrations across the Atlantic (Bairlein et al.,
516 2012). An overlap and exchange of strains between the marine and terrestrial cycles is,
517 therefore, likely, as suggested by previous studies (Comstedt et al., 2011; Gómez-Díaz et al.,
518 2011). However, Gómez-Díaz et al. (2011) reported a population division of *B. garinii* from
519 seabirds between the Atlantic and Pacific basins. These researchers did not use the same
520 MLST as employed in our study, thus, immediate comparison of the results is not possible.

521 When evaluating European *B. garinii* STs only, no pattern of geographical clustering
522 was noticeable in our analysis, probably due to *B. burgdorferi* s.l./or ticks' dispersal promoted
523 by the birds. Similarly, although seabird species show high fidelity to breeding colonies

524 (Schreiber & Burger, 2001), and their main tick *I. uriae* occurs in seabird populations with
525 strong host species associations (McCoy et al., 2005), no geographic structuring was observed
526 in *B. garinii* within the Atlantic and Pacific Oceans (Gómez-Díaz et al., 2011). In contrast, the
527 mammal-associated *B. afzelii* STs were shown to have much less geographical overlap in
528 studies which compared geographical patterns and population structure of the avian-
529 associated *B. garinii* and this mammal-associated genospecies (*B. afzelii*), using the same
530 MLST scheme as this study (Vollmer et al., 2013; Vollmer et al., 2011). Their results
531 illustrated that *B. garinii* showed higher spatial mixing than *B. afzelii* but that *B. garinii*
532 presented population differentiation over a large geographical scale (Europe and China).
533 However, Vollmer et al. (2013) included fewer strains from a smaller geographical range in
534 Europe and China.

535 Although, in general, no overall apparent structure was found for European strains of
536 *B. garinii*, some Norwegian samples were divergent. This could be due to a relative isolation
537 of the study area in Norway, located in the northwest of the country (Tveten, 2013), or a
538 recent invasion event, which would explain the lower diversity of these strains. Recent
539 invasion would be consistent with the reported recent expansion of *I. ricinus* tick populations
540 to northern latitudes in Norway (Gray, Dautel, Estrada-Peña, Kahl, & Lindgren, 2009), which
541 could have caused a population bottleneck. This may also explain the evidence for selection
542 and expansion on MLST genes revealed by the relatively high Tajima's D in this *B. garinii*
543 population.

544 The uniform distribution of *B. garinii* STs among ticks collected from various bird
545 species, and other sources (e.g. questing ticks), does not suggest specialization of certain *B.*
546 *garinii* STs to certain hosts, contrary to the hypothesis of multiple niche polymorphism
547 associated with OspC variation (Dustin Brisson, Drecktrah, Eggers, & Samuels, 2012) found

548 for *Borrelia burgdorferi* sensu stricto (Brisson & Dykhuizen, 2004; Vuong et al., 2014), but
549 not for *B. afzelii* (Raberg et al., 2017). Our results are consistent with birds being the main
550 reservoir hosts of *B. garinii*: they maintain its natural transmission cycle and are the source of
551 infection for questing vector ticks. The lack of clustering of *B. garinii* STs regarding country
552 of origin or isolation source at a finer scale (i.e. Europe), was also supported by the ancestry
553 recombination graph. The clustering pattern between goeBURST (Sup. Mat 6) and that of the
554 ancestry recombination graph was generally similar with only a few exceptions showing
555 recombination among strains that could also be promoted by the avian-associated dispersal,
556 which may increase chances of encounter between different strain types and mixing of strains.

557 We should acknowledge that for migrant bird species, and for those which have both
558 resident and short-distance migrant populations, one cannot be completely confident that the
559 *B. burgdorferi* s.l. infections which the bird-infesting ticks carried were acquired in the
560 geographical area where the birds were captured. Birds (or their ticks) may have acquired the
561 bacteria in a different area where they remained or stopped-over during migration. This may
562 bias prevalence estimates and sequence type origin classification according to geographical
563 location.

564 The results presented in this study demonstrate how *B. burgdorferi* s.l.- vector- host
565 associations and the behaviour of hosts may shape and impact the spread and dispersal, and
566 ultimately the evolutionary biology of *B. burgdorferi* s.l., used here as a model for tick-borne
567 pathogens. Our data which includes *B. garinii* MLST characterization from bird-derived ticks
568 from the largest geographical range investigated so far substantiates that bird migration and
569 dispersal movements appear to be one of the main driving forces to shape *B. garinii*
570 populations, one of the most genetically heterogeneous Lyme borreliosis- causing
571 genospecies (Jacquot et al., 2014; Margos et al., 2008). Because birds are highly mobile and

572 the main reservoir hosts not only for *B. garinii*, but also for other pathogens, they contribute
573 to frequent, fast and long-range spatial mixing of strains and populations. Our study
574 underlines that understanding pathogen variability and spatial distribution, and consequent
575 modulation of transmission rates and evolution of new variants, is essential to understand
576 disease risk.

577

578 PUBLIC DATABASES ACCESSION NUMBERS

579

580 *Borrelia* sp. “*Candidatus Borrelia aligera*” 16S rRNA and *clpX* gene partial sequences
581 obtained in this study have been deposited in GenBank with the accession numbers
582 MH068784 and MH157920, respectively. *Borrelia garinii* MLST sequences have been
583 deposited in *Borrelia* MLST database (<https://pubmlst.org/borrelia/>) with the isolate id
584 numbers 2451 to 2479.

585

586 ACKNOWLEDGEMENTS

587

588 We would like to thank Cecilia Hizo-Teufel and Christine Hartberger for help with laboratory
589 analyses, Marko Mägi, Vallo Tilgar, Oscar Frías, Alejandra Toledo Vásquez, Alexia
590 Mouchet, Josef Heryan, Cinthya Lange, Piet de Goede, Henri Bouwmeester, Esa Lehtikoinen,
591 Franck Théron, Petra Bandelj, Tea Knapič, Irena Kodele Krašna, Pavle Štirn, Katarina
592 Prosenc Trilar, Modest Vengušt, Hanna Holmström, Jorma Nurmi, Miia Rainio, Pablo
593 Sánchez-Virosta, Silvia Espín and Lucy Winder who helped with tick collection, Vítor Hugo
594 Paiva for help with statistical analyses, the Falsterbo Ringing Station and Instituto da

595 Conservação da Natureza e Florestas IP and the Slovenian Bird Ringing Center at Slovenian
596 Museum of Natural History for providing conditions for fieldwork and bird ringing. We thank
597 the two anonymous reviewers by their insightful comments on this paper. This study received
598 financial support from Fundação para a Ciência e a Tecnologia by the strategic program of
599 MARE (MARE - UID/MAR/04292/2013) and the fellowship to Ana Cláudia Norte
600 (SFRH/BPD/108197/2015), and the Portuguese National Institute of Health. Raivo Mänd,
601 Tomi Trilar, Tapio Eeva and Tomas Grim were supported by the Estonian Research
602 Council (research grant # IUT34-8), the Slovenian Research Agency -programme
603 “Communities, relations and communications in the ecosystems” (No. P1-0255), the
604 Academy of Finland (project 265859), and the Internal Grant Agency of Palacky University
605 (PrF_2014_018, PrF_2015_018, PrF_2013_018), respectively. All applicable institutional
606 and/or national guidelines for the care and use of animals were followed in this study.

REFERENCES

- Amore, G., Tomassone, L., Grego, E., Ragagli, C., Bertolotti, L., Nebbia, P., . . . Mannelli, A. (2007). *Borrelia lusitaniae* in immature *Ixodes ricinus* (Acari : Ixodidae) feeding on common wall lizards in Tuscany, central Italy. *Journal of Medical Entomology*, 44(2), 303-307.
- Bairlein, F., Norris, D. R., Nagel, R., Bulte, M., Voigt, C. C., Fox, J. W., . . . Schmaljohann, H. (2012). Cross-hemisphere migration of a 25 g songbird. *Biology Letters*, 8(4), 505-507. doi:10.1098/rsbl.2011.1223
- Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting Linear Mixed-Effects Models Using lme4. 2015, 67(1), 48. doi:10.18637/jss.v067.i01
- Bellet-Edimo, R., Betschart, B., & Gern, L. (2005). Frequency and efficiency of transovarial and subsequent transtadial transmission of *Borrelia burgdorferi* sensu lato in *Ixodes ricinus* ticks. *Bulletin de la Société Neuchâteloise des Sciences Naturelles*, 128, 117-125.
- Bouckaert, R., Vaughan, T. G., Barido-Sottani, J., Duchêne, S., Fourment, M., Gavryushkina, A., . . . Drummond, A. J. (2019). BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. *PLOS Computational Biology*, 15(4), e1006650. doi:10.1371/journal.pcbi.1006650
- Brisson, D., Drecktrah, D., Eggers, C., Samuels, D.S. (2012) Genetics of *Borrelia burgdorferi*. *Annual Review of Genetics*, 46, 515-536.
- Brisson, D. & Dykhuizen, D. E. (2004). ospC diversity in *Borrelia burgdorferi*: different hosts are different niches. *Genetics*, 168(2), 713-722.
- Casjens, S. R., Fraser-Liggett, C. M., Mongodin, E. F., Qiu, W.-G., Dunn, J. J., Luft, B. J., & Schutzer, S. E. (2011). Whole Genome Sequence of an Unusual *Borrelia burgdorferi* Sensu Lato Isolate. *Journal of Bacteriology*, 193(6), 1489-1490. doi:10.1128/jb.01521-10
- Chitimia-Dobler, L., Rieß, R., Kahl, O., Wöfel, S., Dobler, G., Nava, S. & Estrada- Peña, A. (2018). *Ixodes inopinatus* – Occurring also outside the Mediterranean region. *Ticks and Tick-borne Diseases*, 9, 196-200.
- Comstedt, P., Jakobsson, T., & Bergström, S. (2011). Global ecology and epidemiology of *Borrelia garinii* spirochetes. *Infection Ecology and Epidemiology* 1: doi: 10.3402/iee.v1i0.9545.
- De Sousa, R., Lopes de Carvalho, I., Santos, A. S., Bernardes, C., Milhano, N., Jesus, J., . . . Nuncio, M. S. (2012). The Role of *Teira dugesii* lizard as potential host for *Ixodes ricinus* tick-borne pathogens. *Applied and Environmental Microbiology*. doi:10.1128/aem.07945-11
- Diakou, A., Norte, A. C., Lopes de Carvalho, I., Nuncio, S., Novakova, M., Kautman, M., . . . Literak, I. (2016). Ticks and tick-borne pathogens in wild birds in Greece. *Parasitology Research*, 115(5), 2011-2016. doi:10.1007/s00436-016-4943-3
- Didelot, X., Lawson, D., Darling, A., & Falush, D. (2010). Inference of Homologous Recombination in Bacteria Using Whole-Genome Sequences. *Genetics*, 186(4), 1435-1449. doi:10.1534/genetics.110.120121
- Dsouli, N., Younsi-Kabachii, H., Postic, D., Nouria, S., Gern, L., & Bouattour, A. (2006). Reservoir role of the lizard, *Psammotromus algirus*, in the transmission cycle of

- Borrelia burgdorferi* sensu lato (Spirochaetacea) in Tunisia. *Journal of Medical Entomology*, 43, 737-742.
- Dubska, L., Literak, I., Kocianova, E., Taragelova, V., Sverakova, V., & Sychra, O. (2011). Synanthropic birds influence the distribution of *Borrelia* species: analysis of *Ixodes ricinus* ticks feeding on passerine birds. *Applied and Environmental Microbiology*, 77. doi:10.1128/aem.02278-10
- Dubska, L., Literak, I., Kocianova, E., Taragelova, V., & Sychra, O. (2009). Differential Role of Passerine Birds in Distribution of *Borrelia* Spirochetes, Based on Data from Ticks Collected from Birds during the Postbreeding Migration Period in Central Europe. *Applied and Environmental Microbiology*, 75(3), 596-602.
- Eisen, L., & Lane, R. S. (2002). Vectors of *Borrelia burgdorferi* sensu lato. In J. Gray, O. Kahl, R. S. Lane, & G. Stanek (Eds.), *Lyme Borreliosis: Biology, Epidemiology and Control* (pp. 91-115). NY: CAB Publishing.
- Estrada-Peña, A., Bouattour, A., Camicas, J.-L., & Walker, A. R. (2004). *Ticks of domestic animals in the Mediterranean Region - A guide to identification of species*. Zaragoza: University of Zaragoza.
- Estrada-Peña, A., D'Amico, G., Palomar, A. M., Dupraz, M., Fonville, M., Heylen, D., . . . Mihalca, A. D. (2017). A comparative test of ixodid tick identification by a network of European researchers. *Ticks and Tick Borne Diseases*, 8(4), 540-546. doi:10.1016/j.ttbdis.2017.03.001
- Estrada-Peña, A., Nava, S., & Petney, T. (2014). Description of all the stages of *Ixodes inopinatus* n. sp. (Acari: Ixodidae). *Ticks and Tick Borne Diseases*, 5, 734-743. doi:10.1016/j.ttbdis.2014.05.003
- European Centre for Disease Prevention and Control and European Food Safety Authority. *Mosquito maps [internet]*. Stockholm: ECDC; 2018. Available from: <https://ecdc.europa.eu/en/disease-vectors/surveillance-and-disease-data/tick-maps>
- Francisco, A. P., Bugalho, M., Ramirez, M., & Carriço, J. A. (2009). Global optimal eBURST analysis of multilocus typing data using a graphic matroid approach. *BMC Bioinformatics*, 10(1), 152. doi:10.1186/1471-2105-10-152
- Fukunaga, M., Hamase, A., Okada, K., Inoue, H., Tsuruta, Y., Miyamoto, K., & Nakao, M. (1996). Characterization of spirochetes isolated from ticks (*Ixodes tanuki*, *Ixodes turdus*, and *Ixodes columnae*) and comparison of the sequences with those of *Borrelia burgdorferi* sensu lato strains. *Applied and Environmental Microbiology*, 62(7), 2338-2344.
- Gómez-Díaz, E., Boulinier, T., Sertour, N., Cornet, M., Ferquel, E., & McCoy, K. D. (2011). Genetic structure of marine *Borrelia garinii* and population admixture with the terrestrial cycle of Lyme borreliosis. *Environmental Microbiology*, 13(9), 2453-2467. doi:10.1111/j.1462-2920.2011.02515.x
- Gray, J. S., Dautel, H., Estrada-Peña, A., Kahl, O., & Lindgren, E. (2009). Effects of Climate Change on Ticks and Tick-Borne Diseases in Europe. *Interdisciplinary Perspectives on Infectious Diseases*: 593232.
- Gylfe, A., Bergstrom, S., Lunstrom, J., & Olsen, B. (2000). Epidemiology - Reactivation of *Borrelia* infection in birds. *Nature*, 403(6771), 724-725.
- Hanincova, K., Taragelova, V., Koci, J., Schafer, S. M., Hails, R., & Ullmann, A. J. (2003). Association of *Borrelia garinii* and *B. valaisiana* with songbirds in Slovakia. *Applied and Environmental Microbiology*, 69, 2825-2830. doi:10.1128/aem.69.5.2825-2830.2003

- Hasegawa, M., Kishino, H., & Yano, T. (1985). Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *J Mol Evol*, *22*(2), 160-174.
- Hasle, G., Bjune, G. A., Midthjell, L., Røed, K. H., & Leinaas, H. P. (2011). Transport of *Ixodes ricinus* infected with *Borrelia* species to Norway by northward-migrating passerine birds. *Ticks and Tick-borne Diseases*, *2*(1), 37-43.
- Heylen, D. (2016). Ecological interactions between songbirds, ticks, and *Borrelia burgdorferi* s.l. in Europe. In M. A. H. Braks, S. E. van Wieren, W. Takken, & H. Sprong (Eds.), *Ecology and Control of Vector-borne diseases* (Vol. 4, pp. 91 - 101). Wageningen: Wageningen Academic Publishers.
- Heylen, D., Fonville, M., Docters van Leeuwen, A., Stroo, A., Duisterwinkel, M., van Wieren, S., . . . Sprong, H. (2017). Pathogen communities of songbird-derived ticks in Europe's low countries. *Parasites & Vectors*, *10*(1), 497. doi:10.1186/s13071-017-2423-y
- Heylen, D., Krawczyk, A., Lopes de Carvalho, I., Nuncio, M. S., Sprong, H., & Norte, A. C. (2017). Bridging of cryptic *Borrelia* cycles in European songbirds. *Environmental Microbiology*, *19*, 1857-1867. doi:10.1111/1462-2920.13685
- Heylen, D., Matthysen, E., Fonville, M., & Sprong, H. (2014). Songbirds as general transmitters but selective amplifiers of *Borrelia burgdorferi* sensu lato genotypes in *Ixodes ricinus* ticks. *Environmental Microbiology*, *16*(9), 2859-2868. doi:10.1111/1462-2920.12304
- Heylen, D., Sprong, H., van Oers, K., Fonville, M., Leirs, H., & Matthysen, E. (2014). Are the specialized bird ticks, *Ixodes arboricola* and *I. frontalis*, competent vectors for *Borrelia burgdorferi* sensu lato? *Environmental Microbiology*, *16*(4), 1081-1089. doi:10.1111/1462-2920.12332
- Heylen, D., Tijssse, E., Fonville, M., Matthysen, E., & Sprong, H. (2013). Transmission dynamics of *Borrelia burgdorferi* s.l. in a bird tick community. *Environmental Microbiology*, *15*(2), 663-673. doi:10.1111/1462-2920.12059
- Heylen, D. J. A., Sprong, H., Krawczyk, A., Van Houtte, N., Genné, D., Gomez-Chamorro, A., . . . Voordouw, M. J. (2017). Inefficient co-feeding transmission of *Borrelia afzelii* in two common European songbirds. *Scientific Reports*, *7*, 39596. doi:10.1038/srep39596
- Hillyard, P. D. (1996). *Ticks of North-West Europe*. London: Backhuys Publishers.
- Hoën, A. G., Margos, G., Bent, S. J., Diuk-Wasser, M. A., Barbour, A., Kurtenbach, K., & Fish, D. (2009). Phylogeography of *Borrelia burgdorferi* in the eastern United States reflects multiple independent Lyme disease emergence events. *Proceedings of the National Academy of Sciences of the United States of America*, *106*(35), 15013-15018.
- Hubalek, Z. (2009). Epidemiology of lyme borreliosis. *Current Problems in Dermatology*, *37*, 31-50. doi:10.1159/000213069
- Humair, P.-F., & Gern, L. (2000). The wild hidden face of Lyme borreliosis in Europe. *Microbes and Infection*, *2*(8), 915-922.
- Humair, P. F., Postic, D., Wallich, R., & Gern, L. (1998). An avian reservoir (*Turdus merula*) of the Lyme borreliosis spirochetes. *International Journal of Medical Microbiology Virology Parasitology Infectious Diseases*, *287*, 521-538.
- Jacquot, M., Gonnet, M., Ferquel, E., Abrial, D., Claude, A., Gasqui, P., . . . Bailly, X. (2014). Comparative population genomics of the *Borrelia burgdorferi* species complex reveals high degree of genetic isolation among species and underscores benefits and constraints to studying intra-specific epidemiological processes. *PLoS ONE*, *9*(4), e94384. doi:10.1371/journal.pone.0094384

- Johnson, B. J., Happ, C. M., Mayer, L. W., & Piesman, J. (1992). Detection of *Borrelia burgdorferi* in ticks by species-specific amplification gene. *American Journal of Tropical Medicine and Hygiene.*, 47, 730-741.
- Jolley, K. A., & Maiden, M. C. (2010). BIGSdb: Scalable analysis of bacterial genome variation at the population level. *BMC Bioinformatics*, 11(1), 595. doi:10.1186/1471-2105-11-595
- Katoh, K., & Standley, D. M. (2013). MAFFT Multiple Sequence Alignment Software Version 7: Improvements in Performance and Usability. *Molecular Biology and Evolution*, 30(4), 772-780. doi:10.1093/molbev/mst010
- Kipp, S., Goedecke, A., Dorn, W., Wilske, B., & Fingerle, V. (2006). Role of birds in Thuringia, Germany, in the natural cycle of *Borrelia burgdorferi sensu lato*, the Lyme disease spirochaete. *International Journal of Medical Microbiology*, 296 Suppl 40, 125-128.
- Kurtenbach, K., Carey, D., Hoodless, A. N., Nuttall, P. A., & Randolph, S. E. (1998). Competence of pheasants as reservoirs for Lyme disease spirochetes. *Journal of Medical Entomology*, 35(1), 77-81.
- Kurtenbach, K., De Michelis, S., Etti, S., Schafer, S. M., Sewell, H. S., Brade, V., & Kraiczy, P. (2002). Host association of *Borrelia burgdorferi sensu lato* - the key role of host complement. *Trends in Microbiology*, 10(2), 74-79.
- Kurtenbach, K., Gatewood, A., Bent, S. J., Vollmer, S. A., Ogden, and, N. H., & Margos, G. (2010). Population Biology of Lyme Borreliosis Spirochetes. In D. F. a. E. J. F. D. A. Robinson (Ed.), *Bacterial Population Genetics in Infectious Disease*. New Jersey: John Wiley & Sons, Inc. .
- Kurtenbach, K., Hanincova, K., Tsao, J. I., Margos, G., Fish, D., & Ogden, N. H. (2006). Fundamental processes in the evolutionary ecology of Lyme borreliosis. *Nature Reviews Microbiology*, 4(9), 660-669.
- Kurtenbach, K., Peacey, M., Rijpkema, S. G., Hoodless, A. N., Nuttall, P. A., & Randolph, S. E. (1998). Differential transmission of the genospecies of *Borrelia burgdorferi sensu lato* by game birds and small rodents in England. *Applied and Environmental Microbiology*, 64, 1169-1174.
- Kuznetsova, A., Brockhoff, P. B., & Christensen, R. H. B. (2017). lmerTest Package: Tests in Linear Mixed Effects Models. *Journal of Statistical Software*, 82, 1-26.
- Laaksonen, M., Sajanti, E., Sormunen, J. J., Penttinen, R., Hanninen, J., Ruohomaki, K., . . . Klemola, T. (2017). Crowdsourcing-based nationwide tick collection reveals the distribution of *Ixodes ricinus* and *I. persulcatus* and associated pathogens in Finland. *Emerging Microbes and Infection*, 6(5), e31. doi:10.1038/emi.2017.17
- LoGiudice, K., Ostfeld, R.S., Schmidt, K.A. & Keesing, F. (2003). The ecology of infectious disease: Effects of host diversity and community composition on Lyme disease risk. *Proceedings of the National Academy of Sciences of the United States of America*, 100(2): 567-571.
- Lommano, E., Bertaiola, L., Dupasquier, C., & Gern, L. (2012). Infections and coinfections of questing *Ixodes ricinus* ticks by emerging zoonotic pathogens in Western Switzerland. *Applied and Environmental Microbiology*, 78(13), 4606-4612. doi:10.1128/aem.07961-11
- Lopes de Carvalho, I., Zé-Zé, L., Alves, A., Pardal, S., Lopes, R. J., Mendes, L., & Nuncio, M. S. (2012). *Borrelia garinii* and *Francisella tularensis* subsp *holarctica* detected in migratory shorebirds in Portugal. *European Journal of Wildlife Research*, 58, 857-861

- Mangold, A. J., Bargues, M. D., & Mas-Coma, S. (1998). Mitochondrial 16S rDNA sequences and phylogenetic relationships of species of *Rhipicephalus* and other tick genera among Metastriata (Acari: Ixodidae). *Parasitology Research*, *84*(6), 478-484.
- Mannelli, A., Nebbia, P., Tramuta, C., Grego, E., Tomassone, L., Ainaudi, R., . . . Meneguz, P. G. (2005). *Borrelia burgdorferi* sensu lato infection in larval *Ixodes ricinus* (Acari : Ixodidae) feeding on blackbirds in northwestern Italy. *Journal of Medical Entomology*, *42*(2), 168-175.
- Margos, G., Chu, C. Y., Takano, A., Jiang, B. G., Liu, W., Kurtenbach, K., . . . Kawabata, H. (2015). *Borrelia yangtzensis* sp. nov. a rodent associated species in Asia is related to *B. valaisiana*. *International Journal of Systematic and Evolutionary Microbiology*, *65*, 3836-3840. doi:10.1099/ijsem.0.000491
- Margos, G., Gatewood, A. G., Aanensen, D. M., Hanincová, K., Terekhova, D., Vollmer, S. A., . . . Kurtenbach, K. (2008). MLST of housekeeping genes captures geographic population structure and suggests a European origin of *Borrelia burgdorferi*. *Proceedings of the National Academy of Sciences*, *105*(25), 8730-8735. doi:10.1073/pnas.0800323105
- Margos, G., Tsao, J. I., Castillo-Ramírez, S., Girard, Y. A., Hamer, S. A., Hoen, A. G., . . . Ogden, N. H. (2012). Two Boundaries Separate *Borrelia burgdorferi* Populations in North America. *Applied and Environmental Microbiology*, *78*(17), 6059-6067. doi:10.1128/aem.00231-12
- Margos, G., Vollmer, S. A., Ogden, N. H., & Fish, D. (2011). Population genetics, taxonomy, phylogeny and evolution of *Borrelia burgdorferi* sensu lato. *Infection, Genetics and Evolution*, *11*(7), 1545-1563. doi:10.1016/j.meegid.2011.07.022
- McCoy, K. D., Chapuis, E., Tirard, C., Boulonier, T., Michalakis, Y., Bohec, C. L., . . . Gauthier-Clerc, M. (2005). Recurrent evolution of host-specialized races in a globally distributed parasite. *Proceedings of the Royal Society B: Biological Sciences*, *272*(1579), 2389-2395. doi:10.1098/rspb.2005.3230
- Michalik, J., Wodecka, B., Skoracki, M., Sikora, B., & Stanczak, J. (2008). Prevalence of avian-associated *Borrelia burgdorferi* s.l. genospecies in *Ixodes ricinus* ticks collected from blackbirds (*Turdus merula*) and song thrushes (*T. philomelos*). *International Journal of Medical Microbiology*, *298*, 129-138.
- Miyamoto, K., & Masuzawa, T. (2002). Ecology of *Borrelia burgdorferi* sensu lato in Japan and East Asia. In J. Gray, O. Kahl, R. S. Lane, & G. Stanek (Eds.), *Lyme Borreliosis: Biology, Epidemiology and Control* (pp. 201-222). Oxford, UK: CABI International.
- Munro, H. J., Ogden, N. H., Lindsay, L. R., Robertson, G. J., Whitney, H., & Lang, A. S. (2017). Evidence for *Borrelia bavariensis* infections of *Ixodes uriae* within seabird colonies of the North Atlantic Ocean. *Applied and Environmental Microbiology*. doi:10.1128/aem.01087-17
- Nei, M. (1987). *Molecular Evolutionary Genetics* New York: Columbia Univ. Press.
- Norte, A.C., Margos, G., Becker N.S., Ramos, J.A., Nuncio, M.S., Fingerle, V., Araújo, P. M., . . . Lopes de Carvalho, I. (2019) https://pubmlst.org/bigssdb?db=pubmlst_borrelia_isolates (isolate id numbers 2451 to 2479)
- Norte, A.C., Margos, G., Becker N.S., Ramos, J.A., Nuncio, M.S., Fingerle, V., Araújo, P. M., . . . Lopes de Carvalho, I. (2019) <https://www.ncbi.nlm.nih.gov/nucleotide/> (accession numbers MH068784 and MH157920)
- Norte, A., de Carvalho, I., Ramos, J., Gonçalves, M., Gern, L., & Nuncio, M. (2012). Diversity and seasonal patterns of ticks parasitizing wild birds in western Portugal. *Experimental and Applied Acarology*, *58*, 327-339. doi:10.1007/s10493-012-9583-4

- Norte, A. C., Alves da Silva, A., Alves, J., da Silva, L. P., Nuncio, M. S., Escudero, R., & Lopes de Carvalho, I. (2014). The importance of lizards and small mammals as reservoirs for *Borrelia lusitaniae* in Portugal. *Environmental Microbiology Reports*, *7*, 188-193. doi:10.1111/1758-2229.12218
- Norte, A. C., Lopes de Carvalho, I., Nuncio, M. S., Ramos, J. A., & Gern, L. (2013). Blackbirds *Turdus merula* as competent reservoirs for *Borrelia turdi* and *Borrelia valaisiana* in Portugal: evidence from a xenodiagnostic experiment. *Environmental Microbiology Reports*, *5*, 604-607. doi:10.1111/1758-2229.12058
- Norte, A. C., Pascoal da Silva, L., Tenreiro, P. J. Q., Felgueiras, M. S., Araújo, P. M., Lopes, P. B., . . . Lopes de Carvalho, I. (2015). Patterns of tick infestation and their *Borrelia burgdorferi* s.l. infection in wild birds in Portugal. *Ticks and Tick-borne Diseases*, *6*, 743-750. doi:DOI: 10.1016/j.ttbdis.2015.06.010
- Norte, A. C., Ramos, J. A., Gern, L., Nuncio, M. S., & Lopes de Carvalho, I. (2013). Birds as reservoirs for *Borrelia burgdorferi* s.l. in western Europe: circulation of *B. turdi* and other genospecies in bird-tick cycles in Portugal. *Environmental Microbiology*, *15*, 386-397. doi:doi: 10.1111/j.1462-2920.2012.02834.x.
- Ogden, N. H., Lindsay, L. R., Hanincova, K., Barker, I. K., Bigras-Poulin, M., Charron, D. F., & Thompson, R. A. (2008). Role of Migratory Birds in Introduction and Range Expansion of *Ixodes scapularis* Ticks and of *Borrelia burgdorferi* and *Anaplasma phagocytophilum* in Canada, *74*, 1780-1790
- Olsen, B., Jaenson, T. G. T., & Bergstrom, S. (1995). Prevalence of *Borrelia burgdorferi* Sensu Lato- Infected Ticks on Migrating Birds. *Applied and Environmental Microbiology*, *61*(8), 3082-3087.
- Palomar, A. M., Aránzazu Portillo, Paula Santibáñez, David Mazuelas, Lidia Roncero, Óscar Gutiérrez, & Oteo, J. A. (2016). Presence of *Borrelia turdi* and *Borrelia valaisiana* (Spirochaetales: Spirochaetaceae) in Ticks Removed From Birds in the North of Spain, 2009-2011. *Journal of Medical Entomology*, *54*, 243-246.
- Paradis, E. (2010). pegas: an R package for population genetics with an integrated-modular approach. *Bioinformatics*, *26*(3), 419-420. doi:10.1093/bioinformatics/btp696
- Paradis, E., Claude, J., & Strimmer, K. (2004). APE: Analyses of Phylogenetics and Evolution in R language. *Bioinformatics*, *20*(2), 289-290.
- Pérez-Eid, C. (2007). *Les tiques: identification, biologie, importance médicale et vétérinaire* (Editions Médicales Internationales TEC&DOC): Lavoisier.
- Poupon, M. A., Lommano, E., Humair, P. F., Douet, W., Rais, O., Schaad, M., . . . Gern, L. (2006). Prevalence of *Borrelia burgdorferi* sensu lato in ticks collected from migratory birds in Switzerland. *Applied and Environmental Microbiology*, *72*(1), 976-979.
- Raberg, L., Hagstrom, A., Andersson, M., Bartkova, S. Scherman, K., Strandh, M. & Tschirren, B. (2017). Evolution of antigenic diversity in the tick-transmitted bacterium *Borrelia afzelii*: a role for host specialization? *Journal of Evolutionary Biology*, *30*(5): 1034-1041.
- Radulović, Z., Milutinovic, M., Tomanovic, S., & Mulenga, A. (2010). Detection of *Borrelia*-specific 16S rRNA sequence in total RNA extracted from *Ixodes ricinus* ticks. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, *62*, 862-867.
- Rambaut, A., Suchard, M.A., Xie, D. & Drummond, A.J. (2014) Tracer v1.6. Available from <http://beast.bio.ed.ac.uk/Tracer/>

- Randolph, S. E., Gern, L., & Nuttall, P. A. (1996). Co-feeding ticks: Epidemiological significance for tick-borne pathogen transmission. *Parasitology Today*, 12(12), 472-479.
- Rauter, C., & Hartung, T. (2005). Prevalence of *Borrelia burgdorferi* sensu lato genospecies in *Ixodes ricinus* ticks in Europe: a metaanalysis. *Applied and Environmental Microbiology*, 71, 7203-7216. doi:10.1128/aem.71.11.7203-7216.2005
- R Core Team (2013). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>.
- Reed, K. D., Meece, J. K., Henkel, J. S., & Shukla, S. K. (2003). Birds, Migration and Emerging Zoonoses: West Nile Virus, Lyme Disease, Influenza A and Enteropathogens. *Clinical and Medical Research*, 1, 5–12.
- Rijpkema, A. G. T., Molkenboer, M. J. C. H., Schouls, L. M., Jongejan, F., & Schellekens, J. F. P. (1995). Simultaneous detection and genotyping of three genomic groups of *Borrelia burgdorferi* sensu lato in Dutch *Ixodes ricinus* Ticks by characterization of the amplified intergenic spacer region between 5S and 23S rRNA genes. *Journal of Clinical Microbiology*, 33, 3091-3095.
- Santos-Silva, M., Beati, L., Santos, A., De Sousa, R., Nuncio, M., Melo, P., . . . Bacellar, F. (2011). The hard-tick fauna of mainland Portugal (Acari: Ixodidae): an update on geographical distribution and known associations with hosts and pathogens. *Experimental and Applied Acarology*, 55(1), 85-121.
- Schreiber, E. A., & Burger, J. (Eds.). (2001). *Biology of Marine Birds*. Boca Raton: CRC Press.
- Schwan, T. G., & Piesman, J. (2002). Vector Interactions and Molecular Adaptations of Lyme Disease and Relapsing Fever Spirochetes Associated with Transmission by Ticks. *Emerging Infectious Diseases*, 8(2), 115-121. doi:10.3201/eid0802.010198
- Sonenshine, D. E., & Mather, T. N. (1994). *Ecological dynamics of tick-borne zoonoses*. New York: Oxford University Press.
- Szekeres S, Majláthová V, Majláth I & Földvári G (2016) Neglected hosts: the role of lacertid lizards and medium-sized mammals in the eco-epidemiology of Lyme borreliosis. In: Braks, M.A.H., Van Wieren, S.E., Takken, W. & Sprong, H. (eds.) Ecology and prevention of Lyme borreliosis. Ecology and Control of Vector-borne diseases, Volume 4. Wageningen Academic Publishers, Wageningen, the Netherlands, pp. 103-126.
- Tajima, F. (1989). Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics*, 123(3), 585-595.
- Talleklint, L. & Jaenson T. G. (1994). Transmission of *Borrelia burgdorferi* s.l. from mammal reservoirs to the primary vector of Lyme borreliosis, *Ixodes ricinus* (Acari: Ixodidae), in Sweden." *Journal of Medical Entomology*, 31(6): 880-886.
- Taragel'ova, V., Koci, J., Hanincova, K., Kurtenbach, K., Derdakova, M., & Ogden, N. H. (2008). Blackbirds and song thrushes constitute a key reservoir of *Borrelia garinii*, the causative agent of borreliosis in Central Europe. *Applied and Environmental Microbiology*, 74, 1289-1293. doi:10.1128/aem.01060-07
- Thomas, N. J., Hunter, D. B., & Atkinson, C. T. (2007). *Infectious Diseases of Wild Birds*. Oxford: Wiley-Blackwell.
- Toma, L., Mancini, F., Di Luca, M., Cecere, J. G., Bianchi, R., Khoury, C., . . . Ciervo, A. (2014). Detection of microbial agents in ticks collected from migratory birds in central Italy. *Vector Borne and Zoonotic Diseases*, 14(3), 199-205. doi:10.1089/vbz.2013.1458

- Tveten, A.-K. (2013). Prevalence of *Borrelia burgdorferi* sensu stricto, *Borrelia afzelii*, *Borrelia garinii*, and *Borrelia valaisiana* in *Ixodes ricinus* ticks from the northwest of Norway. *Scandinavian Journal of Infectious Diseases*, 45(9), 681-687. doi:10.3109/00365548.2013.799288
- van Duijvendijk, G., Coipan, C., Wagemakers, A., Fonville, M., Ersöz, J., Oei, A., . . . Sprong, H. (2016). Larvae of *Ixodes ricinus* transmit *Borrelia afzelii* and *B. miyamotoi* to vertebrate hosts. *Parasites & Vectors*, 9(1), 1-7. doi:10.1186/s13071-016-1389-5
- Vaughan, T. G., Welch, D., Drummond, A. J., Biggs, P. J., George, T., & French, N. P. (2017). Inferring Ancestral Recombination Graphs from Bacterial Genomic Data. *Genetics*, 205(2), 857-870. doi:10.1534/genetics.116.193425
- Vollmer, S., Feil, E. J., Chu, C.-Y., Raper, S. L., Cao, W.-C., Kurtenbach, K., . . . Margos, G. (2013). *Spatial spread and demographic expansion of Lyme borreliosis spirochaetes in Eurasia*. Poster presented at the ICLB2013.
- Vollmer, S. A., Bormane, A., Dinnis, R. E., Seelig, F., Dobson, A. D. M., Aanensen, D. M., . . . Margos, G. (2011). Host migration impacts on the phylogeography of Lyme Borreliosis spirochaete species in Europe. *Environmental Microbiology*, 13(1), 184-192.
- Vuong, H.B., Canham, C.D., Fonseca, D.M., Brisson, D., Morin, P.J., Smouse, P.E., & Otsfeld, R.S. (2014). Occurrence and transmission efficiencies of *Borrelia burgdorferi* ospC types in avian and mammalian wildlife. *Infection, Genetics and Evolution* 27, 594-600.
- Wilhelmsson, P., Lindblom, P., Fryland, L., Ernerudh, J., Forsberg, P., & Lindgren, P.-E. (2013). Prevalence, Diversity, and Load of *Borrelia* species in Ticks That Have Fed on Humans in Regions of Sweden and Åland Islands, Finland with Different Lyme Borreliosis Incidences. *PLoS ONE*, 8(11), e81433. doi:10.1371/journal.pone.0081433
- Zhioua, E., Bouattour, A., Hu, C. M., Gharbi, M., Aeschliman, A., Ginsberg, H. S., & Gern, L. (1999). Infection of *Ixodes ricinus* (Acari: Ixodidae) by *Borrelia burgdorferi* sensu lato in North Africa. *Journal of Medical Entomology*, 36(2), 216-218.

Table 1. Conditional model averaged coefficient parameters from the generalized linear mixed models (GLMMs) that best explained (lowest AICc, $\Delta AICc < 2$, see Sup. Mat. 2) the prevalence of *B. burgdorferi* s.l. in ticks collected from birds.

Conditional averaged model coefficients					
Parameter	Estimate	SE	Adjusted SE	Z Value	P
Intercept	-5.72	2.07	2.07	2.76	0.0058
Bird_species_ <i>E.rubecula</i>	-2.41	1.25	1.25	1.93	0.054
Bird_species_ <i>F.albicollis</i>	1.27	0.88	0.88	1.44	0.15
Bird_species_ <i>P.major</i>	1.12	0.82	0.83	1.35	0.18
Bird_species_ <i>P.trochilus</i>	0.11	1.24	1.24	0.09	0.93
Bird_species_ <i>P.modularis</i>	-0.30	1.27	1.27	0.24	0.81
Bird_species_ <i>T.iliacus</i>	0.89	1.05	1.05	0.84	0.40
Bird_species_ <i>T.merula</i>	2.50	0.91	0.91	2.76	0.006
Bird_species_ <i>T.philomelos</i>	1.11	0.95	0.95	1.17	0.24
Bird_species_ <i>T.pilaris</i>	4.29	1.51	1.51	2.84	0.0046
Tick_species_ <i>I.frontalis</i>	0.47	1.03	1.03	0.45	0.65
Tick_species_ <i>I.ricinus</i>	1.18	0.70	0.71	1.66	0.096
Tick_species_ <i>I.ventalloi</i>	0.96	1.22	1.22	0.78	0.44
Latitude	0.079	0.03	0.03	2.28	0.022
Latitude*Longitude	0.0008	0.0005	0.0005	1.56	0.12

Table 2. Nucleotide diversity (π) and Tajima's D averaged over the eight MLST genes (*clpA*, *clpX*, *pepX*, *pyrG*, *nifS*, *recG*, *rplB*, *uvrA*) of *B. garinii* strains (strain IDs included in this analysis are available in Sup. Mat. 3).

Population	N strains	Mean π	Mean Tajima's D
Continent			
Europe	227	0.008	-0.222
Asia	85	0.009	-1.184
North America	21	0.008	0.617
Country			
Canada	21	0.008	0.617
China	8	0.009	0.022
Finland	12	0.007	0.157
France	18	0.007	-0.090
Germany	55	0.007	-0.326
Japan	21	0.010	-0.865
Latvia	30	0.010	-0.192
Norway	16	0.005	1.085
Russia	54	0.008	-1.026
Sweden	6	0.005	-0.205
UK	70	0.009	0.219

Figure captions

Fig. 1. Map of countries and sampling locations where birds were screened for infesting ticks. Sampling locations closer than 40 km apart are represented under the same pin and are numbered according to study site listed in Sup. Mat. 1. Light grey pins – sites where birds were screened for ticks but no ticks were found; Dark grey pins – sites where ticks were collected feeding on birds. Details on sampling locations, range of bird capturing dates, bird species, number of bird individuals infested and number of *Ixodes* spp. ticks collected at each location are detailed in Sup. Mat 1. Adapted from the Cartographic Research Lab of the University of Alabama.

Fig. 2. *Borrelia burgdorferi* s.l. prevalence (%) and genospecies in *Ixodes* spp. ticks collected from different bird species. Bird species from which less than 10 ticks were tested were included in the category “other bird spp.”. Numbers at the top of the bars represent the number of ticks tested. L – larva; N + A – nymph and adult.

Fig. 3. *Borrelia burgdorferi* s.l. prevalence (%) and genospecies in *Ixodes* spp. ticks collected feeding on birds per country of collection. Numbers at the top of the bars represent the number of ticks tested. L – larva; N + A – nymph and adult.

Fig. 4. *Borrelia garinii* STs distribution between countries in Europe (blue/green), Asia (red) and North America (purple). GoeBURST analysis included 137 STs, using TLV as parameter. Sixteen major clonal complexes and 4 minor clonal complexes (consisting of only two associated STs) were found. Thirty-three isolates formed singletons. A clonal founder was identified in five out of the 16 major clonal complexes (ST86, ST88, ST184, ST244 and ST251), in red. N refers to the number of *B. garinii* isolates used in the analyses.

Fig. 5. Ancestry Recombination Graph of 85 European Sequence Types reconstructed with BEAST2 v. 2.5 and package bacter v. 2.2. Labels are coloured by geographic origin: green - Northern Europe (Estonia, Finland, Latvia, Norway and Sweden), orange - Eastern Europe (Czech Republic, Hungary, Serbia, Slovenia and Yugoslavia), red - Central Europe (France, Germany, Italy, Netherlands) and blue - British islands (United Kingdom). Branches leading to taxa found in one geographic region only show the corresponding colour. Black labels refer to STs present in several geographic regions indicated in coloured rectangles and corresponding in coloration to the regions defined (North, East, Central Europe and British islands). STs isolated from birds (this study) are marked with a bird next to the label name. Dashed lines show recombination events present in at least 50% of all posterior graphs and stars mark high confidence nodes (present in at least 80% of all posterior graphs).

Sup. Mat. 1 Details on sampling locations, range of bird capturing dates, bird species, number of bird individuals infested and number of *Ixodes* spp. ticks collected at each location.

Sup. Mat. 2 Best model selection using Akaike Information Criterion (AIC) to explain (a) *B. burgdorferi* s.l. and (b) *B. garinii* prevalence in ticks collected from birds. Models were fitted using logit function for binomial error distributions. For each model we present: Maximum Likelihood, AICc = Akaike information criterion corrected for sample size, Δ AIC = difference in AIC to the best ranked model and df = degrees of freedom.

Sup. Mat. 3. List of *B. garinii* isolates retrieved from MLST database and used (a) for goeBURST (n = 172), (b) for phylogenetic analysis (n = 110), and (c) to calculate nucleotide diversity and Tajima's D (n = 304), including ST information and MLST database (<https://pubmlst.org/borrelia/>) ID numbers.

Sup. Mat. 4 *Borrelia burgdorferi* s.l. (Bb) prevalence in different tick species and development stages collected feeding on bird species in which > 10 individuals were infested.

Sup. Mat. 5 Data on specimen detailed source of the *B. burgdorferi* s.l. genospecies detected in our study.

Sup. Mat. 6. Figure caption - *Borrelia garinii* STs distribution in European countries. STs detected in ticks feeding on birds are symbolised with a bird icon and the bird species is indicated by the icon colour. GoeBURST analysis included 85 STs, using TLV as parameter. Thirteen major clonal complexes were found and 5 isolates formed singletons. A founder was identified in five out of the 13 major clonal complexes (ST86, ST88, ST184, ST244 and ST251), these are indicated in red numbers. N refers to the number of *B. garinii* isolates used in the analyses. * cluster of STs from Norway.

Sup. Mat. 7. Figure caption - *Borrelia garinii* STs distribution by isolation source: questing ticks/ human isolates are indicated in rose, while ticks collected from birds are colour coded according to migratory status of birds (resident: dark grey, mixed populations - resident and short distance migrants: light grey, or migrant: blue). GoeBURST analysis included 137 STs, using TLV as parameter. Sixteen major clonal complexes and 4 minor clonal complexes (consisting of only two associated STs) were found. Thirty-three isolates formed singletons. A clonal founder was identified in five out of the 16 major clonal complexes (ST86, ST88, ST184, ST244 and ST251), these are indicated in red numbers. N refers to the number of *B. garinii* isolates used in the analyses.



Fig. 1. Map of countries and sampling locations where birds were screened for infesting ticks. Sampling locations closer than 40 km apart are represented under the same pin and are numbered according to study site listed in Sup. Mat. 1. Light grey pins – sites where birds were screened for ticks but no ticks were found; Dark grey pins – sites where ticks were collected feeding on birds. Details on sampling locations, range of bird capturing dates, bird species, number of bird individuals infested and number of *Ixodes* spp. ticks collected at each location are detailed in Sup. Mat 1. Adapted from the Cartographic Research Lab of the University of Alabama

184x148mm (300 x 300 DPI)

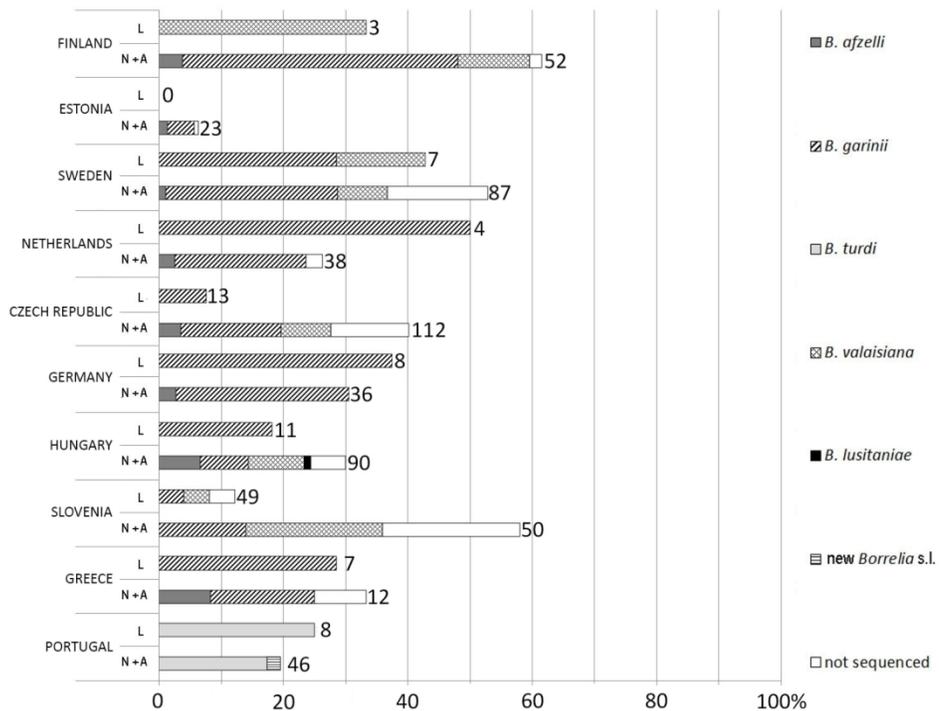


Fig. 2. *Borrelia burgdorferi* s.l. prevalence (%) and genospecies in *Ixodes* spp. ticks collected from different bird species. Bird species from which less than 10 ticks were tested were included in the category "other bird spp.". Numbers at the top of the bars represent the number of ticks tested. L – larva; N + A – nymph and adult.

138x98mm (300 x 300 DPI)

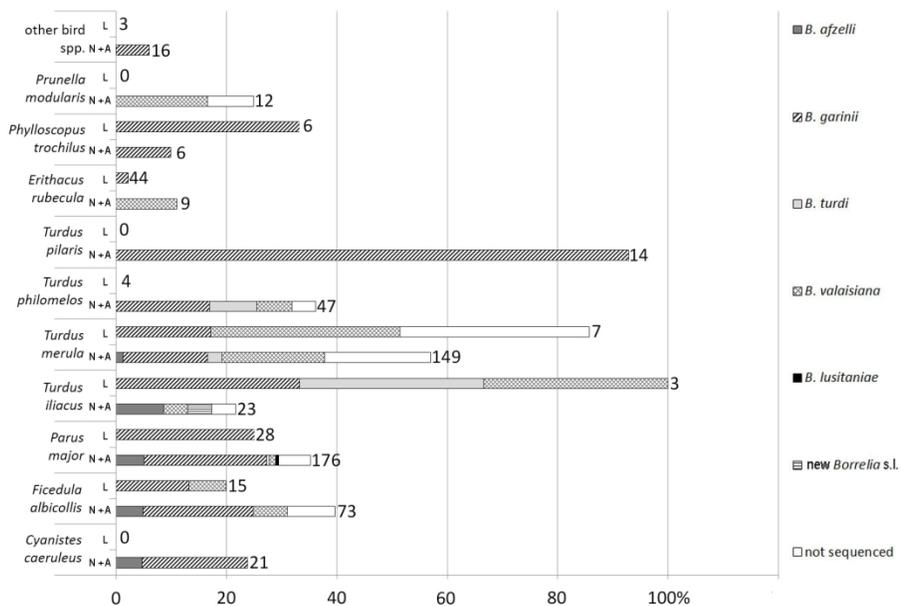


Fig. 3. *Borrelia burgdorferi* s.l. prevalence (%) and genospecies in *Ixodes* spp. ticks collected feeding on birds per country of collection. Numbers at the top of the bars represent the number of ticks tested. L – larva; N + A – nymph and adult.

164x123mm (300 x 300 DPI)

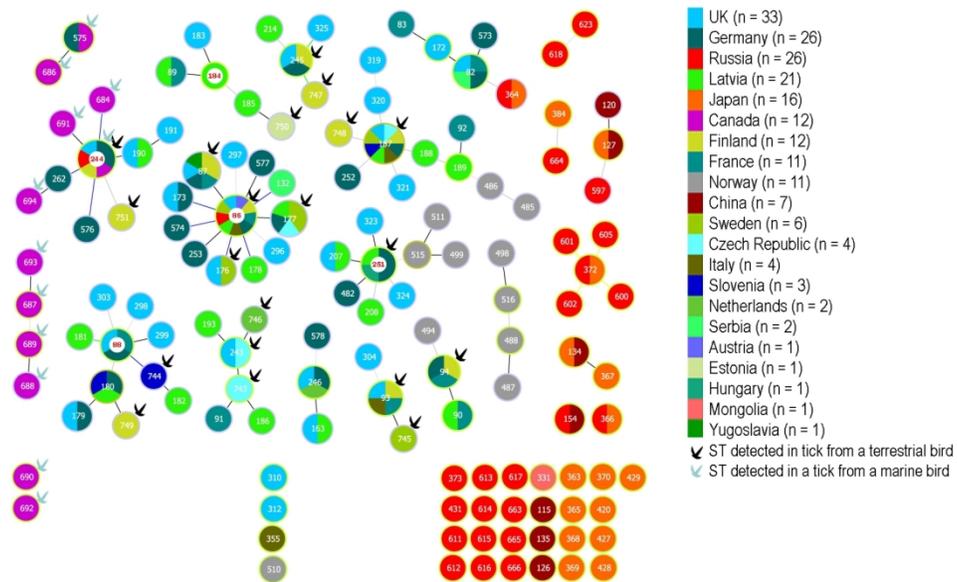


Fig. 4. *Borrelia garinii* STs distribution between countries in Europe (blue/green), Asia (red) and North America (purple). GoeBURST analysis included 137 STs, using TLV as parameter. Sixteen major clonal complexes and 4 minor clonal complexes (consisting of only two associated STs) were found. Thirty-three isolates formed singletons. A clonal founder was identified in five out of the 16 major clonal complexes (ST86, ST88, ST184, ST244 and ST251), in red. N refers to the number of *B. garinii* isolates used in the analyses.

154x93mm (300 x 300 DPI)

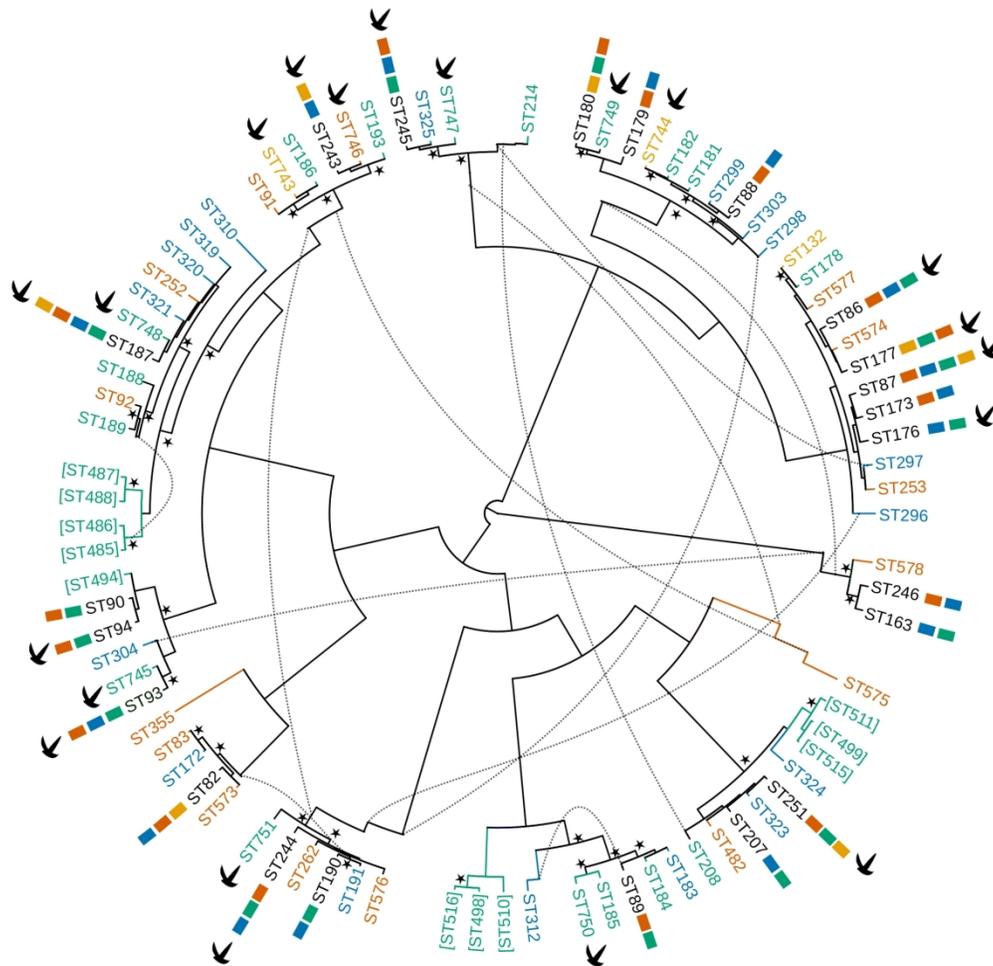


Fig. 5. Ancestry Recombination Graph of 85 European Sequence Types reconstructed with BEAST2 v. 2.5 and package bacter v. 2.2. Labels are coloured by geographic origin: green - Northern Europe (Estonia, Finland, Latvia, Norway and Sweden), orange - Eastern Europe (Czech Republic, Hungary, Serbia, Slovenia and Yugoslavia), red - Central Europe (France, Germany, Italy, Netherlands) and blue - British islands (United Kingdom). Branches leading to taxa found in one geographic region only show the corresponding colour. Black labels refer to STs present in several geographic regions indicated in coloured rectangles and corresponding in coloration to the regions defined (North, East, Central Europe and British islands). STs isolated from birds (this study) are marked with a bird next to the label name. Dashed lines show recombination events present in at least 50% of all posterior graphs and stars mark high confidence nodes (present in at least 80% of all posterior graphs).

189x184mm (300 x 300 DPI)